Evaluation of matrix influence in extractables and leachables analysis of medical device extracts Samanthi Wickramasekara¹, Keaton Nahan¹, Berk Oktem¹ and Eric M. Sussman¹

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Plain Language Summary

Accuracy of quantification of extractables and leachables can be reduced by the presence of matrix in the sample. This study is designed to evaluate the extent of the matrix effect in quantifying common polymer additives and explore the feasibility of preparing a suitable matrix matched reference material for the analysis.

Introduction

Background

Extractables and leachables (E&L) are the chemical species that can be released from the device materials to the medium under laboratory and clinical use conditions, respectively. To provide exposure estimate for biocompatibility evaluation, accurate detection and quantification of these E&L are necessary. Liquid chromatography mass spectrometry (LC/MS) is a commonly used analytical technique to identify and semi quantify the nonvolatile extractables in the medium. Matrix effect (ME) occurs when compounds co elute and compete with the ionization of the analyte which can create under/over estimation of the analyte concentration. Occurrence of ME can alter the accuracy, precision and sensitivity of the LC/MS analysis. **Purpose:**

This study aims to evaluate the matrix effect in LC/MS analysis of device extracts using different polymer materials and extraction conditions. Additionally, the extract stability will be tested to determine the feasibility of developing a matrix matched reference standard.

Experimental Workflow



 ESI(+): 0.1% formic acid in water (A) and 0.1% formic acid in acetonitril ESI(-): 10 mM ammonium acetate in water (A) and acetonitrile (B)
40%B from 0.0-2.0 min, 100%B at 10 min and hold until 18.3 min (at 0.55 r
100-1700
200-600 nm (280 nm for qualitative analysis)
 DSSTox: Extracted from EPA DSSTox (>100,000 compounds consisting of Agilent E&L PCDL:~1000 compounds polymer additive relevant compou Contaminant List:Literature based list of common contaminants (<1000

n • • • • • • • • •	Spiked standards (2ppm) Bisphenol A (BPA) bis(2-ethylhexyl) phthalate (DEHP) diethyl phthalate (DEP) diphenyl phthalate (DPhP) Irganox 1010 Irganox 1330 Stearic acid (SA) 2,5-Bis(5-tert-butyl-benzoxazol-2- yl)thiophene (BBOT) 2,6-Di-tert-butyl-4-methylphenol (BHT) 4,4'-Thiobis(2-tert-butyl-5- methylphenol) (STX) Deuterated-DEHP (D4) Deuterated-BPA (D16)
particles	
le (B)	
mL/min)	
^F organic ac unds compoun	cids, carbamic acids, and carbamates)



Figure 1. Base peak chromatograms (BPC's) for spiked device extracts (left) ethanol water (EW) extracts and IPA extracts (right) in ESI(+).

Table 1: Identification/quantification of top 5 extractables in the extract mixture (all material extracts mixed at 1:1 ratio) [EW: 1:1 Ethanol:water; IPA: Isopropyl alcohol]

Name

Oleamide Palmitamide 20-amino-3,6,9,12,15,18-hexaoxaicosanyl)carbamate

Benzyl[...]carbamate* Triton X-100 Reduced N5

Palmitamide

Stearamide

Triton X-100 Reduced N5

Irgafos 168 Phosphate Didodecyl 3,3'-thiodipropionate 2-Hydroxyethane-1-sulfonic acid Carbamic acid, [(trimethoxysilyl)methyl] methyl ester

(Acetyloxy)-carbamic acid tert-butyl este

Carbamic acid, octyl ester Carbamic acid, dipropylthio-, O-ethyl

Octadecyl-4-hydroxyphenylpropionate 2-methylidenepentadecanoic acid Dodecyl 2-methylacrylate Cyanox 1790

Carbamic acid, dipropylthio-, O-ethyl est Benzyl [(3S,5S,6S,8S)-8-[(3-amino-2,2-dimethyl-3-oxopropyl)carbamoyl]-6-hydroxy-3-{[4-methoxy-3-(3-methoxypropoxy)phenyl]methyl}-2,9-dimethyldecan-5-yl]carbamate

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Results

	RT (min)	Extract Mixture	Ionization	Average Concentration (mg/kg)	%RSD
	9.0	EW	(+)	6	3
	10.0		(+)	22	2
-1-	9.9		(+)	101	1
	14.2		(+)	103	1
	8.0		(+)	187	1
	10.0	IPA	(+)	92	3
	11.3		(+)	108	2
	8.0		(+)	257	1
	14.2		(+)	656	1
	14.3		(+)	730	6
	0.5		(-)	222	2
]-,	0.6		(-)	218	21
4.0.0	0.0		(-)	263	$ \begin{array}{c} 3\\ 2\\ 1\\ 1\\ 1\\ 3\\ 2\\ 1\\ 1\\ 6\\ 2\\ 21\\ 1\\ 1\\ 23\\ 7\\ 17\\ 17\\ 1\\ 2\\ 6\\ 1\\ 2\\ 6\\ 1 \end{array} $
ter	0.8	EW	(-)	352	23
[0.6 11.8		(-)	742	7
•	11.0		()	243	17
	0 7			2310	1 /
	<i>9.2</i> <u>8.0</u>	IPA		1856	1 2
	10.7			1560	2
ter	0.5		(-)	1439	6
~~1				1107	



A noluto	Retention Time	Signal IPA			EW		
Anaryte	(Min)		Recovery (%)	%RSD	Recovery (%)	%RSD	
PA	1.77	UV (280 nm)	C.E.	C.E.	C.E.	C.E.	
EP	2.89	UV (280 nm)	114	2	111	<1	
'hP	7.29	UV (280 nm)	111	2	110	1	
BP	7.86	UV (280 nm)	115	1	88	2	
TX	8.56	UV (280 nm)	105	<1	121	1	
IT	8.90	UV (280 nm)	106	1	112	4	
A	12.20	UV (280 nm)	N.D.	N.D.	N.D.	N.D.	
HP	11.57	UV (280 nm)	C.E.	C.E.	C.E.	C.E.	
ΟΤ	11.85	UV (280 nm)	103	<1	105	1	
x 1010	13.34	UV (280 nm)	101	5	97	1	
x 1330	13.60	UV (280 nm)	114	8	109	1	

Discussion and Conclusion

Device materials produced different extractables profiles, which provided a complex matrix when combined

Multiple databases were used to identify these compounds including DSS-TOX LC/UV spike and recovery (accuracy and precision) was improved in the absence of the internal standards. Internal standards did not improve compound recovery in the presence of matrix from the device materials.

LC/UV was less susceptible to matrix effects compared to LC/MS.

Depending on the ionization mode and adducts, concentrations of hydrophobic extractables were underestimated by 20 percent or more in matrix.

Although IPA and EW are sometimes claimed to be equivalent semi-polar

solvents, they produced non-equivalent recoveries for the spiked samples.