

## **BLOOD GROUPING REAGENT**

### **DG GEL 8 AB (X4)**

**REF 210386**

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#### Instructions for Use

#### **INTENDED USE**

The DG Gel 8 AB (x4) card is for the determination of human A and B antigens on the surface of red blood cells of four separate blood samples.

For use with the DG Gel System.

For *in vitro* diagnostic use.

#### **SUMMARY AND EXPLANATION**

The ABO system was the first human blood group system discovered by Landsteiner in 1900<sup>1</sup> and is still the most important in transfusion practice. The ABO system is defined by the presence or absence of the A and/or B antigens on human red blood cells and by the presence of antibodies in the plasma or serum corresponding to the antigen or antigens missing in the red blood cells. The anti-A and anti-B reagents contained in the DG Gel 8 AB (x4) card are used to perform the ABO blood group typing in four separate samples.

#### **PRINCIPLE OF THE TEST**

The principle of the test is based on the gel technique described by Yves Lapierre<sup>2</sup> in 1985 for detecting red blood cell agglutination reactions. The DG Gel 8 cards are composed of eight microtubes. Each microtube is made of a chamber, also known as incubation chamber, at the top of a long and narrow microtube, referred to as the column. Buffered gel solution containing specific antibody (anti-A or anti-B) has been prefilled into the microtube of the plastic card. The agglutination occurs when the red blood cell antigens react with the corresponding antibodies, present in the gel solution or in the serum or plasma sample (in the case of reverse grouping test). The gel column acts as a filter that traps agglutinated red blood cells as they pass through the gel column during the centrifugation of the card. The gel column separates agglutinated red blood cells from non-agglutinated red blood cells based on size. Any agglutinated red blood cells are captured at the top of or along the gel column, and non-agglutinated red blood cells reach the bottom of the microtube forming a pellet.

#### **REAGENTS**

##### **Observable indications**

Note: Inspect the condition of the cards before use (see Warnings and Precautions).

Cards with an alteration or change in color, trapped bubbles in the gel, cracked gel or gel with fissures, drying gel, presence of other artifacts, and opened or damaged seals may indicate an alteration of the product.

##### **Material provided**

Note: All microtubes contain sodium azide (NaN<sub>3</sub>) as a preservative at a final concentration of 0.09%.

Each microtube of the DG Gel 8 AB (x4) card contains different reagents (monoclonal antibodies) mixed with a gel in buffered medium with preservative. The different microtubes are identified on the front label of the card.

Microtubes **A**: monoclonal antibody anti-A. Mixture of IgM and IgG antibodies of murine origin, clones 16243G2 and 16247E6.

Microtubes **B**: monoclonal antibody anti-B. IgM antibody of murine origin, clone 9621A8. This clone does not react with acquired B cells.

Clones 16243G2, 16247E6 and 9621A8 is are produced using intermediate products produced for Diagnostic Grifols S.A. in a shared manufacturing agreement with DIAGAST, Parc Eurasante, 251 av. Eugene Avinee-BP9, 59374 Loos Cedex France; US License Number 1744.

### **Reagent preparation**

DG Gel 8 AB (x4) cards are supplied as ready to use. The gel cards and samples to be tested should be brought to room temperature (18 - 25 °C) before testing.

### **Material required but not provided**

#### For Manual Method

- Automatic pipettes of 10 µL, 50 µL and 1 mL.
- Disposable pipette tips.
- Glass or plastic test tubes.
- DG Gel 8 Neutral (if a control microtube is needed).
- Grifols Diluent.
- DG SPIN centrifuge.
- DG Reader Net or DG Reader (optional).

#### For Fully Automated Methods

- DG Gel 8 Neutral (if a control microtube is needed).
- Grifols Diluent.
- Grifols Wash Solution A and Grifols Wash Solution B.
- Erytra Eflexis, Erytra or WADiana Compact.

### **STORAGE AND STABILITY**

- Do not use beyond the expiration date.
- Store upright (as indicated by the two arrows on the outer packaging) with seal intact at 2 – 25 °C.
- Do not freeze.
- Do not expose cards to excessive heat.

### **WARNINGS AND PRECAUTIONS**

- The results by themselves alone are not a clinical diagnosis. Evaluate the results together with the patient's clinical information and other data.
- If you observe microbiological contamination, alterations or changes in color, or other artifacts do not use the card.

- If you observe trapped bubbles in the gel, cracked gel or gel with fissures, drying gel, or gel without a visible fine line of supernatant do not use the card.
- Do not use the card if opened or if the aluminum film seal is damaged.
- If you identify incorrect temperature conditions during storage or shipment, do not use the cards.
- If you identify improper storage or shipping conditions that results in dispersed drops observed, the card should be centrifuged with the DG SPIN before use. If after one centrifugation with the DG SPIN the drops do not descend, do not use the card.
- The product should only be used by qualified personnel.
- The use of volumes and/or red blood cell suspensions in concentrations other than those indicated in the method, may modify the reaction and lead to incorrect test results, i.e., false positive or false negative results.
- The use of diluents other than Grifols Diluent for the red blood cell suspension may modify the reaction and lead to incorrect test results.
- Do not use a centrifuge other than the DG SPIN centrifuge.
- Samples collected in sodium citrate or heparin should be tested by manual method.
- The reagents of the DG Gel 8 AB (x4) card of human monoclonal origin are manufactured using materials that have been tested and found non-reactive for the HBs antigen, and for anti-HIV and anti-HCV antibodies. However, there is no known procedure to ensure that products of human origin will not transmit infectious agents. Human blood products and samples should be handled as if they were potentially capable of transmitting infectious diseases.
- All products with animal derived material, and human blood products and samples should be handled as if they were potentially capable of transmitting infectious diseases.
- Once used, dispose of the product in containers for biological waste, according to local, state and national regulations.
- If you have any questions or need further information on the use of this product, please contact your local Grifols service representative.

#### **SPECIMEN COLLECTION AND PREPARATION**

Blood samples collected in EDTA, sodium citrate or heparin should be used. The collection, separation and handling of the blood should be performed by qualified technical personnel according to current standards<sup>3-4</sup>, and following the instructions of the manufacturer of the materials used for collecting the sample.

Do not use grossly hemolyzed, cloudy or contaminated samples.

Use the red blood cells collected for the determination of the antigens of the ABO system. Samples should be tested as soon as possible. If necessary, samples collected in EDTA and stored at 2 - 8 °C can be used up to 7 days after collection. Samples collected in sodium citrate or heparin and stored at 2 - 8 °C can be used up to 3 days after collection.

Red blood cells from bags collected in ACD, CPD, CPDA-1, CP2D AS-1 (Adsol) or AS-3 can also be used up to 7 days after the expiration date indicated on the label of the bag. If red blood cells from the bag segment are used, it is suggested that these be washed with physiological saline solution before preparing the suspension. Do not use if clots or hemolysis are observed.

## PROCEDURE

1. Allow DG Gel 8 AB (x4) cards, additional reagents and the samples to reach room temperature (18 - 25 °C).  
**Note:** For fully automated instruments, skip the next steps and refer to the Instructions for Use of the related instruments.
2. Identify the cards to be used and the samples to be tested.
3. Prepare a 5% red blood cell suspension in Grifols Diluent (50 µL of packed red blood cells in 1 mL of Grifols Diluent).
4. Remove the foil seal from the complete DG Gel 8 card or from the individual microtubes to be used for testing.
5. Carefully peel off the aluminum film to prevent cross-contamination of the microtube contents among them.
6. Ensure the re-suspension of the red blood cells before use.
7. Add 10 µL of the 5% red blood cell suspension into each of the A/B microtubes.  
**Note:** Carefully dispense the red blood cell suspension avoiding contact of the pipette tip with the wall or the contents of the microtubes to prevent carryover.
8. Centrifuge the gel card in the DG SPIN centrifuge.
9. After centrifugation, remove the gel card from the centrifuge and read the results. Alternatively, use the DG Reader to read and to interpret the results.  
**Note:** If a control microtube (a gel in a buffered medium without antibodies) has not been processed in another DG Gel 8 card for the sample tested, a microtube of the DG Gel 8 Neutral card can be used for this purpose. To process a control microtube, add 10 µL of the same 5% red blood cell suspension sample into one microtube of the DG Gel 8 Neutral card, centrifuge the card in the DG SPIN, and read the results.

## RESULTS

Report results as an agglutination grade, absence of agglutination or hemolysis.

**Negative results:** no agglutination and no hemolysis of red blood cells is visible in the microtube. In a negative result the red blood cells are located in the bottom of the gel column.

**Positive results:** agglutination and/or hemolysis of the red blood cells is visible in the microtube.

In a positive result the agglutinated red blood cells may remain throughout the gel column showing different reaction grades (see Reaction Grades and Figure 1 for a picture of example of reaction grades). Some positive reactions may also form a pellet in the bottom of the microtube. Samples with normal expression of ABO antigens provide strong positive reaction grades. Weaker reactions may indicate a weak or partial expression of ABO antigens. Subgroup A<sub>2</sub> of the ABO system may also present a weak expression.

### Notes:

1. Some fibrin, particulates or other artifacts may trap red blood cells at the top of the gel columns erroneously leading to an abnormal result (see limitation number 7).
2. Occasionally red blood cell retention in the incubation chamber may occur with positive 4+ samples, without interfering in the result interpretation

**Reaction Grades**

<b>Negative:</b>	0	Well-defined pellet of non-agglutinated red blood cells at the bottom of the gel column and no visible agglutinated cells in the rest of the gel column.
<b>Positive:</b>	W+	Barely visible small-sized clumps of agglutinated cells in the lower part of the gel column and a pellet of unagglutinated cells at the bottom.
	1+	Some small-sized clumps of agglutinated cells most frequently in the lower half of the gel column. A small pellet may also be observed at the bottom of the gel column.
	2+	Small or medium-sized clumps of agglutinated cells throughout the gel column. A few unagglutinated cells may be visible at the bottom of the gel column.
	3+	Medium-sized clumps of agglutinated cells in the upper half of the gel column.
	4+	A well-defined band of agglutinated red blood cells in the top part gel column. A few agglutinated cells may be visible below the band.
<b>mf</b>		Mixed-field. A band of red blood cells at the top part of the gel or dispersed throughout the gel column, and a pellet in the bottom as a negative result.
<b>H</b>		Hemolysis in the microtube with very few or no red blood cells in the gel column. Report if hemolysis is present in the microtube but not in the sample.

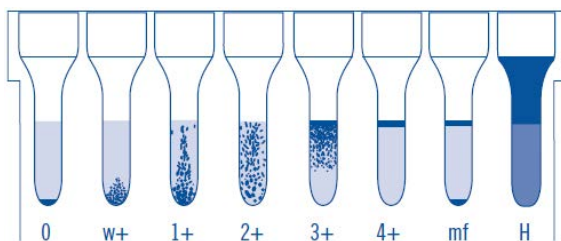


Figure 1. Picture of an example of reaction grades.

**Stability of the results**

After centrifuging the cards it is recommended that the results be read immediately. Do not leave processed cards in a horizontal position. If necessary, a delayed reading can be performed up to 24 hours after processing the cards if they are kept in an upright position, refrigerated (2 - 8 °C) and sealed with a laboratory covering film to avoid evaporation of the supernatant.

**QUALITY CONTROL**

Include positive and negative controls with testing on each day of use. If an unexpected control result is obtained, a complete assessment of the instrument, reagents and material used should be made.

### Interpretation of the results

**ABO system.** The expected reaction with microtubes A and B, and its interpretation are shown in the following table (+ = positive, and 0 = negative)

**Table “Interpretation ABO Forward Group”**

Microtube A	Microtube B	Interpretation
0	0	0
+	0	A
0	+	B
+	+	AB

### Notes:

1. In the control microtube (buffered gel solution without antibodies) the result should be negative. If it is positive, due to the formation of rouleaux, to strong cold autoagglutinins or other causes, invalidate the test. Repeat the determination after washing the red blood cells with physiological saline solution and preparing a new suspension of the washed red blood cells. If the control microtube of the repeat test is negative, the results of the test can be interpreted; if it is positive, invalidate the test.
2. Precaution should be taken in the interpretation of mixed-field events. Not all mixed-cell situations are detected. Additional information on patient history and additional testing will be necessary for resolution. Transfused patients or those subjected to bone marrow transplant may present images of mixed-field<sup>5</sup>. Mixed-field is also observed in some ABO subgroups (A<sub>3</sub>), Tn cryptantigens, in blood group chimerism in fraternal twins, and in the very rare case of mosaicism arising from dispermy<sup>5-6</sup>.
3. The ABO group obtained should be verified comparing the result obtained with a previously typed sample. If a previously typed sample is not available, it is recommended that the reverse group be determined. Forward (cell grouping) and reverse (serum grouping) discrepancies should be investigated before releasing the result.
4. The observation of complete or partial hemolysis (pinkish supernatant and/or gel column) in microtubes should be interpreted as a positive result, after verifying that it is not due to a problem of collection and/or handling of the sample.

### LIMITATIONS OF THE PROCEDURE

1. Grossly hemolyzed, cloudy or contaminated samples or samples with presence of a clot, may cause false positive or false negative results.
2. Aged or hemolyzed specimens may cause weaker reactions compared to those obtained with a fresh sample.
3. Red blood cells from individuals with A or B variants may present a weak expression of the antigens, and may not be detected. These weak antigens may be better detected using reagent anti-AB included in DG Gel 8 ABO/Rh + Kell card.
4. Antigen expression may be weakened in the red blood cells of persons with leukemia or other malignant diseases<sup>5</sup>.

5. Abnormal concentrations of serum proteins, the presence of infused macromolecular solutions in the serum or plasma or the presence of Wharton’s jelly in cord blood samples may cause non-specific agglutination of the red blood cells. It is suggested that red blood cells be washed before performing the test<sup>5</sup>.
6. Samples with high-potency antibodies may coat the red blood cells completely, causing spontaneous agglutination<sup>5</sup>.
7. If poorly anti-coagulated plasma or incompletely clotted serum is used, fibrin residues may trap non-agglutinated red blood cells at the top of the gel, appearing as a pinkish or reddish layer. Although the results could be correctly interpreted, in a negative reaction the false appearance of a mixed field could lead to a misinterpretation. In case of incompletely clotted serum samples, it is recommended to re-clot the serum and repeat the test<sup>5</sup>.
8. The anti-A reagent contained in this card could react with Tn cryptantigens.
9. On occasions, unagglutinated red blood cells may be retained somewhere in the gel column with the appearance of very minute red dot or fleck. However, this nonspecific retention should not interfere with the interpretation of the result.

**SPECIFIC PERFORMANCE CHARACTERISTICS**

- Grifols DG Gel 8 Blood Grouping Reagents Anti-A, Anti-B meet FDA potency requirements for Blood Grouping Reagents.
- Every lot has been tested against a panel of positive and negative samples for the relevant antigens to assure reactivity and specificity in accordance with FDA requirements. Details of specificity test results submitted to the FDA for release of product will be furnished upon request.
- For the manual method, the performance of the reagents was confirmed against FDA-licensed reagents in a comparison study where reagents were tested in parallel at different clinical sites. The estimated percent agreements and their lower limits of 95% one-side confidence interval for all sites combined are indicated on the table below.

**Table “Overall Statistical Analysis Results of the comparison study”**

	Nº of samples	Negative Percent Agreement (Lower 95% CI)	Nº of samples	Positive Percent Agreement (Lower 95% CI)
Anti-A	1863	100.00% (99.84%)	1233	99.84% (99.49%)
Anti-B	2566	100.00% (99.88%)	540	100.00% (99.45%)

- Percent of Agreement only indicates agreement between the Diagnostic Grifols reagents and the FDA-licensed reagents and does not indicate which reagent gave the correct result(s).
- For further information about the performance data for manual method using DG Reader or DG Reader Net and for automated method, please refer to the Instructions for Use of the related instrument.

#### **BIBLIOGRAPHY**

1. Klein HG and Anstee DJ. Mollison's Blood Transfusion in Clinical Medicine, 11th edition, Blackwell Scientific, Publications, Oxford, 2005.
2. Lapiere Y, et al. The gel test: a new way to detect red cells antigen-antibody reactions. Transfusion, 30: 109-113, 1990.
3. CLSI H3-A6: Procedures for the collection of diagnostic blood specimens by venipuncture; Approved Standard, 6th edition, 2007.
4. CLSI H18-A4: Procedures for the handling and processing of blood specimens; Approved Guideline, 4rd edition, 2010
5. Technical Manual, 19th edition, American Association of Blood Banks, Bethesda, Maryland, 2017.
6. Judd WJ, Johnson ST, Storry JR. Judd's Methods in immunohematology, 3rd edition, AABB Press Bethesda, 516, 2008.

#### **PRESENTATION**

210386 DG Gel 8 AB (x4) 50 Cards

#### **Manufactured by:**

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### SYMBOLS KEY

One or more of these symbols may have been used in the labeling/packaging of this product

	<i>In vitro</i> diagnostic medical device
	Batch code
	Use by YYYY-MM-DD or YYYY-MM
	Temperature limitation
	Consult instructions for use
	Catalog number
	This way up
	Fragile, handle with care
	Keep dry