

Improving food & health

Chr. Hansen, Inc.

MilwDivision of Biotechnology and GRAS Notice ReviewU.S./Center for Food Safety & Applied Nutrition (HFS-255)PhonU.S. Food & Drug AdministrationFax

Reference: Pichia kluyveri DSM 33235

9015 West Maple Street Milwaukee, WI 53214 4298 ECEIVED Phone : 414 - 607 - 5700 MAY 0 1 2020 : 414 - 607 - 5959 Fax OFFICE OF FOOD ADDITIVE SAFETY April 29, 2020

Dear Sir or Madam,

In accordance with the Federal Register [81 Fed. Reg. 159 (17 August 2016)] issuance on Generally Recognized as Safe (GRAS) notifications (21 CFR Part 170), Chr. Hansen is pleased to submit a notice that we have concluded, through scientific procedures that *Pichia kluyveri* (*P. kluyveri*) DSM 33235 is generally recognized as safe and is not subject to the pre-market approval requirements used to enhance flavor in fermentation of beverages including but not limited to brewing of alcohol free beer and low alcohol beer. The recommendation is to inoculate the pure culture at a level of 0.1 g/L in fermentation. Prior to bottling, alcohol free and low alcohol beers undergo pasteurization or filtration prior to bottling, which removes most if not all *P. kluyveri*.. Though *P. kluyveri* is safe to consume, *P. kluyveri* would only be present in negligible levels, if at all, in the finished food product due to pasteurization or filtration.

If there are any questions or concerns, please contact us.

Yours sincerely,

Arie Carpenter

Senior Regulatory Affairs Specialist usarbr@chr-hansen.com 414-777-7526 CHR. HANSEN, INC.



# Contents

	List of	Tables a	and Figures	3
	List of	Append	lices	3
	Abbrev	viations		3
1	Sign 1.1		ments and certification ent of intent	
	1.2	Name a	and address of notifier	4
	1.3	Commo	on or usual name	4
	1.4	Conditi	ons of use	4
	1.5	Basis fo	or GRAS determination	4
	1.6	Premar	ket approval status	. 5
	1.7	Availab	ility of information	. 5
	1.8	Freedo	m of Information Act	. 5
	1.9	Certific	ation	. 5
	1.10	Signatu	ıre	. 5
2	Iden 2.1		thod of manufacture, specifications, and physical or technical effect of the GRAS organism	
	2.1.2 2.1.2 2.1.2 2.1.4 2.1.4 2.1.5 2.2	2 GN 3 Sp 4 Ge 5 Ar	burce and description of the GRAS organism M status becies identification enome sequencing and functional gene annotation halysis of the <i>P. kluyveri</i> DSM 33235 genome hypic characteristics	7 7 7 7
	2.2.2 2.2.2 2.3	2 Bio	ntifungal susceptibility ogenic amine production d of manufacture	10
	2.3.2 2.4		w materials and processing aids	
	2.4.2 2.5		enetic stability ed technical effect & amount required	
3 4 5 6	Self- Expe	limiting erience k ative Pichia k	bsure levels of use based on common use in food kluyveri is endogenous in fermented plants and as a yeast isolated from food	14 14 14 14
	6.2	History	of safe use of <i>Pichia</i> genus	15
	6.2.2 6.2.2		story of safe use in food of <i>Pichia kluyveri</i> story of safe use of <i>Pichia kluyveri</i> in fermented beverages	



W		ed	
7	List of s	supporting data and information	
6.	4 Cc	onclusion of GRAS status	
	6.3.4	Literature search on adverse effects of P. kluyveri	
	6.3.3	P. kluyveri DSM 33235 does not produce biogenic amines	
	6.3.2	<i>P. kluyveri</i> DSM 33235 is susceptible to most antifungal agents tested	
	6.3.1	P. kluyveri DSM 33235 does not contain any genes of concern	
6	3 <i>P</i> .	kluyveri DSM 33235 is a non-pathogenic safe strain	

## List of Tables and Figures

Figure 2-1: Fingerprints Profiles of *P. kluyveri* DSM 33235 Reference Stock and Inoculation Materials...12

Table 2-1: MIC values for Pichia kluyveri DSM 33235.	10
Table 2-2: Quality control testing Schedule for P. kluyveri DSM 33235.	12
Table 6-1: Literature review about Pichia kluyveri in the wine microbial ecosystem	17

#### List of Appendices

APPENDIX 1: PRODUCT INFORMATION SHEET APPENDIX 2: PRODUCT SPECIFICATION SHEET APPENDIX 3: HACCP EVALUATION OF APPLICATION: LOW ALCOHOL BEER APPENDIX 4: GMO STATEMENT APPENDIX 5: ALLERGENS IN PLANT APPENDIX 6: SAFETY DATA SHEET APPENDIX 7: FSSC 22000 CERTIFICATE: POHLHEIM APPENDIX 8: ISO 22000 CERTIFICATE: POHLHEIM

#### **Abbreviations**

DSM: Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH EFSA: European Food Safety Authority FDA: Food and Drug Administration **CFR: Code of Federal Regulations** NCBI: National Center for Bioinformatics **GRAS:** Generally Recognized As Safe ISO: International Standardization Organization LAB: Lactic Acid Bacteria P. kluyveri: Pichia kluyveri MIC: Minimum Inhibitory Concentration **BA: Biogenic Amine** AA: amino acid Spp: species GMP: Good manufacturing practice PRP: Prerequisite program **OPRP: Operational Prerequisite Program** 



CCP: Critical control point FALCPA: US Food Allergen Labeling and Consumer Protection Act of 2004 NHANES: National Health and Nutrition Examination Survey OIV: International Organization of Vine and Wine VFDB: Virulence Factor Database EUCAST: European Committee on Antimicrobial Susceptibility Testing ECOFF: Epidemiological Cutoff CLSI: Clinical and Laboratory Standards Institute FALCPA: Food Allergen Labeling And Consumer Protection Act CFU: Colony Forming Units

# 1 Signed statements and certification

# 1.1 Statement of intent

In accordance with the 21 CFR 170 Subpart E, regulations for Generally Recognized as Safe (GRAS) notifications, Chr. Hansen, Inc. is pleased to submit a notice that we have concluded, through scientific procedures, that *Pichia kluyveri* DSM 33235 is GRAS and is not subject to the premarket approval requirements for use in fermentation of beverages including but not limited to brewing of alcohol free beer and low alcohol beer.

## 1.2 Name and address of notifier

Chr. Hansen, Inc. 9015 W Maple St. Milwaukee, WI 53214 Tel: (414) 607-5700 Fax: (414) 607-5959

## 1.3 Common or usual name

Yeast / Fermentation Yeast / Pichia kluyveri / P. kluyveri / DSM 33235 / Food culture

## 1.4 Conditions of use

*Pichia kluyveri* DSM 33235 is intended to be used at an inoculation level of 0.1 g/L (1kg for 10,000 L) as a pure starter culture in fermentation of alcohol-free beer and low alcohol beer characterized by a similar flavor profile of regular beers.

Due to selective fermentation of glucose, low alcohol production and high volatile esters productivity, *P. kluyveri* DSM 33235 exhibits optimal characteristics for the purpose of providing superior flavor in these applications. At the end of beer fermentation, this yeast is removed by filtration or centrifugation and killed by pasteurization, therefore it is not present in viable form in the finished product.

## 1.5 Basis for GRAS determination

Pursuant to the GRAS rule [81 Fed. Reg. 159 (17 August 2016)], Chr. Hansen has concluded that *P. kluyveri* DSM 33235 is GRAS through scientific procedures, in accordance with 21 CFR 170.30 (a) and (b).



## 1.6 Premarket approval status

It is the opinion of Chr. Hansen that *P. kluyveri* DSM 33235 pure culture for brewing is not subject to premarket approval requirements of the Federal Food, Drug and Cosmetics Act, based on our conclusion that the notified substance is GRAS under the intended use conditions.

# 1.7 Availability of information

The data and information that is the basis for Chr. Hansen's conclusion that *P. kluyveri* DSM 33235 is GRAS are available for review and copying by FDA during customary business hours, at the location below, or will be sent to FDA upon request, made to:

Chr. Hansen, Inc. c/o Arie Carpenter Senior Regulatory Affairs Specialist 9015 W Maple St., Milwaukee, WI 53214 usarbr@chr-hansen.com

## 1.8 Freedom of Information Act

It is our opinion that the information contained in this notification is not exempt from disclosure under the Freedom of Information Act.

#### 1.9 Certification

To the best of our knowledge, this GRAS notification is a complete, representative, and balanced submission that includes unfavorable information, as well as favorable information, known to us and pertinent to the evaluation of the safety and GRAS status of the use of *Pichia kluyveri* DSM 33235.

## 1.10 Signature

– Arie	Čarpenter, Šenior Re	- egulatory Affai	rs Specialist	
- 1				

Katharine Urbain, Head of Regulatory Affairs – North America <u>April 29, 2020</u> Date

April 29, 2020

Date

# 2 Identity, method of manufacture, specifications, and physical or technical effect

# 2.1 Name of the GRAS organism

The subject of this GRAS determination is the yeast, Pichia kluyveri, designated as DSM 33235.

# 2.1.1 Source and description of the GRAS organism

*Pichia kluyveri* DSM 33235 was isolated from spontaneous ferment of mature Chardonnay grapes and published by (Goddard, 2008). The strain has been deposited into the German Collection of Microorganisms and Cell Cultures (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH) under the accession number DSM 33235. *P. kluyveri* is placed in the *Pichia* genus (Kurtzman, Robnett, & Basehoar-Powers, 2008), which belongs to the *Pichiaceae* family and the order of *Saccharomycetales* along with other yeast species relevant for food and biotechnology such as *Saccharomyces cerevisiae*, *Debaryomyces hansenii, Yarrowia lipolytica* and *Komagataella pastoris* (reclassified from *Pichia pastoris*).

Yeasts of the *Pichia* genus can be found in diverse natural habitats ranging from soil and water to a variety of plants and mature or rotten fruits. *Pichia* spp. are frequently isolated from food (Villa-Carvajal, Querol, & Belloch, 2006) (Loureiro & Malfeito-Ferreira, 2003). In the microbiome of some agronomically important fruits, such as grape, *Pichia spp*. are even considered predominant (Sabate, Cano, Esteve-Zarzoso, & Guilllamon, 2002).

As a species, *P. kluyveri* is a wild non-pathogenic yeast. True to the genus, its natural habitat is diverse and varies from the fleshy part of plants, fruit insects such as *Drosophila* sp., fruits and vegetables. Literature has shown a diversity of fruits and vegetables in which *Pichia kluyveri* can be found. *Pichia kluyveri* naturally occurs in produce such as grapes, persimmon, apple, cocoa beans, coffee beans, soy, olives, tomato and cactus necrotic tissues (Vaudano, et al., 2019) (Bozoudi & Tsaltas, 2016) (Kim, Lee, Jeon, & Park, 2019) (Wei, Zhang, Yuan, Dai, & Yue, 2019) (Holt, Mukherjee, Lievens, Verstrepen, & Thevelein, 2018) (Gross, Kunz, Muller, Santos Kron, & Freimoser, 2018) (Broissin-Vargas, Snell-Castro, Godon, Gonzalez-Rios, & Suarez-Quiroz, 2018) (Vadkertiova, Molnarova, Vranova, & Slavikova, 2012) (Hamby, Hernandez, Boundy-Mills, & Zalom, 2012)

*Pichia* is characterized by multilateral budding on a narrow base, presence or absence of true hyphae but pseudohyphae may occur, ascospores may be hat-shaped, hemispheroidal, or spherical with or without a ledge, sugars may be fermented and nitrate is not utilized as a source of nitrogen (Kurtzman C. P., 2011) (Kurtzman, Fell, & Boekhout, The Yeasts : a taxininuc study, 2011) (Villa-Carvajal, Querol, & Belloch, 2006). *Pichia kluyveri* Bedford ex Kudryavtsev (1960) – having the synonyms *Pichia kluyveri* Bedford (1942), *Hansenula kluyveri* Bedford ex Kudryavtsev (1960), *Pichia belgica* Dekker (1941) and *Zygosaccharomyces bisporus* Anderson (1917) - is an aerobic teleomorph yeast producing two to four hat-shaped spores in each ascus, able to ferment only glucose and produce pectin degrading enzymes (Kurtzman, Fell, & Boekhout, The Yeasts : a taxininuc study, 2011) (Pichia kluyveri, 2019).



Classification of the organism Pichia kluyveri is as follows:

- Kingdom: Fungi
- Phylum: Ascomycota
- Subphylum: Saccharomycotina
- Class: Saccharomycetes
- Order: Saccharomycetales
- Family: Pichiaceae
- Genus: Pichia
- Species: kluyveri

#### 2.1.2 GM status

*Pichia kluyveri* DSM 33235 is not genetically modified by use of recombinant DNA techniques (APPENDIX 1: PRODUCT INFORMATION SHEET 715390).

#### 2.1.3 Species identification

*Pichia kluyveri* DSM 33235 was species identified at an external laboratory according to the current standard for yeast identification using ITS5/4 and D1/D2 primers as described by Vu *et al.* (2016). The species *Pichia kluyveri* was first described by Bedford (1942) but it has been modified since, thus the correct reference for *Pichia kluyveri* is Kudryavtsev (1960). The *P. kluyveri* type strain is the CBS 188 strain.

#### 2.1.4 Genome sequencing and functional gene annotation

To obtain a high-quality genome sequence of *P. kluyveri* DSM 33235, the strain was genome sequenced using the Illumine MiSeq Technology and Oxford Nanopore Technology. Output from the MiSeq sequencing (4,909,396 raw reads and coverage 78x) was assessed and trimmed as previously described Agersø *et al.* (2018) and combined with the ONT reads (26,619 raw reads and coverage 37x) to obtain high quality hybrid assembly. The hybrid assembly from both sequencing technologies resulted in 92 contigs with a total assembly size of 13.6 Mbp with a GC content of 28.4%.

The genome sequence of the *P. kluyveri* DSM 33235 was subjected to gene finding and functional annotation using the state-of-the-art program GeneMark-ES in 'fungi'-mode and predicted proteins were annotated the using a Swiss-Prot database. GeneMark-ES predicted 6,474 proteins of which 93% were functional annotated.

The genome size, GC content and number of proteins in the *P. kluyveri* DSM 33235 were comparable to the genome sequence of *Pichia kluyveri* CBA6002 (QEFR01.1) published by Kim *et al.* (2019) as well as the genome sequences of other *Pichia* spp. in the NCBI genome database.

#### 2.1.5 Analysis of the P. kluyveri DSM 33235 genome

Databases of virulence genes and antimicrobial resistance genes does not exist for yeast strains in similar fashion as several curated databases for virulence and antibiotic resistance genes are published for bacterial strains. In addition to this, acquired resistance is not a concern in yeast as it is in bacteria (Arendrup & Patterson, 2017) (EFSA Panel on Biological (BIOHAZ), 2020).



Improving food & health

#### 2.1.5.1 Search of functional gene annotations for words of potential safety concern.

The gene annotations of *P. kluyveri* DSM 33235 were searched to identify terms that could be linked to antifungal resistance. A total of 39 proteins included one or more of these words in its annotation. Many proved to be housekeeping genes or transporters, five genes had functional annotations that could be linked with resistance to fluconazole i.e. 'Negative regulator of PDR1-mediated fluconazole resistance JJJ1', 'Fluconazole resistance protein 1', 'Fluconazole resistance protein 3' or 'Lanosterol 14-alpha demethylase' (CYP51/ERG11). As acquired antifungal resistance is not an issue in yeast strains (Arendrup and Patterson, 2017; EFSA Panel on Biological Harzards (BIOHAZ), 2020) and as homologous proteins were observed in other Pichia spp. and other budding yeast, these genes are conserved and therefore not considered a concern. Several of these genes are assumed to play a role in the intrinsic fluconazole resistance in P. kudriavzevii (anamorph form C. krusei). Both Lamping et al. (2009) and Whaley et al. (2017) stated that the main resistance mechanism is assumed to be efflux pump activity i.e. reduced drug accumulation in combination with reduced azole affinity at the target protein, Erg11p. Whereas Feng et al. (2016) find that it is point mutations and increased expression of Erg11 that lead to azole resistance, however they also state that the mechanism is not fully investigated in *P. kudriavzevii* (Feng. Yang, Wang, Chen, & Xi, 2016). The remaining functional annotations that were linked to 'resistance' were associated with drug, multidrug, oligomycin, vanadate, ethionine, alkylphosphocholines, nitrosoguanidine and homologs were observed in other Pichia spp.. These conserved genes are putatively involved in the intrinsic fluconazole resistance observed in the DSM 33235 strain and they were dismissed as safety concerns.

In a similar fashion the gene annotations were searched for terms that could be linked to virulence. This search identified 19 genes of which all were found to be of no safety concern.

Finally, because biogenic amines are produced by decarboxylation of amino acids, the gene annotations were searched for the word 'decarboxylase'. Only one such gene was found, labeled 'ornithine decarboxylase' putatively involved in decarboxylation of ornithine to putrescine. (Rocha & Wilson, 2019) describes that polyamines, such as putrescine, are essential metabolites found in yeasts and the biosynthesis of putrescine to spermidine to spermine are involved in critical roles in the cell. A homolog of ornithine decarboxylase is observed in *Pichia* spp. strains and many different *Saccharomycetales* strains including *S. cerevisiae* supporting the presence in other yeast strains including strains important for food productions.

#### 2.1.5.2 Search for the Candida albicans toxin candidalysin

Candidalysin is regarded as the first and only 'true' virulence factor identified in *C. albicans*. It is a cytolytic peptide toxin secreted by the invasive form of the number one human pathogenic yeast and is critical for mucosal and systemic infections and triggers protective immune responses (Moyes, et al., 2016) (Naglik, Gaffen, & Hube, 2019). Candidalysin is a 31 amino acid peptide produced by cleavage of the Ece1p protein (Moyes, et al., 2016) (Naglik, Gaffen, & Hube, 2019). Blast search of the 31 aa peptide against *P. kluyveri* DSM 33235 strain did not detect any homologous peptides in the DSM 33235 strain.

#### 2.1.5.3 Genome analysis for mycosins in the DSM 33235 genome sequence

A variety of eukaryotic and prokaryotic microorganisms produce antimicrobial substances such as bacteriocins that enable the ability to dominate in a certain environment by antimicrobial activity to closely related microorganisms (Schaffrath, Meinhardt, & Klassen, 2018). Antimicrobial activity or



inhibitory activity of yeast against other yeast is often described to be caused by secondary metabolites known as 'killer toxins' or 'mycocins' (Hatoum, Labrie, & Fliss, 2012). *Pichia* spp. have great potential for antimicrobial activity and screening studies have identified *P. kluyveri* as the most frequently isolated yeast with killer activity (Starmer, Ganter, & Aberdeen, 1992) (Abranches, Morais, Rosa, Mondonca-Hagler, & Hagler, 1997).

Search for killer toxins in the genome of the *P. kluyveri* DSM 33235 strain showed that the strain encodes a putative killer toxin, which has homology to the protein sequence of the killer toxin KT395 and preproteins of other killer toxins. Furthermore, in the DSM 33235 strain is also identified proteins related to  $\beta$ -1,3-glucanase activity, which in some publication referred to killer activity. It is however debated in the literature if lytic enzymes should be referred to as antagonistic toxins (Pretscher, et al., 2018). Pretscher *et al.* (2018) did detect extracellular amylase, cellulase,  $\beta$ -glucosidase and protease activity in two different *P. kluyveri* strains including the 395 strain and the above finding support that DSM 33235 strain might have similar antimicrobial activity.

There are no indications that 'killer toxins' or antimicrobial substances produced by yeast strains including yeasts important for food industry are real toxins and are therefore not harmful to human beings. Thus, they are therefore not of any safety concerns.

#### 2.1.5.4 Search for secondary metabolite biosynthesis pathways in the DSM 33235 strain

To analyze for biosynthesis genes for secondary metabolites, the genome of DSM 33235 and genomes of yeast species relevant for the food industry were analyzed using published methods. The analysis showed that the DSM 33235 strain does not encode any secondary metabolites. The results of the analysis are in line with the general impression that yeasts do not produce secondary metabolites and are of low biosafety concern (Freimoser, Rueda-Mejia, Tilocca, & Migheli, 2019).

# 2.2 Phenotypic characteristics

## 2.2.1 Antifungal susceptibility

Minimum inhibitory concentrations (MICs) of 8 antifungals were determined for *P. kluyveri* DSM 33235 according to the EUCAST standards (EUCAST definitive document E.Def 7.3.1) (Table 2-1). In line with the recommendation by EUCAST, the MICs were compared with the cut-off values established for the breakpoint values for *P. kudriavzevii* (the teleomorph version of *C. krusei*) by EUCAST and CLSI as no breakpoints are published for *P. kluyveri*. The range of antifungals tested are in line with the recommendation given by EFSA 'Scientific Opinion on the maintenance of the list of QPS microorganisms intentionally added to food or feed (2013 update)' to test clinically relevant antifungals (EFSA, 2020).



Antifungal group	Antifungal	MIC value (mg/l)	EUCAST breakpoint Candida kruseiª	EUCAST ECOFF Candida krusei <sup>b</sup>	CLSI Breakpoint <i>Candida krusei</i> <sup>c</sup>
Azoles	Fluconazole	>64	_ d	128	-
	Itraconazole	0.5	-	1	-
	Posaconazole	0.125	-	ND	-
	Voriconazole	1	-	1	≥2
Echinocandins	Anidulafungin	≤0.008	0.064	0.064	≥1
	Micafungin	≤0.008	-	0.25	≥1
Polyenes	Amphotericin	0.06	1	1	-
Pyrimidine analogue	Flucytosine	2	-	-	-

#### Table 2-1: MIC values for Pichia kluyveri DSM 33235.

a: EUCAST Antifungal Clinical Breakpoint for *Candida krusei (P. kudriavzevii)* in as listed Table v. 9.0 valid from 2018-02-12. B: EUCAST ECOFF for *Candida krusei (P. kudriavzevii)* given at https://mic.eucast.org/Eucast2/ (Reference date: February 11<sup>th</sup>, 2020)

c: CLSI breakpoints for resistant *C. krusei (P. kudriavzevii)* as listed in Table 1 'Minimal Inhibitory Concentrations Breakpoints for *In Vitro* Broth Dilution Susceptibility Testing of *Candida* spp. And Selected Antifungal Agents After 24-Hours Incubation' in the CLSI standard M60 'Performance Standards for Antifungal Susceptibility Testing of Yeasts' (2017).

D: no breakpoint or ECOFF provided in M60 or EUCAST.

*Pichia kluyveri* DSM 33235 is sensitive to most of the antifungal tested with MIC values that are below the CLSI breakpoints and the EUCAST ECOFF values for the *P. kudriavzevii (C. krusei)* (Table 2-1). The strain has low susceptibility to fluconazole and this is considered to be intrinsic resistance as a similar resistance pattern is observed for other *P. kluyveri* strains (Data generated internally at Chr. Hansen and (Xiao, *et al.*, 2018). *Pichia* spp. are reported to be intrinsically resistant to fluconazole in literature as well (Cuenca-Estrella, 2013) (Arendrup & Patterson, 2017).

Regarding resistance in other *Pichia* species, *P. kudriavzevii* (*C. krusei*) is described as either intrinsically resistant or insensible to fluconazole in several reviews (Cuenca-Estrella, 2013) (Arendrup & Patterson, 2017) (Wiederhold, 2017) and similar is shown for small population of clinical isolates of *Pichia norvegensis* (the teleomorph version of *Candida norvegensis*) isolated nearly 60 years apart (Sandven, Nilsen, Digranes, Tjade, & Lassen, 1997). The observed fluconazole intrinsic resistance in *Pichia* spp. is supported by larger studies such as a study of 6,082 blood sepsis isolates (Pfaller, *et al.*, 2004) and the ARTEMIS DISK study of 256,882 isolates (Pfaller M. D., 2010). Accordingly, the ECOFF set by EUCAST and the absence of breakpoint set by CLSI for *P. kudriavzevii* (*C. krusei*) are well supported.

#### 2.2.2 Biogenic amine production

The DSM 33235 strain was tested for the production of histamine, tyramine, cadaverine and putrescine using an in-house procedure based on published methods. The strain did not produce any of the four biogenic amine compounds tested when grown in presence of specific amino acid precursors known to induce production of the biogenic amines.

#### 2.3 Method of manufacture

*Pichia kluyveri* DSM 33235 can be manufactured by Chr. Hansen GmbH Giessener Str. 94, Pohlheim, in accordance with current Good Manufacturing Practices (cGMP) consistent with 21 CFR Parts 110 and 117 and following Chr. Hansen's global protocol for the production of yeasts to be used in fermented



beverages. The production plant complies with a set of basic GMP-rules, also called Pre-Requisite Program (PRP) according to Chr. Hansen's Quality, GMPs and Food Safety Principles, which are available from our website: www.chr-hansen.com. In addition, the production plant has an appointed local OPRP (Operational Pre-Requisite Program) that includes PRP issues and CCPs (Critical Control Points), which are documented and are classified as specifically critical for the safety of food ingredients produced in the plant. The Pohlheim production plant maintains the following certifications: FSSC 22000 and ISO 22000 (APPENDIX 7: FSSC 22000 CERTIFICATE POHLHEIM and APPENDIX 8: ISO 22000 CERTIFICATE POHLHEIM).

Chr. Hansen's *P. kluyveri* DSM 33235 product is sold as a frozen block of liquid, and is produced by inoculating the microorganism into sterilized growth substrate. Aerobic conditions are maintained during the fermentation; pH and temperature are controlled. When the microbiological growth peaks, fermentation is stopped by cooling.

Sterile plastic food grade bags are labeled with product name, item number, batch number, amount, and storage temperature. These labeled bags are filled with liquid containing above 1.0E + 09 CFU/g of *P. kluyveri* and frozen for storage. The process flow including critical control points is shown in APPENDIX 3: HACCP FLOW SHEET GLOBAL.

#### 2.3.1 Raw materials and processing aids

Chr. Hansen's *P. kluyveri* DSM 33235 product is manufactured using standard fermentation techniques. This includes the use of fermentation and standardizing ingredients that are safe and suitable for use in human food. These ingredients have no technical function in the finished food product and are all permitted for this application in addition to meeting the specifications of the Food Chemical Codex.

Chr. Hansen's *P. kluyveri* DSM 33235 product does not contain any allergens in accordance with the list of common allergens in accordance with the US Food Allergen Labeling and Consumer Protection Act of 2004 (FALCPA), and by the EU Regulation 1169/2001/EC Annex II, as shown in APPENDIX 5, ALLERGENS IN PLANT. Though there are allergens present in the production facility, there is no risk that those allergens could contaminate *P. kluyveri* production line (APPENDIX 3: HACCP FLOW SHEET).

## 2.4 Specifications

Purity is controlled as described in Table 2-2 and additionally in APPENDIX 2: PRODUCT SPECIFICATION SHEET 715390. Moreover, absence of chemical contamination is assessed per a global monitoring program along with a vendor management program.

Enumeration of *P. kluyveri* DSM 33235 strain is carried out based on ISO 7954:1987, ISO 6611 | IDF 94, along with OIV-OENO 576B-2017 recommendations. Yeast extract Glucose Chloramphenicol Agar (YGC) plates are inoculated with *P. kluyveri* culture and are incubated for 72 hours at 25°C in aerobic conditions.

The testing for contaminating microorganisms is carried out following standard test methods outlined in OIV monograph of non-Saccharomyces yeasts (Aurand, 2017) and published in the International Oenological Codex. While this international standard is focused on wine, it provides purity standards for non-Saccharomyces yeasts in fermented beverages where such an internationally recognized standard does not exist for beer to date.



Microorganism	Specification	Frequency
Coliforms	<100 cfu/g	Every batch
E. coli	Absent in 1 g	Every batch
Lactic Acid Bacteria	<1000 cfu/g	Every batch
Mould	<1000 cfu/g	Every batch
Staphylococci	Absent in 1 g	Every batch
Salmonella	Absent in 25g	Per monitoring
		program

Table 2-2: Quality control testing Schedule for *P. kluyveri* DSM 33235.

#### 2.4.1 Genetic stability

Genetic stability of *P. kluyveri* DSM 33235 has been demonstrated by DNA fingerprinting comparing the stock culture in the cell bank and various batch of inoculation material produced since 2014 (Figure 2-1).

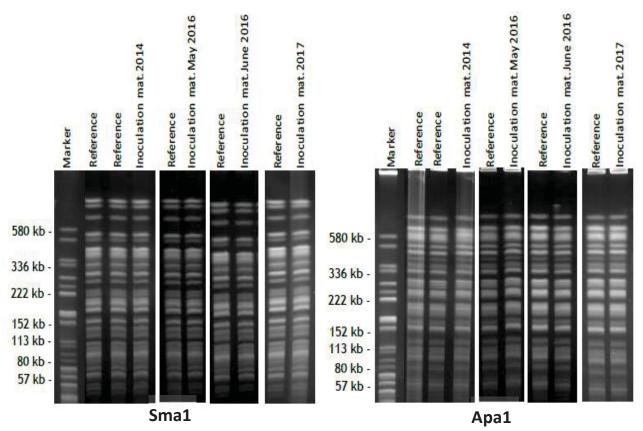


Figure 2-1: Fingerprints Profiles of *P. kluyveri* DSM 33235 Reference Stock and Inoculation Materials.



# 2.5 Intended technical effect & amount required

In nature, *Pichia kluyveri* can be found primarily in fruits, plants, vegetative fermentation and decay. As an example, *P. kluyveri* is one of the organisms that helps to impart organoleptic characteristics such as fruitiness, sweetness, and cocoa aroma in the sensory profile of chocolate (Barista, Ramos, Ribeiro, Pinheiro, & Schwan, 2015) (Crafack, Mikkelsen, Saerens, Knudsen, Blennow, Lowor, Takrama, Swiegers, Petersen, Heimdal, Nielsen, 2013).

Currently, *P. kluyveri* is a commercialized oenological yeast, and it is widely used in winemaking as non-*Saccharomyces* starter culture known as Frootzen<sup>™</sup> (FROOTZEN - first ever Pichia kluyveri yeast, 2019). With this dossier, we would like to demonstrate the safety and versatility of *P. kluyveri* in the production of non-alcoholic and low alcoholic beer applications in order to improve fruity flavors developed during the natural fermentation process.

When used in beer brewing alcohol-free and low alcohol beer, *P. kluyveri* DSM 33235 strain is added at the start of fermentation where it consumes glucose in the wort and produces ester compounds (flavor) together with carbon dioxide. After fermentation the beer is left to mature. Once maturation is complete, alcohol free and low alcohol beer must be filtered or centrifugated prior to pasteurization and bottling. The process of filtration or pasteurization eliminates *P. kluyveri* DSM 33235 culture from the product.

At the time of use, the frozen *P. kluyveri* DSM 33235 culture should be defrosted in lukewarm water (30°C) for 5 – 10 minutes before being directly inoculated into the fermentation tank. The culture containing *P. kluyveri* DSM 33235 has a total cell count above 1.0E + 09 cfu/g and is intended to be inoculated at a concentration of 0.1 g/L (approximately 1.0E + 05 cfu per ml / 1 KG per 10,000L). Frozen *P. kluyveri* DSM 33235 product has a shelf life of 18 months when stored at -50°C.

# 3 Dietary exposure

*P. kluyveri* DSM 33235 is either pasteurized or filtered out of the finished low or non-alcoholic beer. Because of this, only a negligible amount of non-viable microorganisms, if any, would be present in the bottled beverages. The dietary intake of *P. kluyveri* yeast cells, therefore, are not expected to increase due to consumption of the final product.

If by chance *P. kluyveri* DSM 33235 is ingested, literature and the conducted strain specific safety assessment on the DSM 33235 shows that it would be safe to consume (Aponte & *et al.*, 2010) (Banjara, Suhr, & Hallen-Adams, 2015) (DEAK & BEUCHAT, 1993) (Liang, Zhang, Wu, Liu, & Zhang, 2016) (Ogunremi, Sanni, & Agrawal, 2015). Furthermore, it is well established that the microbiome of an adult is very stable and would only shift in microbial composition due to significant dietary changes or extreme weight loss (Faith, et al., 2013). Therefore, if *P. kluyveri* DSM 33235 strain, as viable yeast, would reach the digestive system of consumers, due to sporadic consumption of fermented beverages, it's presence should be considered transient and is not expected to change the gastrointestinal flora.



# 4 Self-limiting levels of use

The proposed use of the *P. kluyveri* DSM 33235 strain is as a food ingredient added for fermentation of low alcohol beer and alcohol-free beer with the intent of enhancing flavor and aromatic profiles. The self-limiting levels of use are:

- Current GMP Following the use level prescribed by Chr. Hansen, the *P. kluyveri* DSM 33235 strain will only be added for fermenting beverages at levels required to achieve the technical effect in the final product. There would be no benefit to the customer to add *P. kluyveri* product at higher levels due to the following:
  - Increased cost to the customer
  - Negative impact on flavor or aromatic profiles

# 5 Experience based on common use in food

The basis for the GRAS conclusion for the *P. kluyveri* DSM 33235 is based on scientific procedures and not common use in food before 1958.

# 6 Narrative

## 6.1 *Pichia kluyveri* is endogenous in fermented plants and as a yeast isolated from food

Yeasts of the *Pichia* genus are widely distributed in diverse natural habitats such as soil, freshwater, tree exudates, plants, mature or rotten fruits, and frequently they are reported as yeasts isolated from food (Villa-Carvajal, Querol, & Belloch, 2006) (Loureiro & Malfeito-Ferreira, 2003). *Pichia* species are considered predominant in the microbiome of some agronomically important fruits such as in grapes (Sabate, Cano, Esteve-Zarzoso, & Guilllamon, 2002). Carbohydrate utilization is species specific and while the genus is heterogeneous, each species within the genus generally assimilates only a few sugars and other carbon compounds (Kurtzman, Fell, & Boekhout, 2011).

*Pichia kluyveri* is a yeast that finds its natural habitat in the fleshy part of plants, fruit insects (e.g. *Drosophila* sp.), fruits and vegetables such as grapes, persimmon, apple, cocoa beans, coffee beans, soy, olives, tomato and cactus necrotic tissues (Vaudano *et al.*, 2010) (Vaudano, *et al.*, 2019) (Bozoudi & Tsaltas, 2016) (Kurtzman, Fell, & Boekhout, The Yeasts : a taxininuc study, 2011) (Kim, Lee, Jeon, & Park, 2019) (Wei, Zhang, Yuan, Dai, & Yue, 2019) (Holt, Mukherjee, Lievens, Verstrepen, & Thevelein, 2018) (Gross, Kunz, Muller, Santos Kron, & Freimoser, 2018) (Broissin-Vargas, Snell-Castro, Godon, Gonzalez-Rios, & Suarez-Quiroz, 2018) (Vadkertiova, Molnarova, Vranova, & Slavikova, 2012) (Hamby, Hernandez, Boundy-Mills, & Zalom, 2012) (Gross, Kunz, Muller, Santos Kron, & Freimoser, 2018) (Kurtzman and Fell, 1998) (Kurtzman & Fell, 1998) (Vadkertiova, Molnarova, 2012) (Hamby, Hernandez, Boundy-Mills, & Zalom, 2012).

Based on the wide array of food products that *P. kluyveri* are associated with, it can be concluded that *P. kluyveri* has a long history of safe human consumption.



# 6.2 History of safe use of *Pichia* genus

Yeast strains of the *Pichia* genus can be found in a variety of foods that we eat as well as beverages that are consumed regularly. Several *Pichia* spp. occur naturally in the spontaneous microbiome of foods and beverages. Some selected species have also been intentionally added as starter cultures for imparting characteristic flavors and aromatic profiles.

## 6.2.1 History of safe use in food of *Pichia kluyveri*

*Pichia* spp. have been isolated, in viable form, in bakery sourdoughs, partially processed vegetables, cheeses, beers, must and wines among others (Banjara, Suhr, & Hallen-Adams, 2015) (Marquina, et al., 1992) (Nuobariene, Arneborg, & Hansen, 2014) (Hammes, et al., 2005) (KING, Magnuson, Torok, & Goodman, 1991) (Pereira-Dias, Potes, Marinho, Malfeito-Ferreira, & Loureiro, 2000) (Jolly, Augustyn, & Pretorius, 2006) (Drumonde-Neves, Franco-Duarte, Lima, Schuller, & Pais, 2017).

*Pichia kluyveri* has a well-established and important role in both coffee fermentation and chocolate making. *P. kluyveri* is one of the microorganisms responsible for natural fermentations in coffee beans, essential for pulp removal and sensory quality development (Broissin-Vargas, Snell-Castro, Godon, Gonzalez-Rios, & Suarez-Quiroz, 2018). When inoculated in cocoa as the starting culture in combination with other yeasts, *P. kluyveri* was found to become predominant in the microbiota during cocoa bean fermentation, influencing the chocolate flavor profile with more intense fruitiness, sweetness and cocoa aroma in sensory evaluation (Barista, Ramos, Ribeiro, Pinheiro, & Schwan, 2015) (Crafack *et al.*, 2013).

In regard to spontaneously fermented vegetables, *P. kluyveri* has been traditionally reported in the representative yeast flora of olive brines. *P. kluyveri* has also been found in homemade *paocai*, a traditional Chinese preparation containing various fermented ingredients such as cabbage, radish, celery, lettuce, pepper and garlic (Aponte & *et al.*, 2010) (Villa-Carvajal, Querol, & Belloch, 2006) (Hurtado, Reguant, Esteve-Zarzoso, Bordons, & Rozes, 2008) (Nisiotou, Chorianopoulos, Nychas, & Panagou, 2010)( (Liang, Zhang, Wu, Liu, & Zhang, 2016) (Chang & *et al.*, 2008). *P. kluyveri* has also been found to naturally occur in the microbiome of kimchi, contributing to the spontaneous fermentation of this traditional Korean food. Documented cases of *P. kluyveri* overgrowth in kimchi were associated with the presence of aesthetically undesirable white colonies, however no health risk has been associated with the overgrowth of this yeast (Kim J. Y., *et al.*, 2019).

#### 6.2.2 History of safe use of *Pichia kluyveri* in fermented beverages

Due to its high environmental stress tolerance (e.g. towards glucose osmolarity, pH variation and ethanol concentration) and superior sensorial contributions (e.g. significant biosynthesis of ester compounds), *P. kluyveri* is appreciated as alternative to *Saccharomyces* yeasts, or in co-fermentation with them, for improving the flavor bouquets of alcoholic beverages (Kim J. Y., 2019) (Contreras, *et al.*, 2015) (Jolly, Augustyn, & Pretorius, 2006) (Padilla, Gil, & Manzanares, 2016) (Kim, Hong, & Park, 2008) (Wei, Zhang, Yuan, Dai, & Yue, 2019). In this respect, *P. kluyveri* has a long and documented history of both intentional and unintentional safe use in traditional production of grape wine, coyol palm (*Acrocomia aculeata*) "taberna" wine, mezcal from agave (*Agave salmiana*), new wine "federweisser", as well as in brewing of spontaneously fermented beers, likely since ancient times. No safety risks have



been ever reported or associated with fermentations involving *P. kluyveri* in beverages (Santiago-Urbina, Arias-Garcia, & Ruiz-Teran, 2015) (Diaz, Molina, Nahring, & Fischer, 2013).

#### 6.2.2.1 Pichia kluyveri in wine

Vinification or wine making was traditionally based on spontaneous fermentations due to the action of various microorganisms either present on the grape surface or in the winery ecosystem. Alcoholic fermentation is the most important biochemical process that is responsible for converting grape must into wine. Yeasts are the driving force in transforming musts into wine, especially yeasts belonging to the *S. cerevisiae* species.

In traditional winemaking, several non-*Saccharomyces* yeast species intervene in the early phases of the fermentation contributing unique properties to the final flavor. One of the most abundant non-*Saccharomyces* oenological yeasts is *P. kluyveri*, as demonstrated by its ubiquitous and consistent detections in wild fermentations of traditional wines (Pardo, Garcia, Zuniga, & Uruburu, 1989) (Lopandic & *et al.*, 2008) (Diaz, Molina, Nahring, & Fischer, 2013). A comprehensive literature review documenting *P. kluyveri* as an indigenous yeast of the natural winery biota is shown in Table 6-1.

In modern winemaking, must is inoculated with selected commercial *Saccharomyces* yeasts to control the microbiological process and prevent undesired spoilage. Recently, however, there has been an increasing interest in the industrial application of non-*Saccharomyces* species that are normally isolated from the wine ecosystem, such as *P. kluyveri*.

In support of this interest, scientific studies investigated the influences on sensorial and organoleptic properties due to the activities of these alternative yeasts isolated from the wine microbiota, and noteworthy positive results were reported (Fleet G., 2003) (Beckner Whitener *et al.*, 2016). When intentionally inoculated, *P. kluyveri* was found to improve the aroma of Sauvignon Blanc and Riesling, by increasing the volatiles thiols content (e.g. 3-mercaptohexyl acetate) and the peach/apricot character, in co-fermentation with *S. cerevisiae* (Anfang, Brajkovich, & Goddard, 2008) (Benito *et al.*, 2015).

*P. kluyveri* nowadays represents a safe and well-established wine microorganism, especially recommended for enhancing tropical fruit aroma in Riesling, Sauvignon Blanc and Chardonnay wines. Additionally, *P. kluyveri* was developed as commercial non-*Saccharomyces* starter culture (Frootzen<sup>™</sup>), and it can be used as starter pure inoculum, in co-fermentation or in sequential inoculation for vinification (FROOTZEN - first ever Pichia kluyveri yeast, 2019) (Benito *et al.*, 2015) (Jolly, Augustyn, & Pretorius, 2006) (Beckner Whitener *et al.*, 2016) (New Zealand government-owned Crown Research Institute, 2014).



Sample of origin	Wine cultivar	Country	Publication
Air vineyard, air cellar, tank surface, grape skin, fermenting must	Pinot Noir, Cornalin, Chardonnay, Gutedel, Resi Ermitage	Switzerland	(Diaz, Molina, Nahring, & Fischer, 2013)
Fermenting must	Zweigelt, Grüner Veltliner	Austria	(Lopandic <i>et al.,</i> 2008)
Grape skin	Žametovka, Modra frankinja	Slovenia	(Raspor, Milek, Polanc, Smole Mozina, & Cadez, 2006)
Fermenting must	Refošk	Slovenia	(Zagorc <i>et al.,</i> 2001)
Fermenting must	Red and white wines from not specified grapes among the ones in this list: Lista'n Noir, Negramoll, Tintilla, Vijariego, Lista'n Blanc, Malvası'a, Gual, Verdello, Marmajuelo, Moscatel	Spain	(Gonzalez, Barrio, & Querol, 2006)
Fermenting must	Carinyena, Garnacha	Spain	(Sabate, Cano, Esteve-Zarzoso, & Guilllamon, 2002)
Fermenting must	Tempranillo, Macabeo, Bobal	Spain	(Pardo, Garcia, Zuniga, & Uruburu, 1989)
Fermenting must	Tempranillo	Spain	(Ocon <i>et al.,</i> 2010)
Fermenting must	Merlot	France	(Fleet, Lafon-Lafourcade, & Ribereau-Gayon, Evolution of yeasts and lactic acid bacteria during fermentation and storage of bordeaux wines, 1984)
Grape skin	Riesling	Canada	(Chamberlain, Husnik, & Subden, 1997)
Grape skins, fermenting must	Grignolino	Italy	(Vaudano, <i>et al.</i> , 2019)
Grape skins, fermenting must	Pinot blanc Riesling	Italy	(Guzzon <i>et al.,</i> 2016)
Fermenting must	Catalanesca	Italy	(Di Maro, Ercolini, & Coppola, 2007)
Grape skin	Trincadeira Preta	Portugal	(Barata, Gonzailez, Malfeito- Gerreira, Querol, & Loureiro, 2008)
Fermenting must	Castelão	Portugal	(Baleiras Couto, Reizinho, & Duarte, 2005)

# **Table 6-1**: Literature review about *Pichia kluyveri* in the wine microbial ecosystem



Grape skin,	Blue Frankish,	Slovakia	(Nemcova, Breierova,
fermenting must	Green Veltliner,		Vadkertiova, & Molnarova,
	Sauvignon blanc		2015)
Fermenting must	Isabel,	Brazil	(Bezerra-Bussoli, Baffi, Gomes,
	Bordeaux,		& Da-Silva, 2013)
	Cabernet Sauvignon		
Fermenting must	Cabernet Sauvignon	China	(Li, Liu, Xue, & Liu, 2011)
Grape skin,	Malbec,	Argentina	(Lopes, Rodriguez, Sangorrin,
fermenting must	Merlot		Querol, & Caballero, 2007)
Grape skin	Cabernet Sauvignon	Israel	(Zahavi, Droby, Cohen, Weiss, &
			Ben-Arie, 2002)
Clarified grape juice	Unknown cultivar from the Stellenbosch	South	(Strauss, Jolly, Lambrechts, &
	region	Africa	van Rensburg, 2001)
Fermenting must	Veltlin green	Czech	(Suranska, Vranova, Omelkova,
		Republic	& Vadkertiova, 2012)
Grape skin	Unknown from Tokaji region	Hungary	(Sipiczki, 2016)

## 6.2.2.2 Beer, beer-like and cereal-based beverages

Cereal-based beverages are widespread and include beer, beer-like beverages and several traditional beverages produced in various parts of the world (Chand Bhalla *et al.*, 2009). Yeast is a traditional and essential ingredient in the fermentation of these beverages (Center for Food Science and A Nutrion) (BierStDB - Verordnung zur Durchfuhrung des Vorlaufigen Biergesetzes, n.d.) (T. Alcohol and Tobacco Tax and Trade Bureau, n.d.). Spoilage of cereal-based beverages by yeast is not easily defined since the "spoilage" may be perceived as positively contributing to the product aroma and flavor (Loureiro & Querol, 1999). Moreover, after a thorough literature investigation, no food poisoning cases linked with yeast spoilage of cereal based beverages could be found (RASFF portal | Food Safety, 2019) (Boulton & Quain, 2001).

*Pichia kluyveri* has a documented history of safe use in fermentation of different types of traditional cereal-based beverages and in beer. The presence of this yeast has been documented in certain popular fermented beer-like beverages such as *champús* in Colombia, *chicha* manufactured in Colombia and Ecuador, *kununzaki* and *burukutu* in Nigeria, and *Caxiri* in Brazil (Osorio-Cadavid, Chaves-Lopez, Tofalo, Paparella, & Suzzi, 2008) (Andres Lopez-Arboleda, Ramirez-Castrillon, Adriana Mambuscay-Mena, & Osorio-Cadavid, 2010) (Pilo *et al.*, 2018) (Ogunremi, Sanni, & Agrawal, 2015) (Miguel, Collela, de Almeida, Dias, & Schwan, 2015). Likewise, *P. kluyveri* was reported as being present during the fermentation of *tchapalo*, a traditional sorghum beer from Cote d'Ivoire (N'guessan, Brou, Jacques, Casaregola, & Dje, 2011).

*Pichia* can be found in spontaneously fermented beers that are already present on the United States market. The American coolship ale (ACA) is a type of beer produced by a small number of breweries in the United States and ACA brewing follows practices of the lambic Belgian beer method. In this adaptation of the lambic process by American craft brewers, *Pichia* was found to be part of a yeast community dominated by *Brettanomyces bruxellensis* (Bokulich, Bamforth, & Mills, 2012).

These studies suggest that *P. kluyveri* can be associated with safe fermentations of a varieties of cereals such as corn, sorghum, barley and millet. Moreover, it can be reasonably assumed that *P. kluyveri* can be considered a non-*Saccharomyces* brewing yeast naturally participating in wild cereal-based, beer-like



and beer fermentations that have been safely consumed by humans for a long time in fermented beverages.

# 6.3 *P. kluyveri* DSM 33235 is a non-pathogenic safe strain

The DSM 33235 strain isolated from spontaneous ferment of mature Chardonnay grapes has been unambiguously species identified to be a *Pichia kluyveri* strain. The Pariza decision tree asks questions concerning an organism's ability to produce harmful substances. Section 6.3 addresses the questions posed in the Pariza decision tree that address pathogenicity.

## 6.3.1 P. kluyveri DSM 33235 does not contain any genes of concern

The genome of *P. kluyveri* DSM 33235 was sequenced and analyzed for genes of concern. The genome did not contain any homologous genes to the one known true virulence factor for yeast, candidalysin, and no functional gene annotations that could be linked with potential safety concerns in regard to 'virulence' were detected.

Furthermore, functional gene annotations were used to search for words connected with potential safety concerns such as "antifungal" or "resistance". As discussed in section 2.2.1, the only hit of potential concern identified might play a role in intrinsic resistance to fluconazole observed in the DSM 33235 strain. When "resistance" was examined, it was associated with drug, oligomycin, vandate, ethionine, aklylphosphochlolines and nitrosoguanidine. Homologues proteins were observed in other *Pichia* species and these were found to be of no safety concern in DSM 33235.

Finally, the genome of DSM 33235 strain was searched genes encoding for antimicrobial substances also referred to as killer toxins in yeast. The *P. kluyveri* DSM 33235 was found to encode genes with putative antimicrobial activity. The genome was also analyzed for genes associated with secondary metabolites and there was no concern of secondary metabolites for the intended uses covered in this dossier.

## 6.3.2 P. kluyveri DSM 33235 is susceptible to most antifungal agents tested

As discussed in Section 2.2.1, *P. kluyveri* DSM 33235 was screened for antifungal resistance against eight common antifungal agents and the strain was sensitive to itraconazole, posaconazole, voriconazole, anidulafungin, micafungin, amphotericin and flucytosine. The *P. kluyveri* DSM 33235 strain was found to be intrinsically resistant to fluconazole, which seem to be a general characteristic of the *Pichia* genus.

## 6.3.3 *P. kluyveri* DSM 33235 does not produce biogenic amines

As discussed in Section 2.2.2, biogenic amines are produced by decarboxylation of amino acids. Gene annotations were searched for the word "decarboxylase" and only one such gene was found. The gene was annotated "ornithine decarboxylase." A homologous gene is identifiable in many yeast strains widely used in food production such as *S. cerevisiae*. The gene is subsequently not of concern in *P. kluyveri* for use in similar applications.

## 6.3.4 Literature search on adverse effects of P. kluyveri

For thousands of years, yeast strains have been an important and essential ingredient in production of wine, beer, bread and dairy production and, as part of daily life, consumers ingest large amounts of yeast cultures without adverse effects on the health (Fleet G. H., 2007) (Jacques, 2008). Therefore, there is in general a low concern on the biosafety of yeasts based on the facts that yeasts only rarely cause



human infections (Jacques, 2008) (Anoop, Rotaru, Shwed, Tayabali, & Arvanitakis, 2015) (Perez-Torrado & Querol, 2016) (EFSA, 2007) (EFSA, 2009) and are not known as prolific producers of harmful secondary metabolites (Freimoser, Rueda-Mejia, Tilocca, & Migheli, 2019).

As with consumption of any live microorganism there is no such thing as zero risk of infection (Saarela, Matto, & Mattila-Sandholm, 2002) and infections caused by *P. kluyveri* in immuno-compromised patients have been reported in rare occasions (Xiao *et al.*, 2018) (Aslani *et al.*, 2018). Aslani *et al.* (2018) reported a case of infection with *P. kluyveri* in one patient out of 162 cancer patient included in a study at Mazandaran University Hospital, Sari, Iran, and Xiao *et al.* (2018) one infection caused by a *P. kluyveri* strain among 884 non-candidal infections isolates isolated from invasive fungal infections over a 5-year period from 65 Chinese hospitals. In many other studies or reviews on pathogenic yeasts, *P. kluyveri* strains are not reported to cause human infections or the species is not considered to be pathogenic (Falagas, Roussos, & Vardakas, 2010) (Fernandez-Ruiz & al., 2017) (Guinea, 2014) (Horn & al., 2012) (Seyedmousavi & al., 2018). Limon, Skalski, & Underhill, 2017) (Yapar, 2014) (Eggimann, Garbino, & Pittet, 2003) (Astvad & al, 2018). The conclusion of the literature study and the performed strain-specific safety assessment of the *P. kluyveri* DSM 33235 strain is that the strain is to be considered safe.

## 6.4 Conclusion of GRAS status

In summary, the data presented in this document fully supports the conclusion that *Pichia kluyveri* DSM 33235 is GRAS for the production of non-alcoholic beer and low alcoholic beer to add desirable flavors to these products. The basis for this conclusion follows the Pariza *et al.* decision tree.

*Pichia kluyveri* DSM 33235 has been unambiguously assigned a genus and species, and the genome has been sequenced. The sequenced genome has been analyzed for encoding virulence factors and toxins associated with pathogenicity and was found to be free of such elements. The genome has also been analyzed for genes involved in antimicrobial resistance and found to have conserved genes putatively involved the intrinsic fluconazole resistance observed in the strain. As acquired antifungal resistance is not an issue in yeast strains (Arendrup and Patterson, 2017; EFSA Panel on Biological Harzards (BIOHAZ), 2020), these conserved genes are not considered a safety concern. *P. kluyveri* DSM 33235 was analyzed for the production of antimicrobial substances and found to produce putative genes having antimicrobial activity in line with other *P. kluyveri* strains and yeasts important for the food industry.

*Pichia kluyveri* DSM 33235 has not been genetically modified and can be found endogenously in many different parts of the plant kingdom. It has a long history of safe use in a variety of foods and has an integral part in the flavor development of those foods and has not been shown to induce any undesirable physiological effects when consumed in food.

*Pichia kluyveri* DSM 33235 is a nonpathogenic and safe yeast that has been subject to an adequate strain-specific safety assessment mitigating the risk of any negative impact on human health and subsequently, Chr. Hansen has concluded that it is GRAS.



# 7 List of supporting data and information

## Works cited

- Abranches, J., Morais, P., Rosa, C., Mondonca-Hagler, L., & Hagler, A. (1997). The incidence of killer activity and extracellular proteases in tropical yeast communities. *Can J. Microbiol*, 328-336.
- Agerso, Y., Stuer-Lauridsen, B., Bjerre, K., Jensen, M., Johansen, E., Bennedsen, M., & et al. (2018).
   Antimicrobial susceptibility testing and tentative epidemiological cut-off values of five Bacillus species relevant for use as animal feed additives or for plant protection. *Appl Environ Microbiol*, 84: AEM.01108-18.
- Andres Lopez-Arboleda, W., Ramirez-Castrillon, M., Adriana Mambuscay-Mena, L., & Osorio-Cadavid, E. (2010). Diversidad de levaduras asociadas a chichas tradicionales de Colombia yeast diversity associated to Colombian traditional chichas.
- Anfang, N., Brajkovich, M., & Goddard, M. R. (2008). Co-fermentation with Pichia kluyveri increases varietal thiol concentrations in Sauvignon Blanc. *Aust. J. Grape Wine Res.*, 1-8.
- Anoop, V., Rotaru, S., Shwed, P., Tayabali, A., & Arvanitakis, G. (2015). Review of current methods for characterizing virulence and pathogenicity potential of industrial Saccharomyces cereviseae strains towards humans. *FEMS Yeast Res*, 1-12.
- Aponte, M., & et al. (2010). Study of green Sicilian table olive fermentations through microbiological chemical and sensory analyses. *Food Microbiol*, 162-170.
- Arendrup, M., & Patterson, T. (2017). Multidrug-resistant candida: Epidemiology, molecular mechanisms, and treatment. *J Infect Dis*, S445-S451.
- Aslani, N., & al., e. (2018). Identificatgion of uncommon oral yeasts from cancer patients by MALDI-TOF mass spectrometry. *BMC Infect. Dis.*, 1-11.
- Astvad, K., & al, e. (2018). Update from a 12-year nationwide fungemia surveillance: Increasing intrinsic and acquired resistance causes concern. J. Clin. Microbiol.
- Aurand, J.-M. (2017, June 2). Monograph of non-Saccharomyces yeasts; Resolution OIV-OENO 576B-2017. In I. O. Wine, *International Oenological Codex* (pp. 1-7). Sofia: International Organization of Vine and Wine.
- Baleiras Couto, M., Reizinho, R., & Duarte, F. (2005). Partial 26S rDNA restriction analysis as a tool to characterise non-Saccharomyces yeasts present during red wine fermentations. *Int. J. Food Microbiol.*, 49-56.
- Banjara, N., Suhr, M., & Hallen-Adams, H. (2015). Diversity of Yeast and Mold SPecies from a Variety of Cheese Types. *Curr. Microbiol.*, 792-800.
- Barata, A., Gonzailez, S., Malfeito-Gerreira, M., Querol, A., & Loureiro, V. (2008). Sour rot-damaged grapes are sources of wine spoilage yeasts. *FEMS Yeast Res.*, 1008-1017.
- Barista, N., Ramos, C., Ribeiro, D., Pinheiro, A., & Schwan, R. (2015). Dynamic behavior of Saccharomyces cerevisiae, Pichia Kluyveri and Hansenianspora uvarum during spontaneous and inoculated



cocoa fermentations and their effect on sensory characteristics of chocolate. *LWT - Food Sci. Technol*, 221-227.

Beckner Whitener, M., & et al. (2016). Untangling the wine metabolome by combining untargeted SPME-GCXGC-TOF-MS and sensory analysis to profile Sauvignon blanc co-fermented with seven different yeasts. *Metabolomics*, 53.

Bedford, C. (1942). A Taxonomic Study of the Genus Hansenula. *Mycologia*, 628-649.

- Benito, S., & et al. (2015). Effect on quality and composition of Riesling wines fermented by sequential inoculation with non-Saccharomyces and Saccharomyces cerevisiae. *Eur. Food Res. Technol.*, 707-717.
- Bezerra-Bussoli, C., Baffi, M., Gomes, E., & Da-Silva, R. (2013). Yeast Diversity Isolated from Grape Musts During Spontaneous Fermentation from a Brazilian Winery. *Curr. Microbiol.*, 356-361.
- *BierStDB Verordnung zur Durchfuhrung des Vorlaufigen Biergesetzes*. (n.d.). Retrieved Feb 12, 2019, from http://www.gesetze-in-internet.de/bierstbd/BJNR701350931
- Bokulich, N., Bamforth, C., & Mills, D. (2012). Brewhouse-Resident Microbiota Are Responsible for Multi-Stage Fermentation of American Coolship Ale. *PLoS One*, e35507.
- Boulton, C., & Quain, D. (2001). Brewing yeast and fermentation. Blackwell Science.
- Bourdichon, F., & et al. (2012). Food fermentations: Microorganisms with technological beneficial use. International Journal of Food Microbiology, 87-97.
- Bozoudi, D., & Tsaltas, D. (2016). Grape Microbiome: Potential and Opportunities as a Source of Starter Cultures. In A. Morata, & I. Loira, *Grape and Wine Biotechnology*. InTech.
- Broissin-Vargas, L., Snell-Castro, R., Godon, J., Gonzalez-Rios, O., & Suarez-Quiroz, M. (2018). Impact of storage conditions on fungal community composition of green coffee beans Coffea arabica L. stored in jute sacks during 1 year. *Journal of Applied Microbiology*, 547-558.
- Center for Food Science and A Nutrion. (n.d.). *Guidance for Industry: Labeling of Certain Beers Subject to the Labeling Jurisdiction of the Food and Drug Administration.*
- Chamberlain, G., Husnik, J., & Subden, R. (1997). Freeze-desiccation survival in wild yeasts in the bloom of icewine grapes. *Food Res. Int.*, 435-439.
- Chand Bhalla, T., & et al. (2009). Cereal based alcoholic beverages.
- Chang, H.-W., & et al. (2008). Analysis of yeast and archaeal population dynamics in kimchi using denaturing gradient gel electrophoresis.
- Contreras, A., Hidalgo, C., Schmidt, S., Henschke, P., Curtin, C., & Varela, C. (2015). The application of non-Saccharomyces yeast in fermentations with limited aeration as a strategy for the production of wine with reduced alcohol content. *Int. J. Food Microbiol*, 7-15.



- Crafack, Mikkelsen, Saerens, Knudsen, Blennow, Lowor, Takrama, Swiegers, Petersen, Heimdal, Nielsen. (2013). Influencing cocoa flavour using Pichia Kluyveri and Kluyveromyces marxianus in a defined mixed starter culture for cocoa fermentation. *Int. J. Food Microbiol*, 103-116.
- Cuenca-Estrella, M. (2013). Antifungal drug resistance mechanisms in pathogenic fungi : from bench to bedside. *Clin Microbiol Infect*, 54-59.
- DEAK, T., & BEUCHAT, L. (1993). Yeasts Associated with Fruit Juice Concentrates. J. Food Prot., 777-782.
- Di Maro, E., Ercolini, D., & Coppola, S. (2007). Yeast dynamics during spontaneous wine fermentation of the Catalanesca grape. *Int. J. Food Microbiol.*, 201-210.
- Diaz, C., Molina, A., Nahring, J., & Fischer, R. (2013). Characterization and dynamic behavior of wild yeast during spontaneous wine fermentation in steel tanks and amphorae. *Biomed Res Int*, 540465.
- Diversa Corporation. (2006). *GRAS Notices, GRN No. 204 Phospholipase C enzyme preparation from Pichia pastoris expressing a heterologous phospholipase C gene.* San Diego: C. 92121 4995 Directors Place.
- Drumonde-Neves, J., Franco-Duarte, R., Lima, T., Schuller, D., & Pais, C. (2017). Association between Grape Yeast Communities and Vineyard Ecosystems. *PLoS One*, e0169883.
- EFSA . (2009). Scientific Opinion on the maintenance of the list of QPS microorganisms intentionally added to food or feed (2009 update). *EFSA J.*, 1431.
- EFSA (European Food Safety Authority). (2007). Ethyl carbamate and hydrocyanic acid in food and beverages. *EFSA J*, 1-44.
- EFSA. (2007). Introduction of a Qualified Presumption of Safety (QPS) approach for assessment of selected microorganisms referred to EFSA. *EFSA J.*, 1-16.
- EFSA. (2007, December). Introduction of a Qualified Presumption of Safety (QPS) approach for assessment of selected microorganisms referred to EFSA Opinion of the Scientific Committee. *EFSA Journal, 5*(12), 587.
- EFSA Panel on Biological (BIOHAZ). (2020). Scientific opinion on the update of the list of QPSrecommended biological agents intentionally added to food or feed as notified to EFSA. *EFSA J*, 5966.
- Eggimann, P., Garbino, J., & Pittet, D. (2003). Epidemiology of Candida species infections in critically ill non-immunosuppressed patient. *Lancet Infect. Dis.*, 685-702.
- Esteve-Zarzoso, B., Manzanares, P., Ramon, D., & Querol, A. (1998). The role of non-Saccharomyces yeasts in industrial winemaking. *Int. Microbiol.*, 143-8.
- Faith, J., Guruge, J., Charbonneau, M., Subramanian, S., Seedorf, H., & Goodman, A. (2013). The Long-Term Stability of the Human Gut Microbiota. Retrieved from Science: http://science.sciencemag.org/content/341/6141/1237439



- Falagas, M., Roussos, N., & Vardakas, K. (2010). Relative frequency of albicans and the various nonalbicans Candida spp among candidemia isolates from inpatients in various parts of the world: A systematic review. *Int. J. Infect Dis.*, e954-e966.
- Feng, W., Yang, J., Wang, Y., Chen, J., & Xi, Z. (2016). ERG11 mutations and upregulation in clinical itraconazole-resistant isolates of Candida krusei. *Can J Microbiol*, 938-943.
- Fernandez-Ruiz, M., & al., e. (2017). Fungemia due to rare opportunistic yeasts: Data from a populationbased surveillance in Spain. *Med. Mycol.*, 125-136.
- Fleet, G. (2003). Yeast interactions and wine flavour. Int. J. Food Microbiol., 11-22.
- Fleet, G. H. (2007). Yeasts in foods and bevereages: impact on product quality and safety. *Curr. Opin. biotechnol*, 170-175.
- Fleet, G., Lafon-Lafourcade, S., & Ribereau-Gayon, P. (1984). Evolution of yeasts and lactic acid bacteria during fermentation and storage of bordeaux wines. *Appl. Environ. Microbiol.*, 1034-1038.
- Freimoser, F., Rueda-Mejia, M., Tilocca, B., & Migheli, Q. (2019). Biocontrol yeasts: mechanisms and applications. *World J Microbiol Biotechnol*, 1-19.
- FROOTZEN first ever Pichia kluyveri yeast. (2019, Jan 22). Retrieved from http://www.chrhansen.com/en/food-cultures-and-enzymes/fermented-beverages/cards/productcards/frootzen-first-ever-pichia-kluyvery-yeast
- Goddard, M. (2008). Quantifying the complexities of Saccharomyces cerevisiae's ecosystem engineering via fermentation. *Ecology*, 2077-2082.
- Gonzalez, S., Barrio, E., & Querol, A. (2006). Molecular identification and characterization of wine yeasts isolated from Tenerife (Canary Island, Spain). *J. Appl. Microbiol.*
- Gross, S., Kunz, L., Muller, D., Santos Kron, A., & Freimoser, F. (2018). Characterization of antagonistic yeasts for biocontrol applications on apples or in soil by quantitative analyses of synthetic yeast communities. *Yeast*, 559-566.
- Guinea, J. (2014). Global trends in the distribution of Candida species causing candidemia. *Clin. Microbiol. Infect.*, 5-10.
- Guzzon, R., & et al. (2016). Evaluation of the oenological suitability of grapes grown using biodynamic agriculture: the case of a bad vintage. *J. Appl. Microbiol.*, 355-365.
- Hamby, K., Hernandez, A., Boundy-Mills, K., & Zalom, F. (2012). Associations of yeasts with spotted-wing Drosophila (Drosophila suzukii; Diptera:drosophilidae) in cherries and raspberries. *Applied environmental Microbiology*, 4869-4873.
- Hammes, W., Brandt, M., Francis, K., Rosenheim, J., Seitter, M., & Vogelmann, S. (2005). Microbial ecology of cereal fermentations. *Trends Food Sci Technol*, 4-11.
- Hatoum, R., Labrie, S., & Fliss, I. (2012). Antimicrobial and probiotic properties of yeasts: from fundamental to novel applications. *Front Microbiol*, 421.



- Holt, S., Mukherjee, V., Lievens, B., Verstrepen, K., & Thevelein, J. (2018). Bioflavoring by nonconventional yeasts in sequential beer fermentations. *Food Microbiology*, 55-66.
- Horn, F., & al., e. (2012). Systems biology of fungal infection. Front. Microbiol., 1-20.
- Hurtado, A., Reguant, C., Esteve-Zarzoso, B., Bordons, A., & Rozes, N. (2008). Microbial population dynamics during the processing of Arbequina table olives. *Food Res. Int.*, 738-744.
- Impossible Foods Inc. (2018). *GRAS Notice GRN No 737 Soy leghemoglobin preparation from a strain of Pichia pastoris.* Redwood City.
- Jacques, N. &. (2008). Safety assessment of dairy microorganisms: The hemiacomycetous yeast. *Int. J.* Food Microbiol, 321-326.
- Jolly, N., Augustyn, O., & Pretorius, I. (2006). The Role and Use of Non-Saccharomyces Yeasts in Wine Production. *South African J. Enol. Vitic.*, 15-39.
- Jolly, N., Varela, C., & Pretorius, I. (2014). Not your ordinary yeast: non-Saccharomyces yeasts in wine production uncovered. *FEMS Yeast Res.*, 215-237.
- Kim, D., Lee, S., Jeon, J., & Park, H. (2019). Development of air-blast dried non-Saccharomyces yeast starter for improving quality of Korean persimmon wine and apple cider. *International Journal of Food Microbiology*, 193-204.
- Kim, D.-H., Hong, Y.-A., & Park, H.-D. (2008). Co-fermentation of grape must by Issatchenkia orientalis and Saccharomyces cerevisiae reduces the malic acid content in wine. *Biotechnol. Lett.*, 1633-1638.
- Kim, J. Y. (2019). Community structures and genomic features of undesirable white colony-forming yeasts on fermented vegetables. *J. Microbiol.*, 30-37.
- KING, A., Magnuson, J., Torok, T., & Goodman, N. (1991). Microbial Flora and Storage Quality of Partially Proessed Lettuce. *J. Food Sci*, 459-461.
- Kudryavtsev. (1960). Bot. Mater. Otd. Sporov. Rast. Bot. Inst. Komarova Akad. Nauk S.S.S.R., 13: 145.
- Kurtzman, C. P. (2011, January). Plylogeny of the ascomycetous yeasts and the renaming of Pichia anomala to Wickerhamomyces anomalus. *Antonie van Leeuwenhoek*, 99(1), 13-23.
- Kurtzman, C., & Fell, J. (1998). The yeasts : a taxonomic study, 4th ed.
- Kurtzman, C., Fell, J., & Boekhout, T. (2011). *The Yeasts : a taxininuc study* (5th ed.). Elsevier Science and Technology.
- Kurtzman, C., Robnett, C., & Basehoar-Powers, E. (2008). Phylogenetic relationships among species of Pichia, Issatchenkia and Williopsis determined from multigene sequence analysis, and the proposal of Barnettozyma gen. nov., Lindnera gen. nov. and Wickerhamomyces gen. FEMS Yeast Res, 939-954.



- Lamping, E., Ranchod, A., Nakamura, K., Tyndall, J., Niimi, K., Holmes, A., & et al. (2009). Abc1p is a multidrug efflux transporter that tips the balance in favor of innate azole resistance in Candida krusei. *Antimicrob Agents Chemother*, 354-369.
- Li, E., Liu, A., Xue, B., & Liu, Y. (2011). Yeast species associated with spontaneous wine fermentation of Cabernet Sauvignon from Ningxia, China. *World J. Microbiol. Biotechnol.*, 2475-2482.
- Liang, H., Zhang, A., Wu, Z., Liu, C., & Zhang, W. (2016). Characterization of Microbial Community during the Fermentation of Chinese Homemade paocai; a Traditional Fermented Vegetable Food. *Food Sci. Technol. Res.*, 467-475.
- Limon, J., Skalski, J., & Underhill, D. (2017). Commensal Fungi in Health and Disease. *Cell Host Microbe*, 156-165.
- Lopandic, K., & et al. (2008). Molecular profiling of yeasts isolated during spontaneous fermentations of Austrian wines. *FEMS Yeast Res.*, 1063-1075.
- Lopes, C., Rodriguez, M., Sangorrin, M., Querol, A., & Caballero, A. (2007). Patagonian wines: the selection of an indigenous yeast starter. *J. Ind. Microbiol. Biotechnol.*, 539-546.
- Loureiro, V., & Malfeito-Ferreira, M. (2003). Spoilage yeasts in the wine industry. *International Journal of Food Microbiology*, 23-50.
- Loureiro, V., & Querol, A. (1999). The prevalence and control of spoilage yeasts in foods and beverages. *Trends Food Sci Technol*, 356-365.
- Marquina, D., Peres, C., Caldas, F., Marques, J., Peinado, J., & Spencer-Martins, I. (1992). Characterization of the yeast population in olive brines. *Lett. Appl. Microbiol.*, 279-283.
- Miguel, M., Collela, C., de Almeida, E., Dias, D., & Schwan, R. (2015). Physiochemical and microbiological description of Caxiri a cassava and corn alcoholic beverage . *Int. J. Food Sci. Technol.*, 2537-2544.
- Moyes, D., Wilson, D., Richardson, D., Mogavero, J., Tang, S., Wernecke, J., & et al. (2016). Candidalysin is a fungal peptide toxin critical for mucosal infection. *Nature*, 64-68.
- Naglik, J., Gaffen, S., & Hube, B. (2019). Candidalysin: discovery and function in Candida albicans infections. *Curr Opin Microbiol*, 100-109.
- Nemcova, K., Breierova, E., Vadkertiova, R., & Molnarova, J. (2015). The diversity of yeasts associated with grapes and musts of the Strekov winegrowing region, Slovakia. *Folia Microbiol. (Praha)*, 103-109.
- New Zealand government-owned Crown Research Institute. (2014). *Plant & Food Research*. Retrieved Feb 4, 2019, from Growing Futures: Unique yeasts for winemaking: http://www.plantandfood.co.nz/growingfutures/case-studies/unique-yeasts-for-winemaking
- N'guessan, K., Brou, K., Jacques, N., Casaregola, S., & Dje, K. (2011). Identification of yeasts during alcoholic fermentation of tchapalo, a traditional sorghum beer from Cote d'Ivoire. *Antonie Van Leewenhoek*, 855-864.



- Nisiotou, A., Chorianopoulos, N., Nychas, G.-J., & Panagou, E. (2010). Yeast heterogeneity during spontaneous fermentation of black Conservolea olives in different brine solutions. *J. Appl. Microbiol.*, 396-405.
- Nuobariene, L., Arneborg, N., & Hansen, A. (2014). Phytase Activie Yeasts Isolated from Bakery Sourdoughs.
- Ocon, E., & et al. (2010). Quantitative and qualitative analysis of non-Saccharomyces yeasts in spontaneous alcoholic fermentations. *Eur. Food Res. Technol.*, 885-891.
- Ogunremi, O., Sanni, A., & Agrawal, R. (2015). Probiotic potentials of yeasts isolated from some cerealbased Nigerian traditional fermented food products. *J. Appl. Microbiol.*, 797-808.
- Osorio-Cadavid, E., Chaves-Lopez, C., Tofalo, R., Paparella, A., & Suzzi, G. (2008). Detection and identification of wild yeasts in Champus, a fermented Colombian maize beverage. *Food Microbiol*, 771-777.
- Padilla, B., Gil, J., & Manzanares, P. (2016). Past and Future of Non-Saccharomyces Yeasts: From Spoilage Microorganisms to Biotechnological Tools for Improving Wine Aroma Complexity. *Front. Microbiol.*, 411.
- Pardo, I., Garcia, M., Zuniga, M., & Uruburu, F. (1989). Dynamics of Microbial Populations during Fermentation of Wines from Utiel=Requena Region of Sptain. *Appl. Environ. Micobiol*, 539-41.
- Pereira-Dias, S., Potes, M., Marinho, A., Malfeito-Ferreira, M., & Loureiro, V. (2000). Characterisation of yeast flora isolated from an artisanal Portugese ewe's cheese. *Int. J. Food Microbiol*, 55-63.
- Perez-Torrado, R., & Querol, A. (2016). Opportunistic strains of Saccharomyces cerevisiae: A potential risk sold in food products. *Front. Microbiol.*, 1-5.
- Pfaller, M. D. (2010). Results from the artemis disk global antifungal surveillance study, 1997 to 2007: A 10.5-year analysis of susceptibilities of candida species to fluconazole and voriconazole as determined by CLSI standardized disk diffusion. *J Clin Microbiol*, 1366-1377.
- Pfaller, M., Diekema, D., Steel-Moore, L., Denys, G., Staley, C., Dipersio, J., & et al. (2004). Twelve years of fluconazole in clinical practice: Global-trends in species distribution and fluconazole susceptibility of bloodstream isolates of Candida. *Clin Microbiol Infect*, 11-23.
- Pichia kluyveri. (2019, January 29). Retrieved from Mycobank: http://www.mycobank.org/BioloMICS.aspx?TableKey=14682616000000089&Rec=965&Fields=A II
- Pilo, F., & et al. (2018). Saccharomyces cerevisiae poulations and other yeasts associated with indigenous beers (chicha) of Ecuador. *Brazilian J. Microbiol.*, 808-815.
- Pretscher, J., Fischkal, T., Branscheidt, S., Jager, L., Schlander, M., & et al. (2018). Yeasts from different habitats and their potential as biocontrol agents. *Fermentation*.
- RASFF portal | Food Safety. (2019, Feb 12). Retrieved from https://ec.europa.eu/food.safety/rasff/portal\_en.



- Raspor, P., Milek, D., Polanc, J., Smole Mozina, S., & Cadez, N. (2006). Yeasts isolated from three varieties of grapes cultivated in different locations of the Dolenjska vine-growing region, Slovenia. *Int. J. Food Microbiol.*, 97-102.
- Ricci, A. e. (2017). Scientific Opinion on the update of the list of QPS-recommended biological agents intentionally added to food or feed as notified to EFSA. *EFSA J.*
- Rocha, R., & Wilson, R. (2019). Essential , deadly, enigmatic: Polyamine metabolism and roles in fungal cells. *Fungal Biol Rev*, 47-57.
- Saarela, M., Matto, J., & Mattila-Sandholm, T. (2002). Safety Aspects of Lactobacillus and Bifidobacterium Species Originating from Human Oro-gastrointestinal Tract or from Probiotic Products. *Microb. Ecol. Health Dis.*, 234-241.
- Sabate, J., Cano, J., Esteve-Zarzoso, B., & Guilllamon, J. (2002). Isolation and identification of yeasts associated with vineyard and windery by RFLP analysis of ribosomal genes and mitochondrial DNA. *Microbiology Res.*, 267-274.
- Sandven, P., Nilsen, K., Digranes, A., Tjade, T., & Lassen, J. (1997). Candida norvegensis: A fluconazoleresistant species. *Antimicrob Agents Chemother*, 1375-1376.
- Santiago-Urbina, J., Arias-Garcia, J., & Ruiz-Teran, F. (2015). Yeast species associated with spontaneous fermentation of taberna, a traditional palm wine from southeast of Mexico. *Ann. Microbiol.*, 287-296.
- Schaffrath, R., Meinhardt, F., & Klassen, R. (2018). Yeast Killer TOxins: Fundimentals and Applications. In *The Mycota XV Physiology and Genetics* (2nd Edition ed., pp. 87-118).
- Seyedmousavi, S., & al., e. (2018). Fungal infections in animals: A patchwork of different situations. *Med. Mycol.*, S165-S187.
- Sipiczki, M. (2016). Overwintering of Vineyard Yeasts: Survival of Interacting Yeast Communities in Grapes Mummified on Vines. *Front. Microbiol*, 212.
- Spencer, D., Spencer, J., De Figueroa, L., & Heluane, H. (1992). Yeasts associated with rotting citrus fruits in Tucuman, Argentina. *Mycology Res*, 891-892.
- Starmer, W., Ganter, P., & Aberdeen, V. (1992). Geographic distribution and genetics of killer phenotypes for the yeast Pichia kluyveri across the United States. *Appl Environ Microbiol*, 990-997.
- Strauss, M., Jolly, N., Lambrechts, M., & van Rensburg, P. (2001). Screening for the production of extracellular hydrolytic enzymes by non-Saccharomyces wine yeasts. J. Appl. Microbiol., 182-190.
- Suranska, H., Vranova, D., Omelkova, J., & Vadkertiova, R. (2012). Monitoring of yeast population isolated during spontaneous fermentation of Moravian wine. *Chem. Pap.*, 861-868.
- T. Alcohol and Tobacco Tax and Trade Bureau. (n.d.). *Federal Register Labeling and Advertising of Wines, Distilled Spirits and Malt Beverages*. Retrieved Feb 12, 2019, from Title 27 of the Code of



Federal Regulations, Part 7: https://www.federalregister.gov/documents/2007/07/31/E7-14774/labeling-and-advertising-of-wines-distilled-spirits-and-malt-beverages

- Vadkertiova, R., Molnarova, J., Vranova, D., & Slavikova, E. (2012). Yeasts and yeast-like organisms associated with fruits and blossoms of different fruit trees. *Canada Journal of Microbiology*, 1344-1352.
- Vaudano, E., Quinterno, G., Costantini, A., Pulcini, L., Pessione, E., & Garcia-Moruno, E. (2019). Yeast distribution in Gringnolino grapes growing in a new vineyard in Piedmont and the technological characterization of indigenous Saccharomyces spp. strains. *International Journal of Food Microbiology, 289*, 154-161.
- Villa-Carvajal, M., Querol, A., & Belloch, C. (2006, August). Identification of species in the genus Pichia by restriction of the internal transcribed spacers (ITS1 and ITS2) and the 5.8S ribosomal DNA gene. Antonie Van Leeuwenhoek, 90(2), 171-181.
- Vu, D., Groenewald, M., Szoke, S., Cardinali, G., Eberhardt, U., Stielow, B., & et al. (2016). DNA barcoding analysis of more than 9,000 yeast isolates contributes to quantitative thresholds for yeast species and genera delimitation. *Stud Mycol*(85), 91-105.
- Wei, J., Zhang, Y., Yuan, Y., Dai, L., & Yue, T. (2019). Characteristic fruit wine production via reciprocal selection of juice and non-Saccharomyces species. *Food Microbiology*, 66-74.
- Whaley, S., Berkow, E., RFybak, J., Nishimoto, A., Barker, K., & Rogers, P. (2017). Azole antifungal resistance in Candida albicans and emerging non-albicans Candida Species. *Front Microbiol*, 1-12.
- Wiederhold, N. (2017). Antifungal resistance: current trends and future strategies to combat. *Infect Drug Resist*, 249-259.
- Xiao, M., Chen, S.--., Kong, F., Fan, X., Cheng, J., Hou, X., & et al. (2018). Five-year China hospital invasive fungal surveillance net (CHIF-NET) study of invasive fungal infections caused by noncandidal yeasts: Species distribution and azole susceptibility. *Infect Drug Resist*, 1659-1667.
- Yapar, N. (2014). Epidemiology and risk factors for invasive candidiasis. *Ther. Clin. Risk Manag.*, 95-105.
- Zagorc, T., & et al. (2001). Indigenous wine killer yeasts and their application as a starter culture in wine fermentation. *Food Microbiol*, 441-451.
- Zahavi, T., Droby, S., Cohen, L., Weiss, B., & Ben-Arie, R. (2002). Characterization of the yeast flora on the surface of grape berries in Israel.



# NEER™ Product Information Version: 3 PI GLOB EN 07-02-2018

#### Description

NEER<sup>™</sup> is a pure culture of *Pichia kluyveri* to be used in non-alcoholic fermented food and beverages: malt or other cereals, fermented based products, fruits, nuts or vegetables juices. The product is delivered as a deeply frozen (-45°C) bag ready for direct inoculation; it does not require re-hydration or acclimatization.

Culture composition: Pichia kluyveri.

Material No:	715390
Size	1000 U
Туре	Bag(s) in box

Color: Format: Form: Pale, yellowish brown F-DVS Frozen liquid

# Storage

-50 °C / -58 °F

#### Shelf life

When stored according to recommendation the product has a shelf life of 18 months.

#### Dosage

It is recommended to use one bag 1000 U in 50000 L / 500 hl / 13200 US gallons

#### Application

This specific and pure strain of Pichia kluyveri ensures a safe and reliable start to fermentation in wort, fruit or vegetables juices. It gives producers the opportunity to boost fruit flavours, optimising the conversion of soluble derived fruit flavour precursors into volatile flavours, increasing: flavor intensity, spectrum and longevity, sweetness intensity and smoothering the final product.

Product fermented with NEER<sup>™</sup> will have simultaneously several of the following improved features:

- Increased fruit flavour intensity
- Larger spectrum of fruit flavours adding complexity
- Increased sweetness perception
- Lower volatile acidity
- Rounder mouth-feel

#### Directions for use

1. Defrosting step: Open the freezer, take one box of product. Open the box with gloves; remove the cap protection placed on top of the bag and place the bag in a bucket of lukewarm water (30°C) for 5 to 10 minutes. This step will help unstick the frozen block from the plastic bag.

2. Activation: Due to the unique production process the *Pichia kluyveri* yeast cells are already activated for inoculation. No further activation is required.

3. Direct inoculation: Cut the top of the bag containing the frozen liquid yeast from end to end with scissors. Pour the content (frozen block of yeast) into your tank. Note: Never place the product in a -18°C freezer, follow carefully the instruction and move the product out of the -45°C freezer right before inoculation. SO  $_2$  will reduce the culture population. Check sulfites before inoculation, limit SO  $_2$  dosage to the minimum possible and always refer to the maximum level indicated for the product.

#### Technical Data

#### www.chr-hansen.com

Page: 1 (3)

The information contained herein is to the best of our knowledge and belief, true and accurate and the product(s) mentioned herein do(es) not infringe the intellectual property rights of any third party. The product(s) may be covered by pending or issued patents, registered or unregistered trademarks, or similar intellectual property rights. All rights reserved.



#### NEER™

Product Information Version: 3 PI GLOB EN 07-02-2018

#### Fermentation characteristics

Flavors	Acidic balance	Mouth-feel	Other
Enhance fruit flavors (thiols, terpenes,	Low volatile acidity	Medium production of	Low production of $SO_2$
esters)	Low acetic acid	polysaccharides	
Very low volatile phenols			
Very low H <sub>2</sub> S			
No diacetyl production			

#### Timing for inoculation

Dhysiological data

Application related - consult Chr. Hansen Application specialist

Parameter	Value(s)	Comment
Temperature*		
Tolerance limits	10-28°C (50-82°F)	
Optimum	15-25°C (59-77°F)	
SO <sub>2</sub> tolerance*	45 ppm	
Alcohol tolerance*	6.0%	
		Check YAN or FAN
Nitrogen requirements	medium	before inoculation
		Application related -
	From 0 to 5% depending	consult Chr. Hansen
Ethanol production	applied conditions	Application specialist
Glycerol yield	5-8 g/L	Standard

\* note that these inhibitory factors are antagonistic towards each other.

The individual tolerances are valid only if other conditions are favourable.

#### Legislation

The product is intended for use in food. Chr. Hansen 's cultures comply with the general requirements on food safety laid down in Regulation 178/2002/EC and with Council Regulation (EC) No 606/2009 of 10 July 2009, as amended.

#### Food Safety

No guarantee of food safety is implied or inferred should this product be used in applications other than those stated above. Should you wish to use this product in another application, please contact your Chr. Hansen representative for assistance.

#### Labeling

No labeling required, however please consult local legislation if in doubt.

#### Trademarks

Product names, names of concepts, logos, brands and other trademarks referred to in this document, whether or not appearing in large print, bold or with the ® or TM symbol are the property of Chr. Hansen A/S or an affiliate thereof or used under license. Trademarks appearing in this document may not be registered in your country, even if they are marked with an ®.

#### www.chr-hansen.com

The information contained herein is to the best of our knowledge and belief, true and accurate and the product(s) mentioned herein do(es) not infringe the intellectual property rights of any third party. The product(s) may be covered by pending or issued patents, registered or unregistered trademarks, or similar intellectual property rights. All rights reserved.



#### NEER™

Product Information Version: 3 PI GLOB EN 07-02-2018

#### Additional Information

Chr. Hansen's worldwide facilities and the personnel of our Application and Technology Center are at your disposal with assistance and instructions.

#### Technical support

Chr. Hansen's Application and Product Development Laboratories and personnel are available if you need further information.

#### GMO Information

In accordance with the legislation in the European Union<sup>\*</sup> <u>NEER<sup>M</sup> does not contain GMOs and does not contain GM</u> <u>labeled raw materials<sup>\*\*</sup></u>. In accordance with European legislation on labeling of final food products<sup>\*\*</sup> we can inform that the use of <u>NEER<sup>M</sup> does not trigger a GM labeling</u> of the final food product. Chr. Hansen's position on GMO can be found on: www.chr-hansen.com

\* Directive 2001/18/EC of the European Parliament and of the Council of 12 March 2001 on the deliberate release into the environment of genetically modified organisms with later amendments, and repealing Council Directive 90/220/EEC.

\*\* Regulation (EC) No 1829/2003 of the European Parliament and of the Council of 22 September 2003 on genetically modified food and feed with later amendments.

Regulation (EC) No 1830/2003 of the European Parliament and of the Council of 22 September 2003 concerning the traceability and labeling of genetically modified organisms and the traceability of food and feed products produced from genetically modified organisms amending Directive 2001/18/EC, and with later amendments.

#### Allergen Information

List of common allergens in accordance with the US Food Allergen Labeling and	Present as an
Consumer Protection Act of 2004 (FALCPA) and EU Regulation 1169/2011/EC with later	ingredient in
amendments	the product
Cereals containing gluten* and products thereof	No
Crustaceans and products thereof	No
Eggs and products thereof	No
Fish and products thereof	No
Peanuts and products thereof	No
Soybeans and products thereof	No
Milk and products thereof (including lactose)	No
Nuts* and products thereof	No
List of allergens in accordance with EU Regulation 1169/2011/EC only	
Celery and products thereof	No
Mustard and products thereof	No
Sesame seeds and products thereof	No
Lupine and products thereof	No
Mollusks and products thereof	No
Sulphur dioxide and sulphites (added) at concentrations of more than	
10 mg/kg or 10 mg/litre expressed as SO <sub>2</sub>	No

\* Please consult the EU Regulation 1169/2011 Annex II for a legal definition of common allergens, see European Union law at: www.eur-lex.europa.eu



# NEER™

**Product Specification** 

Form:	Frozen DVS
Material No:	715390
Culture	
Composition:	Pichia kluyveri

Performance

- Total cell count cfu/g
- Purity Coliforms cfu/g E.coli Lactic acid bacteria cfu/g Mould cfu/g Staphylococci Salmonella \*

\* Environmental and statistically based product testing is carried out on an ongoing basis, details can be supplied on request.

References and analytical methods are available upon request The information contained herein is to our knowledge true and correct and presented in good faith. No guarantee against patent infringement is implied or inferred.

Storage and shelf life:

See labels and product packaging

Page: 1/1

www.chr-hansen.com

Version: 29/JAN/2017 English

Electronically generated - signature not required

Specification >1.0E+09

**Specification** 

Absent in 1 g

Absent in 1 g

Absent in 25 g

<100

<1000

<1000



# HACCP evaluation of Application: Low alcohol beer

Introduction	Generic HACCP risk evaluations of Application of Cultures & Enzymes is part of our ISO 22000 certification
inter ou de crom	

# Table of contents

Procedure1	
Records7	
Flow	

# Responsibility

Who	What
Innovation responsible	Fill in template with all information's regarding all informations
HACCP responsible	Confirm results and make overall conclusion
Corporate Quality Partner	Approve conclusion

Yeast fermentation in beer including alcoholic fermentation,				
Procedure	and protection			



# Innovation HACCP evaluation Application: Low Alcohol Beer, pasteurized

Project owne	er:	cation responsible_ Hentie Swiegers / Wine Business Group				
Innovation Application Kristine Bjerre Kristine						
Intended Use		Yeast including both <i>Saccharomyces</i> and non- <i>Saccharomyces</i> in low alcohol (< 3 vol%), pasteurized beer				
	Product			nal Health:		
		Dairy: V		Wine		
				Other	r: X	
	Dosage of Culture in Final Application	Cultures included: S. cerevisiae, T. delbrueckii, and K. thermotolerans: All fluid bed dried P. kluyveri: Frozen product Intended inoculation level in final application is 0.1 g/L for frozen culture (1 kg for 10,000 L), and 0.1-0.2 g/L for dried cultures 500 g package for 25-50 hl. See appendix for more information on inoculation levels.				
			All within the range: 0.01-1.0 g/l or <b>max 0.1 %</b>			
c	Is the application	Yes:			No: X	
Application	already covered by a HACCP evaluation?	Reference to HACCP for generic application		Prepare HACCP for New application See below		
	Specifications in this intended Application	Pathogen		According to external specifications		Reference
	Pathogens	Salmonella (/25g)		Absent		
	Fill in appropriate	Listeria monocytogenes (/25g)		Not on specifications		
	microorganisms	S. aureus (cfu/g)		<1		
	and specification	Enterobacteriaceae (cfu/	g)	<1		
		<b>E. coli</b> (/1g) Only on specs for Prelude, Concerto, and Merit		Absent		
		Moulds (cfu/g)		< 1000		
		B. cereus (cfu/g)		Not on specifications		
		* 5x25 g testet (<5/125 g)	-			
SU	Other Contaminants	Microorganism				
Specifications in this intended		Acetic acid bacteria < 10000 (cfu/g)		000		
cificati in this ntended	CH- routine analysis	Lactic acid bacteria <1E+05 (cfu/g)				

To be filled in by Innovation Application responsible



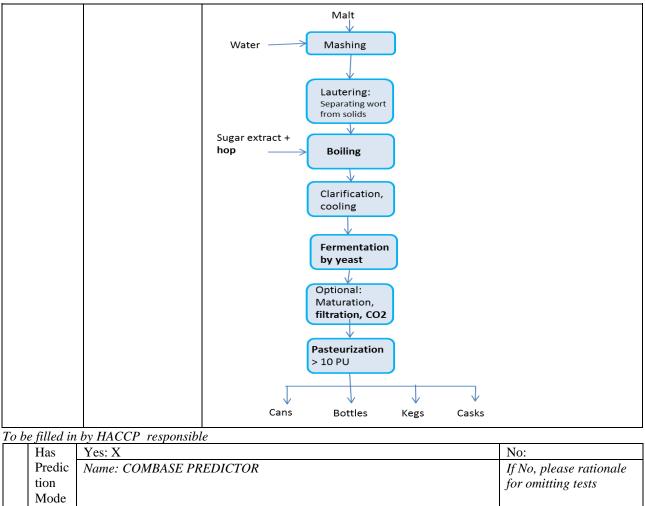
#### HACCP evaluation New Application template Low alc beer Document number: Error! Unknown document property name. Version: Error! Unknown document property name.

Fill in appropriate microorganisms and specification				
Hurdle 1: Aw Water activity in Food produced with culture	Aw= 0.98 => No hurdle	Not critical for Safety: X	Yes critical for Safety: PI –Info	
Hurdle 2: pH	pH=< 4.5	Not critical for Safety:	<b>Yes c</b> ritical for Safety: <b>X</b> <i>PI –Info</i>	
Hurdle 3. Temperature T °C	T °C=25	Not critical for Safety: X	Yes critical for Safety: PI –Info	
Hurdle 4: Max Storage time days/weeks/months	days/ weeks/ months	Not critical for Safety: X	<b>Yes c</b> ritical for Safety: <i>PI –Info</i>	
Hurdle 5: please specify	Hop iso-α-acids and Pasteurization	Not critical for Safety:	Yes critical for Safety: X PI –Info	
Pathogens relevant in the Food in question, see tableCl. botulinum, E coli, Salmonella spp, and Yersinia can be r $\leq 4.5.$ gram-positive pathogens (L. monocytogenes and St. aureus)				
	microorganisms and specification by Innovation Applica Hurdle 1: Aw Water activity in Food produced with culture Hurdle 2: pH Hurdle 3. Temperature T °C Hurdle 4: Max Storage time days/weeks/months Hurdle 5: please specify Pathogens relevant in the Food in	microorganisms and specificationby Innovation Application responsible_Hurdle 1: Aw Water activity in Food produced with cultureHurdle 2: Hurdle 2: PHHurdle 3. T °C=25Temperature T °CHurdle 4: Max days/weeks/monthsHurdle 5: please specify nesses pecifyPathogens relevant in the Food in question, see tableCl. botulinum, E cal gram-positive path	microorganisms and specificationAw=Notby Innovation Application responsible_Hurdle 1: Aw Water activity in Food produced with cultureAw= $0.98 =>$ No hurdleNot critical for Safety: XHurdle 2: Hurdle 3. Temperature T °CpH=< 4.5	

#### To be filled in by Innovation Application responsible\_

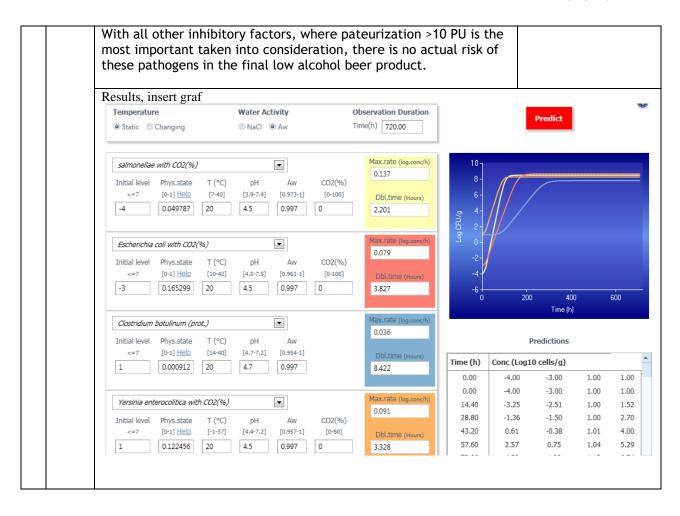
C	Hurdles critical for	$pH \le 4.5$ , Hops $\ge 15$ IBU, Pasteurization $\ge 10$ PU
ō v	safety, list values	Boiling
atic	during processing	Filtration
ic.		
pl rc		
Q d		
$\triangleleft$		





	tion			for om	itting tests		
	Mode			-			
	ls						
	been						
	perfor						
	med						
	Inocul	Salmonella absent in 25 g. => Theoretically 4/100 g					
de	ation						
0		Maximum inoculation level 1.0 g/l					
u u		Salmonella : $(4E-2)x(1E-3) = 4E-5 = Roughly 1 pr E-4 g final$	product				
ve		Max level 1CFU/25g= 4xE-2	•				
Ť.		Max dosage $0.1\% = 1xE-3$					
lic	Maximum inoculation level 1.0 g/l Salmonella : (4E-2)x(1E-3) = 4E-5 = Roughly 1 pr E-4 g final product Max level 1CFU/25g= 4xE-2 Max dosage 0.1% = 1xE-3 E. coli absent in 1 g. => Theoretically 100/100g. Max dosage = 1xE-3. Max level = 1 E. coli 1 x (1E-3) = 1 pr E-3 g final product						
ĕ							
P		E. coli: $1x (1E-3) = 1 \text{ pr } E-3 \text{ g final product}$					
		Clostridium and yersinia not in specifications.					
		Aw= 0.997	pH= 4.5		Temp= 20°C		
	nel		r		r		
	Salmonel la with	Conclusion: All 4 species tested will potentially be able to re	each	Storag	e time: 720 hrs		
	w	1E+08 CFU/g of final product after storage around 1 week. H		= 1  mo			
	Sí la	clostridium will only grow at $pH > 4.7$ .	10110101	1 1110			







### To be filled in by HACCP responsible

	Has Prediction	Yes:		No: X	
Listeria	Models been	Name: COMBASE PREDICTOR		If No, please rationale for omitting	
te	performed			tests	
, is	Inoculation				
	Listeria monocytogenes (+acetic acid)	Aw=	pH=		Temp= °C
model		Conclusion:	L	Storage time:	
Predictive m		Results, insert graf Not relevant since listeria pasteurization).	is inhibited by	the presence	of hops (and

### To be filled in by Innovation responsible

	Has Challenge tests	Yes:	No: X
ts	been performed		If No, please rationale for omitting
tests			tests
	Results of	•	
enge	Challenge test		
en			
hall			
Lh.			
	Reference		
	Challenge test		

### To be filled in by HACCP responsible

	Science supporting prolongation of shelf life beyond	Prevalence of contaminant in Production	Salmonella and Listeria not detected in 10.000 samples in plants and products.
References	estimation in Predictive Model	Menz, Aldred, and Vriesekoop. 2011. Growth and survival of foodborne pathogens in beer. Jour. of food protection. <b>74</b> (10):1670-1675	Presence of even low levels of hop iso-α-acids prohibit growth and limit survival of gram- positive pathogens ( <i>L.</i> <i>monocytogenes</i> and <i>S. aureus</i> )

To be filled in by HACCP team manager Innovation and Corporate Quality Partner

Conclusion	low alc. levels a growth of patho be employed an	by alcohol and alcohol free beer will because of the and high presence of nutrients involve a risk for ogens. Therefore pasteurization $> 10$ PU must always ad monitored for production of low alc. beer. ed for Low alcohol beer with ph<4,5 and h min. 10PU.
Date/ signature QA Innovation	27.11.2014	Tine Olesen
Date/signature	28.11.2014	Mirsad Ajanovic

Undates	Version	Who	What
opulles	1.	KBr,TiO	New



Low Terr	nperature	e <1	0 C					
Aw	pH							
		Au	ria	ot	sn	ili	alla	jia
		St.Au	Listeria	CI.bot	B.cereus	E.coli	Salmonella	Yersinia
					B.C		l	×e
							s	
0,83-0,92	<4,0							
	4,0-4,9	х						
	5,0-5,9	х						
	>6,0	х						
		_						
0,92-0,93	<4,0							
	4,0-4,9	х	х					
	5,0-5,9	х	х					
	>6,0	х	х					
0,93-0,94	<4,0							
	4,0-4,9	Х	х	х				
	5,0-5,9	х	х	х	х			
	>6,0	X	Х	Х	х			
0,94-0,95	<4,0						v	
0,94-0,95	4,0-4,9	x	x	x			x x	x
	5,0-5,9	x	x	x	х		x	x
	>6,0	x	x	x	x		x	x
	/0,0	^	^	^	^		^	^
>0,95	<4,0						x	
	4,0-4,9	х	x	x		х	х	х
	5,0-5,9	х	х	х	х	х	х	х
	>6,0	х	х	х	х	х	х	х

<b>Records</b> R: HACCP-Innovation/ Application or Innovation Quality Team-Site
---

Flow NA
---------



### Appendix

### **Beer pasteurization**

Kirk-Othmer "Food and Feed Technology"

but have made little progress in the United States to the present time.

Most beer in bottles or cans is *pasteurized*. That is, the beer is heated briefly to kill any microorganisms that might be present that could spoil beer flavor. As previously noted, no pathogenic (disease causing) organisms can survive in beer. Because brewers assure that the brewing process is extremely sanitary, few microbes enter the beer, and as a result they use a mild heat treatment. A pasteurizer is a large tunnel through which the beer cans or bottles move on an endless belt. The containers are sprayed with increasingly hot water to raise their temperature to 60-62°C. They are held at this temperature as long as required, and then cooled by water sprays. One pasteurization unit (PU) is 1 min at 60°C (or its heat equivalent) and most beers are pasteurized in the range of 5-15 PUs. There are two alternative techniques to tunnel pasteurization for dealing with the few microbes that might enter beer. The first is "flash" pasteurization in which the beer before packaging flows through a heat exchanger and is rapidly heated up and cooled down. This minimizes heat damage to the beer, but aseptic (sterile or microbe-free) packaging must follow and that is a challenging and expensive technology. Second, bacteria present can be filtered out of the beer by extremely tight membrane filtration. Again, aseptic packaging must follow this, but advantageously the beer can be marketed as "draft" beer in a bottle or can, because the definition of draft beer (in the United States) is that it be unpasteurized. The bottle is now ready for labeling. The packages are loading into six-pack holders, cased, and enter the warehouse from whence the product is distributed to wholesalers and eventually to consumers.



# Pasteurisation Units (PUs)

The Pasteurisation Unit (PU) is defined as relating to the sterilising effect observed when the product is held for one minute at a temperature termed the Base value. At this temperature therefore, 1 PU per minute is achieved.

Experiments on various mixtures of the common brewery biological contaminants showed that at temperatures over about 50°C there is an approximately ten-fold increase in sterilising effect for every 7°C increase in temperature. For example, if the time required to kill a population of micro-organisms at 60°C is found to be 5 minutes then if the temperature were to be increased to 67°C the time required would be only 0.5 minutes. The increase in temperature required to produce a ten-fold increase in kill rate is termed the Z value. The Z value in this case is therefore 7°C.

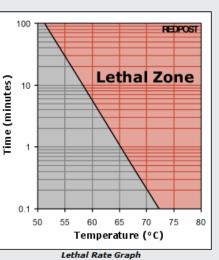
The formula for PUs is normally shown like this:  $PU = t \times 10^{rac{T-60}{7}}$ 

PU = Pasteurisation Units

t = Time(minutes)

 $T = Temperature({}^{\circ}C)$ 

Note this assumes a Base value of 60°C and a Z value of 7°C.



The same definition is often used for soft drinks and other beverages. Where there is any possibility of spore-forming bacterial contamination being present (for example in tomato juice) very much higher temperatures are required for pasteurisation. In these situations a Base value of 80°C and a Z value of 10°C is often used.



### **External specification for CH yeast:**

Pichia (and Concerto) dosage (cfu/ml) is just kept a little low compared to the other dried products in order not to having to apply large volumes to the wine.

	specs CFU/g	batch: CFU/g	Dosage accoring to PI	min CFU/ml
Frootzen	>1E+09	3008018	1 kg to 100 hl or 0.1 g/l	1,0E+05
Prelude	>5E+09	1022011: 1.9E+10	500 g to 25-50 hl or 0.1-0.2 g/l	5,0E+05
Concerto	>1E+09	705079	500 g to 25-50 hl or 0.1-0.2 g/l	1,0E+05
Merit	>1E+10	1082011	500 g to 25-50 hl or 0.1-0.2 g/l	1,0E+06

### CHR\_HANSEN

Improving food & health

### Viniflora® FrootZen

Product Specification

Form:	Liquid	
Item no:	703559	
Culture		
Composition:	Pichia kluyverii	
Performance		Specification
Total cell count cfu/g		>1.0E+09
Purity		Specification
<b>Purity</b> Coliforms cfu/g		Specification
	u/g	
Coliforms cfu/g	u/g	<100
Coliforms cfu/g Lactic acid bacteria cfu		<100 <100000
Coliforms cfu/g Lactic acid bacteria cfu Mould cfu/g		<100 <100000 <1000

References and analytical methods are available upon request

The information contained herein is to our knowledge true and correct and presented in good faith. No guarantee





## Viniflora® PRELUDE¿

**Product Specification** 

Form:Fluid Bed Dried YeastItem no:699118CultureTorulaspora delbrueckii

### Performance

Total cell count cfu/g

### Purity

Acetic acid bacteria cfu/g Coliforms cfu/g Escherichia coli Lactic acid bacteria cfu/g Mould cfu/g Staphylococci Salmonella spp. \*

### Specification

>=5.0E+09

### Specification

<10000 <100 Negative in 1 g <100000 <1000 Negative in 1 g Absent in 25 g



CHR HANSEN Improving food & health

# Viniflora® CONCERTO¿

Product Specification

Form:	Fluid Bed Dried Yeast
ltem no:	705079
Culture	
Composition:	Kluyveromyces thermotolerans

### Performance

Total cell count cfu/g

### Purity

Acetic acid bacteria cfu/g Coliforms cfu/g Escherichia coli Lactic acid bacteria cfu/g Mould cfu/g Staphylococci Salmonella spp. \*

### Specification

>=1.0E+09

### Specification

<10000 <100 Negative in 1 g <100000 <1000 Negative in 1 g Absent in 25 g

\* Environmental and statistically based product testing is carried out on an ongoing basis, details can be supplied on request.



Improving food & nearth

# Viniflora® MERIT

Product Specification

Form:	Fluid Bed Dried Yeast
ltem no:	673398
Culture	
Composition:	Saccharomyces cerevisiae

### Performance

Total cell count cfu/g

### Purity

Acetic acid bacteria cfu/g Coliforms cfu/g Escherichia coli Lactic acid bacteria cfu/g Mould cfu/g Staphylococci Yeasts cfu/g Salmonella spp. \*

# Specification >=1.0E+10

### Specification <10000 <100 Negative in 1 g <100000 <1000 Negative in 1 g

<100000

Absent in 25 g

\* Environmental and statistically based product testing is carried out on an ongoing basis, details can be supplied on request.



### NEER<sup>™</sup> GMO statement Material No: 715390 Version: 1 GMO EN 04-11-2016

### GMO statement concerning NEER™

According to the legislation in the European Union genetic modification occurs if certain techniques have been used<sup>\*</sup>. The same legislation also defines techniques, which will not result in genetic modification. In accordance with this legislation we can state that

NEER<sup>™</sup> does not contain GMOs and does not contain GM labeled raw materials\*\*.

### GM labeling information concerning NEER™

Legislation in the European Union states that a final food product must be labeled if it is a GMO itself, if it contains GMOs, or if it contains ingredients derived from GMOs<sup>\*\*</sup>. In accordance with European legislation on labeling of final food products we can inform that

The use of NEER<sup>™</sup> does not trigger a GM labeling of the final food product.

Chr. Hansen's position on GMO can be found on:

www.chr-hansen.com/About us/Policies and positions/Quality and product safety.

\* Directive 2001/18/EC of the European Parliament and of the Council of 12 March 2001 on the deliberate release into the environment of genetically modified organisms with later amendments, and repealing Council Directive 90/220/EEC. \*\* Regulation (EC) No 1829/2003 of the European Parliament and of the Council of 22 September 2003 on genetically modified food and feed with later amendments.

Regulation (EC) No 1830/2003 of the European Parliament and of the Council of 22 September 2003 concerning the traceability and labeling of genetically modified organisms and the traceability of food and feed products produced from genetically modified organisms amending Directive 2001/18/EC, and with later amendments.



Allergen Information Material No: 715390 Version: 1 AL EN 04-11-2016

List of common allergens in accordance with the US Food Allergen Labeling and Consumer Protection Act of 2004 (FALCPA) and EU Regulation 1169/2011/EC with later amendments.	Present as an ingredient in the product	Ingredient species or type
Cereals containing gluten* and products thereof	No	
Crustaceans and products thereof	No	
Eggs and products thereof	No	Not applicable
Fish and products thereof	No	
Peanuts and products thereof	No	Not applicable
Soybeans and products thereof	No	Not applicable
Milk and products thereof (including lactose)	No	Not applicable
Nuts* and products thereof	No	
List of allergens in accordance with EU Regulation 1169/2011/EC only		
Celery and products thereof	No	Not applicable
Mustard and products thereof	No	Not applicable
Sesame seeds and products thereof	No	Not applicable
Lupine and products thereof	No	Not applicable
Mollusks and products thereof	No	Not applicable
Sulphur dioxide and sulphites at concentrations of more than		
10 mg/kg or 10 mg/litre expressed as SO <sub>2</sub>	No	
Yes = Allergen labeling required		
No = Allergen labeling not required		

Please consult the EU Regulation 1169/2011 Annex II for a legal definition of common allergens, see European Union law at: http://eur-lex.europa.eu/

Page: 1 (1)

The information contained herein is to the best of our knowledge and belief, true and accurate and the product(s) mentioned herein do(es) not infringe the intellectual property rights of any third party. The product(s) may be covered by pending or issued patents, registered or unregistered trademarks, or similar intellectual property rights. Copyright © Chr. Hansen A/S. All rights reserved.



## Safety Data Sheet NEER™

Version: 3 GHS / EN Revision Date: 04-28-2020

### 1. IDENTIFICATION OF THE SUBSTANCE/PREPARATION AND OF THE COMPANY/UNDERTAKING

Phone: +1 414 607-5700

1.1	Product identifier	
	Product name:	NEER™
	Material No:	715390

- 1.2 Relevant identified uses of the substance or mixture and uses advised against Application: For beer or juice application.
- 1.3 Details of the supplier of the safety data sheet Supplier: Chr. Hansen Inc. 9015 West Maple Street 53214-4298 Milwaukee - WI

Headquarters:	Chr. Hansen A/S
	Boge Allé 10-12
	DK-2970 Horsholm
	Tel. +45 45 74 74 74

1.4Emergency telephone number<br/>Emergency telephone:+45 45 74 74 74

### 2. HAZARDS IDENTIFICATION

- 2.1 Classification of the substance or mixture The product is not classified.
- 2.2 Label elements

Signal Word None.

Hazard statements None

Precautionary statements None

2.3 Other hazards



Physical and Chemical Hazards: The hazardous properties of the product are considered to be limited.

Human health: Risk of local frostbite. Prolonged skin contact may cause redness and irritation.

The product does not contain any carcinogenic substances in amounts to be declared.

Environment: The harmful effects of the product in the environment are considered to be limited.

### 3. COMPOSITION/INFORMATION ON INGREDIENTS

3.2 Mixtures The product contains: yeast.

Does not contain substances that must be indicated according to current regulations.

4. FIRST-AID MEASURES

### 4.1 Description of first aid measures

Inhalation: Move into fresh air and keep at rest.

Skin contact: Remove contaminated clothes and rinse skin thoroughly with water.

Eye contact: Do not rub eye. Immediately flush with plenty of water for up to 15 minutes. Remove any contact lenses and open eyelids widely. If irritation persists: Seek medical attention and bring these instructions.

Ingestion: Rinse mouth thoroughly. If uncomfortable: Get medical attention.

- 4.2 Most important symptoms and effects, both acute and delayed Risk of local frostbite. Prolonged skin contact may cause redness and irritation.
- 4.3 Indication of any immediate medical attention and special treatment needed Symptomatic treatment.

### 5. FIRE-FIGHTING MEASURES

- 5.1 Extinguishing media Use fire-extinguishing media appropriate for surrounding materials.
- 5.2 Special hazards arising from the substance or mixture No specific precautions. The explosion limits and the flash point are stated in section 9.

### 5.3 Advice for firefighters Selection of respiratory protection for fire fighting: follow the general fire precautions indicated in the workplace.

### 6. ACCIDENTAL RELEASE MEASURES



- 6.1 Personal precautions, protective equipment and emergency procedures Avoid contact with skin and eyes. Follow precautions for safe handling described in this safety data sheet.
- 6.2 Environmental precautions Avoid discharge into drains, water courses or onto the ground.
- 6.3 Methods and material for containment and cleaning up Absorb spillage with suitable absorbent material. Flush contaminated area with plenty of water.
- 6.4 Reference to other sections For personal protection, see section 8. For waste disposal, see section 13.

#### 7. HANDLING AND STORAGE

7.1 Precautions for safe handling Safe handling advice: Avoid contact with skin and eyes. Observe good industrial hygiene practices.

Technical measures: Keep the workplace clean.

Technical precautions: No special precautions.

7.2 Conditions for safe storage, including any incompatibilities Store in tightly closed original container. Store at super frozen temperature conditions. For detailed information consult the PI sheet. Store in a dry place.

Technical measures for safe storage: No special precautions.

### 8. EXPOSURE CONTROLS/PERSONAL PROTECTION

- 8.1 Control parameters Exposure limits are listed below. No data - no exposure limits noted for ingredient(s).
- 8.2 Exposure controls

Engineering measures: Provide adequate ventilation.

Respiratory equipment: Not relevant, due to the form of the product. Risk of inhalation of dust or aerosols use suitable respirator. Use respiratory equipment with particle filter: EU: FFP3 filter [e.g. 3M 8835 mask] US: P100 filter [e.g. 3M 8293 mask]

For daily use of more than 3 hours a respirator with a powered air blower should be used.

Hand protection: Wear protective gloves against low temperatures.

Eye protection: Risk of contact: Wear goggles/face shield.

Skin protection: No special precautions.

Hygiene measures: Wash hands after contact.

Environmental Exposure Controls: None.



### 9. PHYSICAL AND CHEMICAL PROPERTIES

- 9.1 Information on basic physical and chemical properties
  - Appearance: Color: Odor: pH: Melting point: Boiling point: Decomposition temperature: Flash point: Relative density: Solubility:

Frozen liquid Pale, yellowish brown Fruity, Peptone-like 6,00 - 7,00 Not relevant Not relevant Not relevant Not relevant No data available Water soluble

9.2 Other information No information available.

### 10. STABILITY AND REACTIVITY

- 10.1 Reactivity None known.
- 10.2 Chemical stability Stable under normal temperature conditions and recommended use.
- 10.3 Possibility of hazardous reactions None known.
- 10.4 Conditions to avoid None known.
- 10.5 Incompatible materials None known.
- 10.6 Hazardous decomposition products None known.

### 11. TOXICOLOGICAL INFORMATION

11.1 Information on toxicological effects

Inhalation: Not relevant, due to the form of the product. Inhalation of high concentrations of dust or aerosols may cause toxic alveolitis. Symptoms like fever, coldshivering, coughing, difficulties in breathing, headache, muscle and joint pains etc. may appear 6 to 8 hours after exposure. The symptoms normally disappear completely over night without any treatment.

Skin contact: Risk of local frostbite. Prolonged contact may cause redness and irritation.

Eye contact: Risk of local frostbite. Ingestion: Risk of local frostbite. May irritate and cause malaise.

Specific effects: Pro-longed exposure to dust/aerosols containing microorganisms may cause allergic alveolitis. The product does not contain any carcinogenic substances in amounts to be declared.



### 12. ECOLOGICAL INFORMATION

### 12.1 Ecotoxicity The harmful effects of the product in the environment are considered to be limited.

- 12.2 Persistence and degradability The product is expected to be biodegradable.
- 12.3 Bioaccumulative potential Bioaccumulation: Is not expected to be bio-accumulable.
- 12.4 Mobility in soil The product is water soluble and may spread in water systems.
- 12.6 Other adverse effects None known.

#### 13. DISPOSAL CONSIDERATIONS

13.1 Waste treatment methods

Dispose of waste and residues in accordance with local authority requirements.

### 14. TRANSPORT INFORMATION

The product is not covered by international regulations on the transport of dangerous goods (IMDG, IATA, DOT).

14.1	UN	number
------	----	--------

<u>Air (ICAO/IATA):</u> 14.3 Transport hazard class(es) 14.4 Packing group	-
<u>Sea (IMDG):</u> 14.3 Transport hazard class(es) 14.4 Packing group EmS MFAG	
Land (DOT): 14.3 Transport hazard class(es) 14.4 Packing group	-
14.5 Environmental hazards Marine pollutant (IMDG): -	
14.6 Special precautions for user None known.	
14.7 Transport in bulk according to Annex II of MARPOL Not relevant.	73/78 and the IBC Code

### 15. REGULATORY INFORMATION



15.1 Safety, health and environmental regulations/legislation specific for the substance or mixture

NFPA: Health: 1 Fire: 0 Reactivity: 0 Other: -GHS regulation

Globally Harmonized System of Classification and Labelling of Chemicals (GHS)

16. OTHER INFORMATION

The user must be instructed in the proper work procedure and be familiar with the contents of these instructions.

The following sections contain revisions or new statements : 1.

Wording of Hazard Statements

The information in this Safety Data Sheet has been obtained from current and reliable sources. However, the data is provided without warranty, express or implied, regarding its correctness or accuracy. It is the user's responsibility to determine safe conditions for use of this product and to assume liability for loss injury, damage, or expense resulting from improper use of this product.



# Chr. Hansen GmbH

Certification

### Gießener Straße 94, D35415 Pohlheim, Germany

Bureau Veritas Certification Holding SAS, UK Branch certifies that the food safety management system of the above organization has been assessed and complies with the requirements of:

Standard

### FOOD SAFETY SYSTEM CERTIFICATION (FSSC) 22000 Certification scheme for food safety management systems, consisting of the following

elements:

ISO 22000:2005, ISO/TS 22002-1:2009 and Additional FSSC 22000 requirements V4.1

This certificate is applicable for the scope of

### Production of cultures for the food industry.

Product category: K

Certification cycle start date: 24-04-2018

Subject to the continued satisfactory operation of the organization's Management System, this certificate expires on: 23-04-2021

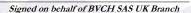
Original certification date: 23-09-2006

Certificate No./Version: DK009342-2

Contract No. 10435618

Issue date: 20-04-2018





Certification body address: 66 Prescot Street, London E1 8HG, United Kingdom Local office: Oldenborggade 25-31, DK-7000 Fredericia, Denmark

Further clarifications regarding the scope of this certificate and the applicability of the management system requirements may be obtained by consulting the organization. To check this certificate validity please contact fooddkmail@dk.bureauveritas.com This certificate remains the property of Bureau Veritas Certification Holding SAS - UK Branch Validity of this certificate can be verified in the FSSC 22000 database of certified organizations available on www.fssc22000.com.



0008





### **Chr. Hansen GmbH** Giessener Strasse 94, D-35415 Pohlheim, Germany

Bureau Veritas Certification Holding SAS – UK Branch certifies that the Management System of the above organization has been audited and found to be in accordance with the requirements of the management system standards detailed below.

Standard

# ISO 22000:2005

Scope of certification

Research, development, production, applied technology and distribution of biotechnological, functional and natural ingredients and processing aids for the food industry.

### **Product category: K**

Original cycle start date:	22-09-2006
Expiry date of previous cycle:	NA
Certification/Recertification Audit date:	NA
Certification/Recertification cycle start date:	24-04-2018

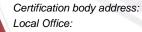
Subject to the continued satisfactory operation of the organization's Management System, this certificate expires on: **23-04-2021** 

Certificate No.: DK009343

Version:1 R

Revision date: 20-04-2018

MIX Paper from responsible sources FSC° C074440



5<sup>th</sup> Floor, 66 Prescot Street, London, E1 8HG, United Kingdom Oldenborggade 25-31, 7000 Fredericia, Denmark



Further clarifications regarding the scope of this certificate and the applicability of the Management System requirements may be obtained by consulting the organization. To check this certificate validity, please call **(+45) 77 311 000**.