



June 4, 2021

Center for Food Safety and Applied Nutrition
Office of Food Additive Safety
U.S. Food and Drug Administration
CPK-2 Building, Room 2092
5001 Campus Drive, HFS-225
College Park, MD 20740



Dear GRAS Filing Team:

Enclosed is a GRAS Determination entitled "GRAS Determination for the Use of Lacto-N-tetraose (LNT) in Selected Conventional Foods and Enteral Tube Feeding Formulas". We believe that this GRAS Determination should be considered as a new notification because Chr. Hansen A/S intends to expand the use of its lacto-N-tetraose ingredient to selected conventional foods and enteral tube feeding formulas.

We thank you for taking the time to review this GRAS Determination. Should you have additional questions, please let us know.

Sincerely,



Dietrich B. Conze, Ph.D.
Managing Partner

Enclosure: CD containing
Form 3667
Cover Letter
GRAS Determination for the Use of Lacto-N-tetraose (LNT) in Selected
Conventional Foods and Enteral Tube Feeding Formulas
References

GRAS Determination for the Use of Lacto-*N*-tetraose (LNT) in Selected Conventional Foods and Enteral Tube Feeding Formulas

Prepared for:

Chr. Hansen A/S¹
9015 W Maple St.
West Allis, WI 53214

Prepared by:

Spherix Consulting Group, Inc.
751 Rockville Pike, Unit 30-B
Rockville, MD 20852

May 18, 2021

¹ Jennewein Biotechnology GmbH is now Chr. Hansen HMO GmbH. The legal entity (including the same company identification number), manufacturing premises, manufacturing processes and quality systems and certifications remains the same.

All documentation bearing the name of Jennewein Biotechnologie GmbH is in the process of being updated to Chr. Hansen HMO GmbH/Chr. Hansen A/S as appropriate. This is however an ongoing process; Chr. Hansen assures that the documents released with the Jennewein Biotechnologie GmbH's name, remain valid until the full update is completed.

Likewise, updated certificates and commercial registrations will be issued by the relevant competent authorities in due course; meanwhile the current certificates and commercial registrations remain valid until further notice.

TABLE OF CONTENTS

I. SIGNED STATEMENT OF THE CONCLUSION OF GENERALLY RECOGNIZED AS SAFE (GRAS) AND CERTIFICATION OF CONFORMITY TO 21 CFR §170.205-170.260....	1
A. SUBMISSION OF GRAS NOTICE	1
B. NAME AND ADDRESS OF THE SPONSOR	1
C. COMMON OR USUAL NAME.....	1
D. TRADE SECRET OR CONFIDENTIAL INFORMATION.....	1
E. INTENDED USE	1
F. BASIS FOR GRAS DETERMINATION	2
G. PREMARKET APPROVAL	4
H. AVAILABILITY OF INFORMATION	4
I. FREEDOM OF INFORMATION ACT (FOIA).....	5
J. INFORMATION INCLUDED IN THE GRAS NOTIFICATION.....	5
II. IDENTITY, METHOD OF MANUFACTURE, SPECIFICATIONS, AND PHYSICAL OR TECHNICAL EFFECT OF THE NOTIFIED SUBSTANCE.....	6
A. COMMON OR USUAL NAME.....	6
B. CHEMICAL NAME	6
C. MOLECULAR FORMULA AND MASS.....	6
D. STRUCTURAL FORMULA	6
E. DESCRIPTION OF LACTO-N-TETRAOSE	6
F. PRODUCTION PROCESS.....	7
G. FINISHED PRODUCT SPECIFICATIONS	7
H. STABILITY	7
III. DIETARY EXPOSURE	10
A. INTENDED EFFECT	10
B. HISTORY OF EXPOSURE.....	10
C. INTENDED USES.....	10
D. ESTIMATED DAILY INTAKE	11
1. Estimated Daily Intake of LNT from Oral Electrolyte Solutions.....	11
2. Estimated Daily Intake of LNT from Selected Conventional Foods and Enteral Tube Feeding Formula	12
3. Food Consumption Survey Data.....	13
4. Food Usage.....	14

IV. SELF-LIMITING LEVELS OF USE	17
V. COMMON USE IN FOOD BEFORE 1958	18
VI. NARRATIVE ON THE CONCLUSION OF GRAS STATUS	19
A. SAFETY OF THE PRODUCTION ORGANISMS	20
B. ABSORPTION, DISTRIBUTION, METABOLISM, AND EXCRETION	20
C. TOXICOLOGY STUDIES	20
D. TOLERANCE STUDY IN NEONATAL PIGLETS.....	21
E. CLINICAL STUDIES.....	22
1. Clinical Studies with HMOs in Infants and Adults	22
2. Clinical Studies with Other Non-digestible Carbohydrates and Enteral Tube Feeding Formulas	36
3. Clinical Studies with Other Non-digestible Carbohydrates and Oral Electrolyte Solutions	46
F. ALLERGENICITY	50
G. REGULATORY APPROVALS AROUND THE WORLD.....	50
VII. SUPPORTING DATA AND INFORMATION.....	51
A. REFERENCES.....	51
B. EXPERT PANEL STATEMENT	62

LIST OF TABLES

Table 1. Intended Uses and Use Levels for LNT.....	1
Table 2. Product Specifications and Batch Data for Lacto- <i>N</i> -tetraose	8
Table 3. Comparison of Uses and Use levels That Are GRAS with the Intended Uses and Use Levels.....	11
Table 4. Estimated “All-user” Daily Intake (EDI) of LNT from Chr. Hansen A/S’s Food Uses by Population Group (2015-2016 NHANES Data)	15
Table 5. Cumulative Estimated “All-user” Daily Intake (EDI) of LNT in All Food Uses by Population Group (2015-2016 NHANES Data)	16
Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants.....	24
Table 7. Clinical Studies with Human Milk Oligosaccharides and Adults	32
Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding ¹	37
Table 9. Studies of Oral Electrolyte Solutions (OES) with Added Nondigestible Carbohydrate	48

LIST OF ABBREVIATIONS

2'-FL: 2'-Fucosyllactose
3-FL: 3-Fucosyllactose
3'-SL: 3'-Sialyllactose
6'-SL: 6'-Sialyllactose
Alb: Albumin
ALT: Alanine aminotransferase
araA: Arabinose isomerase
BMI: Body mass index
BW: Body weight
CBPI: Cytokinesis-block proliferation index
CFR: United States Code of Federal Regulations
CFU: Colony forming units
CHO: Chinese hamster ovary cells
CI: Confidence interval
COSY: Correlation spectroscopy
DSMZ: Deutsche Sammlung für Mikroorganismen und Zellkulturen
DW: Dry weight
EDI: Estimated Daily Intake
EFSA: European Food Safety Authority
EU: Endotoxin unit
F6PPK: Fructose-6-phosphate phosphoketolase
FCC: Food Chemicals Codex
FDA: United States Food and Drug Administration
FFDCA: Federal Food, Drug, and Cosmetic Act
FOIA: Freedom of information Act
FOS: Fructooligosaccharides
Fru-1,6-BP: Fructose-1,6-bisphosphate
Fru-6-P: Fructose-6-phosphate
FSSC: Food Safety System Certification
FUT: Fucosyltransferase
GI: Gastrointestinal
Glc-1-P: Glucose-1-phosphate

Glc-6-P: Glucose-6-phosphate

Gln-1-P: Glucosamine-1-phosphate

Gln-6-P: Glucosamine-6-phosphate

Glob: Globulin

GluNAc-6-P: *N*-acetylglucosamine-6-phosphate

GMO: Genetically modified organism

GMP: Good manufacturing practices

GOS: Galactooligosaccharides

GRAS: Generally Recognized As Safe

GRN: GRAS Notification

HCD: Historical control data

HDL-C: High-density lipoprotein cholesterol

HMBC: $^1\text{H}^{13}\text{C}$ -heteronuclear multiple bond correlation

HMO: Human milk oligosaccharides

HPAEC-PAD: High performance anion exchange chromatography coupled with pulsed amperometric detection

HSQC: $^1\text{H}^{13}\text{C}$ -Heteronuclear single quantum correlation

ICP-MS: Inductively coupled plasma mass spectrometry

IFN γ : Interferon gamma

LC-MS: Liquid chromatography coupled with mass spectrometry

LDL-C: Low-density lipoprotein cholesterol

LDPE: Low-density polyethylene

LNDFHI: lacto-*N*-difucohexaose I

LNnT: Lacto-*N*-neotetraose

LNT: Lacto-*N*-tetraose

LOD: Limit of detection

LOQ: Limit of quantitation

MCH: Mean corpuscular hemoglobin

MCV: Mean corpuscular volume

ND: Not detected

NHANES: National Health and Nutrition Examination Surveys

NIH: National Institutes of Health

NMR: Nuclear magnetic resonance

NOAEL: No Observed Adverse Effect Level

OECD: Organization for Economic Cooperation and Development

PCR: Polymerase chain reaction

Ph Eur: European Pharmacopoeia

pLNnH: Para-lacto-*N*-neohexaose

qPCR: Quantitative polymerase chain reaction

RI: Replicative index

TP: Total protein

UDP-Gal: UDP-galactose

UDP-Glc: UDP-glucose

UDP-GlcNAc: UDP-*N*-acetylglucosamine

**I. SIGNED STATEMENT OF THE CONCLUSION OF GENERALLY
RECOGNIZED AS SAFE (GRAS) AND CERTIFICATION OF
CONFORMITY TO 21 CFR §170.205-170.260**

A. SUBMISSION OF GRAS NOTICE

Chr. Hansen A/S is hereby submitting a GRAS notice in accordance with subpart E of part 170.

B. NAME AND ADDRESS OF THE SPONSOR

Chr. Hansen A/S
9015 W Maple St.
West Allis, WI 53214

C. COMMON OR USUAL NAME

Lacto-*N*-tetraose (LNT)

D. TRADE SECRET OR CONFIDENTIAL INFORMATION

This notification does not contain any trade secret or confidential information.

E. INTENDED USE

Chr. Hansen A/S intends to use LNT as an ingredient in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas (Table 1).

Table 1. Intended Uses and Use Levels for LNT	
Intended Uses	Intended Use Levels (g/kg or g/L)
Toddler formula (Go and Grow by Similac®)	0.6
Milk-based meal replacement beverages for children (Pediasure®)	6
Cereals, prepared, ready-to-serve, for infants and young children	6
Cereals, dry instant, for infants and young children	6
Meal replacement drinks for adults including dairy and non-dairy drinks for weight reduction and formulas for pregnant women	6
Non-carbonated drinks (e.g. fitness water, thirst quenchers, sports and isotonic drinks)	3
Meal-replacement bars, and breakfast bars	6
Bars, snack	6
Oral Electrolyte Solutions	0.6
Enteral tube feeds used as sole source nutrition (Ensure®, Glucerna®, and Boost®)	10

F. BASIS FOR GRAS DETERMINATION

Lacto-*N*-tetraose for the intended use and use level specified above has been shown to be safe and GRAS, using scientific procedures, under the Federal Food, Drug, and Cosmetic Act (FFDCA), as described under 21 CFR §170.30(b). The safety of the intake of LNT has been determined to be GRAS by demonstrating that the safety of the intended level of intake is generally recognized as safe by experts qualified by both scientific training and experience to evaluate the safety of substances directly added to food and is based on generally available and accepted information.

The use of LNT as an ingredient for the intended use in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas has been determined to be safe through scientific procedures set forth under 21 CFR §170.30(b) based on the following:

1. The subject of this GRAS Determination is a spray-dried, powdered food ingredient that contains not less than 75% LNT dry weight. The remaining components include carbohydrate by-products, ash, and moisture.
 - a. LNT is a neutral, non-fucosylated oligosaccharide in human milk.
 - b. Published studies show that the amount of LNT in human milk ranges from 0.003 to 6.7 g/L, with means and medians ranging from 0.1 to 3.9 and 0.2 to 2.1 g/L, respectively.
 - c. Human milk oligosaccharides, including LNT, are resistant to the digestive enzymes in the gastrointestinal tract, poorly absorbed, and pass through the gastrointestinal tract where they are either fermented by the microbiota or excreted unchanged.
2. The subject of this GRAS Determination is also the subject of GRAS Notice 923, which received a “no questions” letter on February 2, 2021 for the use of LNT in non-exempt term infant formula.
 - a. The subject of this GRAS Determination is manufactured using a genetically engineered strain of *Escherichia coli* BL21(DE3) by Chr. Hansen A/S in Food Safety System Certification (FSSC) 22000-, ISO 9001:2015-, GMP-, and/or International Featured Standards Food 6.1-compliant facilities. Chr. Hansen A/S is a Food Facility registered with FDA.

- b. The genetically engineered strain of *Escherichia coli* BL21(DE3) used by Chr. Hansen A/S is not toxigenic and not capable of DNA transfer to other organisms and has the same virulence profile as *E. coli* BL21(DE3).
 - c. All raw materials, processing aids, and food contact substances are GRAS and/or conform to the specifications stated in 21 CFR and/or the Food Chemicals Codex (FCC).
 - d. Process procedures and product specifications are in place to control the levels of carbohydrate by-products, as well as heavy metals, microbes, and production organism-derived DNA and possible endotoxin, ensuring a consistent, safe, food-grade finished ingredient.
 - e. The available stability studies indicate a shelf-life of two years when stored from the date of production under ambient conditions.
 - f. Use of the subject of this GRAS Determination in the intended selected conventional foods and enteral tube feeding formulas results in mean and 90th percentile estimated daily intakes (EDIs) of 1.02 and 2.33 g/day (0.015 and 0.035 g/kg bw/day) for consumers not less than 2 years old.
 - g. Use of the subject of this GRAS Determination in selected conventional foods and enteral tube feeding formulas results in mean and 90th percentile cumulative estimated daily intakes (EDIs) of 0.463 and 1.05 g/day (0.007 and 0.016 g/kg bw/day) for consumers not less than 2 years old.
 - h. Use of the subject of this GRAS Determination in oral electrolyte solutions results in an estimated daily intake of 0.8 – 1.6 g LNT (equivalent to 59.3 – 118.5 mg of LNT/kg bw/day assuming a 13.5 kg toddler and 11.4 – 22.9 mg of LNT/kg bw/day assuming a 70 kg adult). Because OESs are intended for short term use, intake of LNT from OESs will not impact the cumulative LNT intake resulting from the use of LNT in selected conventional foods and enteral tube feeding formulas.
3. Genotoxicology and rat subchronic toxicology studies published by Phipps et al. (2018) show that LNT is not genotoxic and has a no observed adverse effect level (NOAEL) of 4 g/kg bw/day, which was the highest dose tested.
 4. The safety of exposure to Chr. Hansen A/S's LNT per its intended uses and intended use levels is supported by:

- a. Data demonstrating the qualitative and quantitative similarities between the subject of this GRAS Notice and the LNT ingredient tested in the pivotal genotoxicology and subchronic toxicology studies conducted by Phipps et al. (2018);
- b. The lack of genotoxicity and no observed adverse effect level (NOAEL) for LNT established in the 90-day subchronic dietary toxicology conducted by Phipps et al. (2018);
- c. Published genotoxicology, 90-day subchronic dietary toxicology, and neonatal piglet studies with mixtures containing the subject of this GRAS Determination (Parschat et al., 2019; Hanlon, 2020);
- d. Clinical data showing the ingestion of synthetic forms of HMOs is well tolerated in infants up to 1.0 g/day and adults up to 20 g/day;
- e. Clinical data showing that the use of other non-digestible carbohydrates in infants, adults, enteral tube feeding products, and oral electrolyte solutions is well tolerated up to 63 g/day;
- f. The GRAS status of the subject of this GRAS Determination for use in infant formula (GRN 923);
- g. The GRAS status of LNT for use in selected conventional foods (GRN 833).

Therefore, LNT is safe and GRAS at the proposed level of addition to the toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas is, therefore, excluded from the definition of a food additive, and may be used in the U.S. without the promulgation of a food additive regulation by the FDA under 21 CFR.

G. PREMARKET APPROVAL

The notified substance is not subject to the premarket approval requirements of the FD&C Act based on our conclusion that the substance is GRAS under the conditions of intended use.

H. AVAILABILITY OF INFORMATION

The data and information that serve as the basis for this GRAS determination will be available for review and copying at reasonable times at the office of Dietrich Conze, PhD, Managing Partner, Spherix Consulting Group Inc., at 751 Rockville Pike, Unit 30-B, Rockville, MD 20852; Telephone: 240-367-6089; Email: dconze@spherixgroup.com; or be sent to FDA upon request.

I. FREEDOM OF INFORMATION ACT (FOIA)

Parts 2 through 7 of this notification do not contain data or information that is exempt from disclosure under the FOIA.

J. INFORMATION INCLUDED IN THE GRAS NOTIFICATION

To the best of our knowledge, the information contained in this GRAS notification is complete, representative and balanced. It contains both favorable and unfavorable information, known to Chr. Hansen A/S and pertinent to the evaluation of the safety and GRAS status of the use of this substance.



Signature of Authorized Representative of
Chr. Hansen A/S

20 May 2021
Date

II. IDENTITY, METHOD OF MANUFACTURE, SPECIFICATIONS, AND PHYSICAL OR TECHNICAL EFFECT OF THE NOTIFIED SUBSTANCE

A. COMMON OR USUAL NAME

Lacto-*N*-tetraose (LNT; CAS No. 14116-68-8)

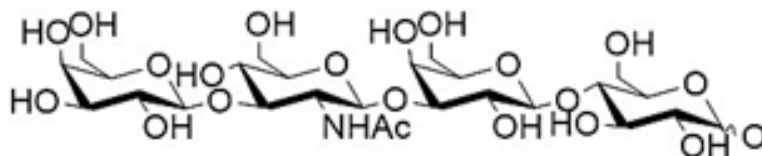
B. CHEMICAL NAME

β -D-Galactopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)-D-glucopyranose

C. MOLECULAR FORMULA AND MASS

C₂₆H₄₅NO₂₁; 707.632 g/mol

D. STRUCTURAL FORMULA



E. DESCRIPTION OF LACTO-*N*-TETRAOSE

Lacto-*N*-tetraose is one of the most abundant oligosaccharides and one of the most abundant core structures in human milk. It is a linear tetrasaccharide consisting of a terminal D-galactose linked through a β -(1 \rightarrow 3) bond to *N*-acetyl-D-glucosamine (GlcNAc), linked through a β -(1 \rightarrow 3) bond to D-galactose, linked through a β -(1 \rightarrow 4) bond to the reducing end D-glucose (see Structural Formula) and can be further decorated with fucosyl or sialyl residues, resulting in over 200 different HMOs identified to date. It is also an isomer of LNnT (lacto-*N*-neotetraose), contains the same monosaccharide moieties with the linkage between the terminal galactose and GlcNAc being β -(1-4), and considered to be a Type I oligosaccharide due to the presence of the characteristic galactose (Gal) (β 1 \rightarrow 3) *N*-acetyl-D-glucosamine [lacto-*N*-biose I (LNB)] unit (Urashima et al., 2012).

The lacto-*N*-tetraose (LNT) that is the subject of this GRAS Determination is also the subject of GRAS Notice (GRN) 923, which received a “no questions” letter from FDA in February 2, 2021. As summarized in GRN 923, the subject of this GRAS Notification is produced by Chr. Hansen A/S using fermentation and a genetically engineered strain of *Escherichia coli* BL21 (DE3). The ingredient is purified from the culture medium and residual impurities include lactose and carbohydrate by-products. Importantly, the structure of the LNT

that is the subject of this GRAS notification is structurally identical to the LNT in human milk as confirmed by liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS), ¹H, ¹³C, double-quantum filtered ¹H¹H-COSY, phase-sensitive ¹H¹³C-heteronuclear single quantum correlation (HSQC), and phase-sensitive ¹H¹³C-heteronuclear multiple bond correlation (HMBC) NMR spectroscopy.

Chr. Hansen A/S intends to expand the intended uses to toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas.

F. PRODUCTION PROCESS

Because the production process was extensively reviewed in GRN 923, the description of the production strain and manufacturing process are incorporated by reference (see pages 6 – 10 of GRN 923). The only difference between the LNT that was the subject of GRN 923 and this GRAS Determination is that cobalt is no longer added to the fermentation medium. Briefly, LNT is produced by fermentation using the genetically engineered strain of *E. coli* BL21(DE3) *JBT-LNT* in a contained, sterile production line at the Chr. Hansen A/S production facility, which is Food Safety System Certification (FSSC) 22000- and ISO 9001:2015- compliant, and FDA-registered (Registration # 1303109037512). Following synthesis, LNT is purified from the fermentation medium and the resulting concentrate is dried into a powder. All other manufacturers involved in the LNT manufacturing are Chr. Hansen A/S-qualified and either GMP-, ISO-, and International Featured Standards Food 6.1-compliant.

G. FINISHED PRODUCT SPECIFICATIONS

To ensure a consistent food-grade product that is free of genetically modified ingredients, each batch of LNT is evaluated against the same product specifications as those specified in GRN 923 using compendial or validated methods that are fit-for-use. Data from five non-consecutive batches of LNT show that the manufacturing process reproducibly produces a finished product that complies with the product specifications and removes the production organism from the finished product (Table 2).

H. STABILITY

The production strain and finished ingredient stability were extensively reviewed in GRN 923. Therefore, the summaries of the genomic and finished product stability are incorporated by reference (see pages 12 -15 of GRN 923). Briefly, the production strain is not expected to lose its ability to produce a consistent finished product because it contains stably integrated genes and no plasmids or episomal vectors. The shelf-life of the finished ingredient is expected to be 2 years from the date of production when stored under ambient conditions.

Table 2. Product Specifications and Batch Data for Lacto-*N*-tetraose

Parameter	Analytical method	Specification	Batch Number				
			16125019	11043019	11047029	21005010	26143020
Physical Parameters							
Appearance (Color) ⁴	visual	White to ivory-colored	Complies	Complies	Complies	Complies	Complies
Appearance (Form) ⁴		Spray-dried powder	Complies	Complies	Complies	Complies	Complies
Chemical Parameters							
Lacto- <i>N</i> -Tetraose ⁴	HPAEC-PAD	≥ 75 % DW (w/w)	84.2	81.8	81.6	83.5	92.7
Sum of Other carbohydrates ^{4,5}		≤ 25 % (% Area)	4.5	3.9	4.4	3.3	5.4
Lactose ⁴		≤ 5 % (% Area)	1.8	0.3	0.4	1.1	< LOQ
Lacto- <i>N</i> -triose II ⁴		≤ 5 % (% Area)	2.4	2.5	2.5	0.5	< LOQ
para-Lacto- <i>N</i> -Hexose ⁴		≤ 5 % (% Area)	1.1	1.6	1.7	1.4	0.5
Glucose/galactose ⁴		≤ 5 % (% Area)	< LOQ	1.1	1.1	< LOQ	< LOQ
Protein ⁴	Nanoquant (modified Bradford)	≤ 100 µg/g	< LOQ	< LOQ	< LOQ	< LOQ	< LOQ
Ash ¹	ASU L 06.00-4	≤ 1 % (w/w)	0.44	0.23	0.34	0.35	0.14
Moisture ⁴	KF titration	≤ 9% (w/w)	5.5	5.4	5.5	5.6	5.8
Endotoxins ³	Ph. Eur. 2.6.14	≤ 10 EU/mg	0.228	0.034	0.031	< LOQ	< LOQ
Aflatoxin M1 ¹	DIN EN ISO 14501	≤ 0.025 µg/kg	< LOQ	< LOQ	< LOQ	< LOQ	< LOQ
GMO residues ²	qPCR	≤ 0.01%	Negative	Negative	Negative	Negative	Negative
Heavy Metals							
Arsenic ¹	ASU L 00.00-135 – ICP-MS	≤ 0.2 mg/kg	ND	ND	ND	ND	ND
Cadmium ¹		≤ 0.1 mg/kg	ND	ND	ND	ND	ND
Lead ¹		≤ 0.02 mg/kg	ND	ND	0.016	ND	ND
Mercury ¹		≤ 0.5 mg/kg	ND	ND	ND	ND	ND
Microbes							
Standard Plate Count ¹	ISO 4833-2	≤ 10000 cfu/g	< 10	< 10	20	< 10	< 10
Yeast and Mold ¹	ISO 21527-2	≤ 100 cfu/g	< 20	< 20	< 20	< 20	< 20
<i>Enterobacteriaceae</i> ¹	ISO 21528-1	≤ 10 cfu/g	< 10	< 10	< 10	< 10	< 10
<i>Salmonella</i> ¹	ISO 6579	Absent/25 g	Absent	Absent	Absent	Absent	Absent
<i>Cronobacter sakazakii</i> ¹	ISO/TS 22964	Absent/10g	Absent	Absent	Absent	Absent	Absent
Abbreviations: DW, dry weight; cfu, colony forming units; STDEV, standard deviation; KF, Karl-Fischer; HPAEC-PAD, high performance anion exchange chromatography coupled with pulsed amperometric detection; qPCR, quantitative polymerase chain reaction; ICP-MS, inductively coupled plasma mass spectrometry; EU, endotoxin unit; Ph Eur., European Pharmacopoeia; LOQ, limit of quantitation; ND, not detected.							
¹ Determined by the Institut für Produktqualität GmbH, which is a DIN EN ISO/IEC 17025-accredited laboratory; Ash LOQ = 0.01 %. Arsenic limit of detection (LOD) = 0.05 mg/kg; Cadmium LOD = 0.01 mg/kg; Mercury LOD = 0.005 mg/kg; Lead LOD = 0.01 ppm; Aflatoxin M1 LOQ = 0.025 µg/kg.							
² Determined by GeneCon International GmbH, which is a DIN EN ISO/IEC 17025-accredited laboratory. Limit of detection = 0.01% of the finished product.							

Table 2. Product Specifications and Batch Data for Lacto-*N*-tetraose

Parameter	Analytical method	Specification	Batch Number				
			16125019	11043019	11047029	21005010	26143020
³ Determined by Mikrobiologisches Labor. Dr. Michael Lohmeyer GmbH, which is a DIN EN ISO/IEC 17025-accredited laboratory; limit of quantitation = 0.005 EU/mg. ⁴ Determined by Chr. Hansen A/S using internally validated methods. Protein LOQ = 10 µg/g; carbohydrate by-products with a percent area greater than 0.5% (limit of quantitation) are considered. ⁵ Defined as the sum of N-acetylglucosamine (GlcNAc); lacto- <i>N</i> -biose (LNB); galactosyllactose; glucosyllactose; LNT-(LNB) _n , which includes LNT-(LNB) ₂ , also known as para-lacto- <i>N</i> -octaose (pLNO); LNT-(LNB) _n -N-GlcNAc, which includes LNT-(LNB) ₀ , also known as LNT-GlcNAc; total unspecific impurities. “n” denotes the number of repeating LNB units attached to LNT and can be any number starting from zero.							

III. DIETARY EXPOSURE

A. INTENDED EFFECT

The intended effect of adding LNT to toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas is to increase LNT intake.

B. HISTORY OF EXPOSURE

LNT is one of the most abundant oligosaccharides in human milk and its levels have been quantitated in 30 studies with greater than 5 donors (Thurl et al., 1997; Thurl et al., 2010; Galeotti et al., 2012; Erney et al., 2000; Coppa et al., 1999; Sumiyoshi et al., 2003; Asakuma et al., 2008; Leo et al., 2010; Gabrielli et al., 2011; Bao et al., 2013; Nakhla et al., 1999; Chaturvedi et al., 1997; Chaturvedi et al., 2001; Asakuma et al., 2011). The results of 11 of these studies are summarized in a systematic review conducted by Thurl et al. (2017). Although the levels of LNT in human milk vary with ethnicity, Secretor and Lewis-blood group status, lactation period, and term vs preterm birth, the available data show that the concentration of LNT in human milk generally ranges from 0.003 to 6.7 g/L with means and medians ranging from 0.1 to 3.9 and 0.2 to 2.1 g/L, respectively. Thurl et al. (2017) reported a means and 95% percentile limits for the amount of LNT in the milk of secretor mothers of term infants of 0.79 g/L and 0.59 – 0.98 g/L and preterm infants of 1.04 g/L and 0.39 – 1.68 g/L.

Synthetic forms of LNT are used in infant formula and selected conventional foods (GRN 833; GRN 923). Thus, humans are exposed to LNT either through the ingestion of human milk and/or products containing synthetic forms of LNT.

C. INTENDED USES

LNT is GRAS for use in non-exempt infant formula at 0.8 g/L and selected conventional foods at 0.6 to 20 g/kg (GRN 833; GRN 923). The subject of GRN 833 is GRAS for use in selected in non-exempt infant formula at 0.8 g/L and conventional foods at 0.6 to 20 g/kg. The subject of this GRAS Determination is GRAS for use in only non-exempt term infant formula at 0.8 g/L as a substitute for other forms of LNT that are GRAS (GRN 923). Chr. Hansen A/S intends to expand the use of the subject of GRN 923 to toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas at levels ranging from 0.6 to 10 g/L (Table 3). Importantly, these expanded uses include new uses, substitutional uses for other forms of LNT that are GRAS for use in infant formula and conventional foods, and increases to LNT use levels in uses that have already been determined GRAS. Therefore, a cumulative estimated daily intake must be calculated using the maximum use level for all uses to determine if Chr. Hansen A/S's expanded intended uses increase overall LNT exposure.

Table 3. Comparison of Uses and Use levels That Are GRAS with the Intended Uses and Use Levels

Uses That Are GRAS ¹	Use Levels That are GRAS (g/kg or g/L) ¹	Intended Uses	Chr. Hansen A/S Intended Use Levels (g/kg or g/L)	Maximum Use Level Used for Cumulative EDI Calculation (g/kg or g/L)
Non-exempt term infant formula	0.8	-	-	0.8
Toddler formula	0.6	Toddler formula (Go and Grow by Similac [®])	0.8	0.8
-	-	Milk-based meal replacement beverages for children (Pediasure [®])	6	6
Baby food	5	-	-	5
-	-	Cereals, prepared, ready-to-serve, for infants and young children	6	6
-	-	Cereals, dry instant, for infants and young children	6	6
Drinks for children	0.6	-	-	0.6
Meal replacement drinks including dairy and non-dairy drinks for weight reduction	2	Meal replacement drinks for adults including dairy and non-dairy drinks for weight reduction and formulas for pregnant women	6	6
Sports, isotonic, and energy drinks including fortified water and soft drinks	1	Non-carbonated drinks (e.g. fitness water, thirst quenchers, sports and isotonic drinks)	3	3
Meal replacement bars for weight loss	20	Meal-replacement bars, and breakfast bars	6	20
Bars, snack	10	Bars, snack	6	10
Unflavored, pasteurized milk	1	-	-	1
Buttermilk	1	-	-	1
Flavored milk	1	-	-	1
Yogurt	10	-	-	10
-	-	Oral Electrolyte Solutions	0.6	- ²
Enteral tube feeds used as sole source nutrition		Enteral tube feeds used as sole source nutrition (Ensure [®] , Glucerna [®] , and Boost [®])	10	10

¹Obtained from GRN 833 and GRN 923.

²Not included in the cumulative estimated daily intake calculation because these products are intended for short-term use only.

D. ESTIMATED DAILY INTAKE

1. Estimated Daily Intake of LNT from Oral Electrolyte Solutions

Oral electrolyte solutions (OESs), such as Pedialyte, are specially formulated to replenish fluids and minerals and recommended to be used under medical supervision to prevent dehydration caused by vomiting, diarrhea, exercise, travel, or heat exhaustion. Conditions of use state 1-2 L of OES, such as Pedialyte, may be ingested per day to maintain proper hydration,

however, a medical professional should be consulted if vomiting, fever, or diarrhea continues beyond 24 hr or if consumption needs are greater than 2 L/day. Due to its infrequent use and low number of users within the database (1 user), calculation of an estimated daily intake (EDI) using the National Center for Health Statistics' (NCHS) 2015-2016 National Health and Nutrition Examination Surveys (NHANES) is not appropriate.

A conservative EDI can be calculated from the intended use of OES. Consumption of a maximum of 1-2 L of an OES per day at a use level of 0.8 g of LNT/L would result in a daily intake of 0.8 – 1.6 g of LNT (equivalent to 59.3 – 118.5 mg of LNT/kg bw/day assuming a 13.5 kg toddler and 11.4 – 22.9 mg of LNT/kg bw/day assuming a 70 kg adult). Because OESs are intended for short term use, intake of LNT from OESs will not impact the cumulative LNT intake resulting from the use of LNT in other select conventional foods and enteral tube feeding formulas.

2. Estimated Daily Intake of LNT from Selected Conventional Foods and Enteral Tube Feeding Formula

Estimates for the intake of Chr. Hansen A/S's intended uses of LNT were based on the food uses and Chr. Hansen A/S's use level in Table 3, in conjunction with food consumption data included in the National Center for Health Statistics' (NCHS) 2015-2016 National Health and Nutrition Examination Surveys (NHANES) (CDC, 2018; USDA, 2018). Nutritional beverages such as Boost, Ensure, and Glucerna were used as surrogates for enteral and tube-feeding formulas. A total of 110 food codes representative of each approved use were chosen from the Food and Nutrition Database for Dietary Studies (FNDDS) for the corresponding biennial NHANES survey. Calculations from NHANES for the mean and 90th percentile intakes were performed for Chr. Hansen A/S's representative food uses of LNT.

To determine the impact of Chr. Hansen A/S's intended uses on the cumulative estimated daily intake of LNT from all uses, a cumulative estimated daily intake was calculated using the maximum use level for all uses with the food consumption data included in the National Center for Health Statistics' (NCHS) 2015-2016 National Health and Nutrition Examination Surveys (NHANES) (Table 3; CDC, 2018; USDA, 2018). A total of 638 food codes representative of each use were chosen from the Food and Nutrition Database for Dietary Studies (FNDDS) for the corresponding biennial NHANES survey. As described previously, nutritional beverages such as Boost, Ensure, and Glucerna were used as surrogates for enteral and tube-feeding formulas. Calculations from NHANES for the mean and 90th percentile intakes were performed for all representative food uses of LNT.

3. Food Consumption Survey Data

a. Survey Description

The most recent NHANES data for the years 2015-2016 are available for public use. NHANES are conducted as a continuous, annual survey, and are released in 2-year cycles. In each cycle, approximately 10,000 people across the U.S. completed the health examination component of the survey. Any combination of consecutive years of data collection is a nationally representative sample of the U.S. population. It is well established that the length of a dietary survey affects the estimated consumption of individual users and that short-term surveys, such as the typical 1-day dietary survey, overestimate consumption over longer time periods (Hayes et al., 2014). Because two 24-hour dietary recalls administered on 2 non-consecutive days (Day 1 and Day 2) are available from the NHANES 2015-2016 survey, these data were used to generate estimates for the current intake analysis.

The NHANES provide the most appropriate data for evaluating food-use and food-consumption patterns in the United States, containing 2 years of data on individuals selected via stratified multistage probability sample of a civilian non-institutionalized population of the U.S. NHANES survey data were collected from individuals and households via 24-hour dietary recalls administered on 2 non-consecutive days (Day 1 and Day 2) throughout all 4 seasons of the year. Day 1 data were collected in-person in the Mobile Examination Center (MEC), and Day 2 data were collected by telephone in the following 3 to 10 days, on different days of the week, to achieve the desired degree of statistical independence. The data were collected by first selecting Primary Sampling Units (PSUs), which were counties throughout the U.S. Small counties were combined to attain a minimum population size. These PSUs were segmented and households were chosen within each segment. One or more participants within a household were interviewed. Fifteen PSUs are visited each year. For example, in the 2009-2010 NHANES, there were 13,272 persons selected; of these 10,253 were considered respondents to the MEC examination and data collection. 9754 of the MEC respondents provided complete dietary intakes for Day 1 and of those providing the Day 1 data, 8,405 provided complete dietary intakes for Day 2. The release data do not necessarily include all the questions asked in a section. Data items may have been removed due to confidentiality, quality, or other considerations. For this reason, it is possible that a dataset does not completely match all the questions asked in a questionnaire section. Each data file has been edited to include only those sample persons eligible for that particular section or component, so the numbers vary.

In addition to collecting information on the types and quantities of foods being consumed, the NHANES surveys collect socioeconomic, physiological, and demographic information from individual participants in the survey, such as sex, age, height and weight, and

other variables useful in characterizing consumption. The inclusion of this information allows for further assessment of food intake based on consumption by specific population groups of interest within the total population.

Sample weights are incorporated with NHANES surveys to compensate for the potential under-representation of intakes from specific population groups as a result of sample variability due to survey design, differential non-response rates, or other factors, such as deficiencies in the sampling frame (CDC, 2006; USDA, 2012).

b. Statistical Methods

Consumption data from individual dietary records, detailing food items ingested by each survey participant, were collated by computer in Octave and used to generate estimates for the intake of LNT by the U.S. population. Estimates for the daily intake of LNT represent projected 2-day averages for each individual from Day 1 and Day 2 of NHANES data; these average amounts comprised the distribution from which mean and percentile intake estimates were produced. Mean and percentile estimates were generated incorporating sample weights to provide representative intakes for the entire U.S. population. “All-user” intake refers to the estimated intake of LNT by those individuals consuming food products containing LNT. Individuals were considered users if they consumed one or more food products containing LNT on either Day 1 or Day 2 of the survey.

4. Food Usage

The estimated “all-user” total intakes of LNT from Chr. Hansen A/S’s intended uses only from 110 proposed food uses listed in NHANES in the U.S. by population group is described in Table 4. In summary, 9.38% of the total U.S. population 2+ years was identified as consumers of Chr. Hansen A/S’s intended uses of LNT in the 2015-2016 survey. The mean intakes by LNT consumers age 2+ from Chr. Hansen A/S’s intended food uses were estimated to be 1.02 g/person/day or 0.015 g/kg body weight/day. The heavy consumer (90th percentile) intakes were estimated to be 2.33 g/person/day or 0.035 g/kg body weight/day. The highest consumers on a mean EDI by body weight basis were ages 13 months to 2 years at 0.038 g/kg body weight/day (0.479 g/day).

The cumulative estimated “all-user” total intakes of LNT from 638 proposed food uses listed in NHANES in the U.S. by population group is described in Table 5. In summary, 62.0% of the total U.S. population 2+ years was identified as consumers of LNT from the selected food uses in the 2015-2016 survey. The mean intakes by all LNT consumers age 2+ from all LNT food uses were estimated to be 0.463 g/person/day or 0.007 g/kg body weight/day. The heavy consumer (90th percentile) intakes were estimated to be 1.05 g/person/day or 0.016 g/kg body

weight/day. The highest consumers on a mean EDI by body weight basis were ages 13 months to 2 years at 0.025 g/kg body weight/day (0.31 g/day).

Importantly, a comparison of the mean and 90th percentile EDIs of LNT ages 2+ from Chr. Hansen A/S's food uses and all food uses shows that the EDI decreases from 1.02 and 2.33 to 0.463 to 1.05 g/day, which is likely due to dilution from a broader range of uses and an increased number of users (Table 2; compare Tables 4 and 5, respectively). Thus, Chr. Hansen A/S's intended uses and use levels do not increase LNT exposure.

Table 4. Estimated "All-user" Daily Intake (EDI) of LNT from Chr. Hansen A/S's Food Uses by Population Group (2015-2016 NHANES Data)								
Population Group	N users	N population	% Users	Mean mass (kg)	Mean EDI (g)	90th % EDI (g)	Mean EDI (g/kg)	90th % EDI (g/kg)
ages 0-6 months	49	197	24.87	7.00	0.052	0.091	0.007	0.013
ages 7-12 months	72	207	34.78	9.44	0.143	0.277	0.015	0.029
ages 13 months-2 years	44	535	8.22	12.56	0.479	1.090	0.038	0.087
ages 2-5 years	69	915	7.54	16.92	0.587	1.49	0.035	0.088
ages 6-12 years	146	1505	9.70	36.58	0.797	1.83	0.022	0.050
ages 13-19 years	145	1143	12.69	67.35	1.04	2.42	0.015	0.036
ages 20 years and up	513	5748	8.92	80.76	1.11	2.46	0.014	0.031
ages 2 years and up	873	9311	9.38	67.35	1.02	2.33	0.015	0.035

Table 5. Cumulative Estimated “All-user” Daily Intake (EDI) of LNT in All Food Uses by Population Group (2015-2016 NHANES Data)								
Population Group	N users	N population	% Users	Mean mass (kg)	Mean EDI (g)	90th % EDI (g)	Mean EDI (g/kg)	90th % EDI (g/kg)
ages 0-6 months	142	197	72.08	7.00	0.131	0.195	0.019	0.028
ages 7-12 months	169	207	81.64	9.44	0.210	0.531	0.022	0.056
ages 13 months-2 years	373	535	69.72	12.56	0.31	0.662	0.025	0.053
ages 2-5 years	566	915	61.86	16.92	0.329	0.692	0.019	0.041
ages 6-12 years	975	1505	64.78	36.58	0.352	0.64	0.010	0.017
ages 13-19 years	821	1143	71.83	67.35	0.459	0.896	0.007	0.013
ages 20 years and up	3415	5748	59.41	80.76	0.519	1.30	0.006	0.016
ages 2 years and up	5777	9311	62.04	67.35	0.463	1.05	0.007	0.016

IV. SELF-LIMITING LEVELS OF USE

This part does not apply.

V. COMMON USE IN FOOD BEFORE 1958

This part does not apply.

VI. NARRATIVE ON THE CONCLUSION OF GRAS STATUS

The subject of this GRAS determination is a synthetic form of LNT, which is a non-digestible oligosaccharide found in human milk, also known as a human milk oligosaccharide (HMO). Non-fucosylated neutral HMOs, such as LNT, constitute 42-55% of the total HMO fraction in human milk and LNT is one of the most abundant non-fucosylated neutral HMOs (Van Niekerk et al., 2014; Hong et al., 2014; Thurl et al., 2017). As summarized in GRN 923, published studies indicate that the levels of LNT in human milk ranges from approximately 0.003 to 6.7 g/L with means and medians ranging from 0.1 to 3.9 and 0.2 to 2.1 g/L, respectively.

To obtain a thorough and comprehensive understanding of the safety of LNT per the intended uses and use levels, searches of the published scientific literature were conducted using Pubmed. All articles published up to May 10, 2021 that evaluated the safe use of LNT in conventional foods, oral electrolytes solutions (OESs), and enteral tube feeding formulas were retrieved and reviewed. Consistent with the requirements of the GRAS standard, Chr. Hansen A/S considered the totality of publicly available data and information relevant to the safety of LNT including the use of other HMOs in selected conventional foods and oral electrolyte solutions, and non-digestible carbohydrates in enteral tube feeding products. This document includes the entire results of these searches.

Currently, two synthetic LNT products are GRAS (GRN 833; GRN 923). The subject of GRN 833 is manufactured by Glycom A/S using a genetically engineered strain of *E. coli* and is GRAS for use in non-exempt term infant formula and selected conventional foods. The subject of GRN 923, which is the subject of this GRAS determination, is also produced using a genetically engineered strain of *E. coli* and GRAS for use in non-exempt term infant formula. As summarized in GRN 923, the subjects of GRN 833 and this GRAS determination are structurally identical to the LNT in human milk, qualitatively comparable and quantitatively similar to each other, and supported by a battery of published genotoxicology, subchronic toxicology, and neonatal piglet tolerance studies conducted with LNT and mixtures of HMOs containing LNT. LNT is not genotoxic, has a NOAEL (no observed adverse effect level) of at least 4 g/kg bw/day, and, when administered with other HMOs, is well-tolerated in neonatal piglets. Additionally, publicly available clinical data show that the ingestion of other HMOs, such as 2'-fucosyllactose (2'-FL), 3'-sialyllactose (3'-SL) and 6'-sialyllactose (6'-SL), and other non-digestible carbohydrates are also well tolerated in infants, children, adults, oral electrolyte solutions and susceptible population groups that received enteral tube feeding formulas.

Because infants are considered a susceptible population group from a safety perspective and the subject of this GRAS determination is qualitatively comparable and quantitatively similar to the LNT tested by Phipps et al. (2018) and the subject of GRN 833 (Scientific Committee on Food, 1998; GRN 833, 2019; GRN 923, 2021), there is reasonable certainty that

the use of the subject of this GRAS determination per the intended uses will also be safe in children, adults, and enteral tube feeding formulas. Chr. Hansen A/S therefore concludes that the subject of this GRAS Determination is GRAS as an ingredient in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas at the intended use levels.

A. SAFETY OF THE PRODUCTION ORGANISMS

The safety of the host organism *E. coli* BL21(DE3) and the production strain *JBT-LNT* was thoroughly summarized in GRN 923. Therefore, the summaries of the safety of the host organism and the production strain are incorporated by reference (see pages 26 and 27 of GRN 923). Importantly, because it was engineered with genes with known function, which do not confer toxicogenicity, virulence, or DNA, using site-specific homologous recombination or transposition, *JBT-LNT* is non-toxicogenic, not capable of DNA transfer to other organisms, and has the same virulence profile as *E. coli* BL21(DE3). Therefore, based on the widespread use of *E. coli* BL21(DE3) as a host strain, the strategy used to genetically engineer *JBT-LNT*, and its comprehensive characterization, use of *JBT-LNT* as the production strain is not expected to result in safety concerns.

B. ABSORPTION, DISTRIBUTION, METABOLISM, AND EXCRETION

The ADME of human milk oligosaccharides (HMOs) and other non-digestible carbohydrates, such as galactooligosaccharides, have been extensively summarized in previous GRAS Notices and opinions published by worldwide authoritative bodies, including GRN 923 which summarizes the GRAS status of the subject of this GRAS Determination in infant formula (GRN 484, 2014; GRN 546, 2015; GRN 547, 2014; GRN 571, 2015; GRN 650, 2016; GRN 659, 2016; GRN 735, 2018; GRN 749, 2018; GRN 766, 2018; GRN 815, 2019; GRN 833, 2019; GRN 919, 2020; GRN 921; 2020; GRN 923, 2020; EFSA Panel on Dietetic Products, 2015; EFSA Panel on Nutrition et al., 2020). As summarized on pages 27 and 28 of GRN 923, HMOs, including LNT, are highly resistant to the digestive enzymes of the gastrointestinal (GI) tract and poorly absorbed.

C. TOXICOLOGY STUDIES

The toxicology studies that support the safe use of LNT in foods include a battery of published genotoxicity and subchronic toxicity studies conducted and published by Phipps et al. (2018) and Parschat et al. (2020). All of these studies were extensively summarized in GRN 923 and therefore their summaries are incorporated by reference (see pages 28 – 44 of GRN 923). Briefly, Phipps et al. (2018) evaluated the genotoxicity and subchronic toxicity of a product manufactured by fermentation by Glycom A/S containing 77% LNT in an OECD 408-compliant bacterial reverse mutation test, an OECD 471-compliant in vitro micronucleus test, a 14-day

range-finding oral toxicity test in neonatal rats, and an OECD 408-compliant 90-day oral toxicity test in neonatal rats. Parschat et al. (2020) evaluated the genotoxicity and subchronic toxicity of a mixture of HMOs containing 2'-FL, 3-fucosyllactose (3-FL), LNT, 3'-SL, and 6'-SL manufactured by Chr. Hansen A/S in a corroborative OECD-compliant bacterial reverse mutation assay, an OECD-compliant *in vitro* micronucleus assay, a seven-day pilot dietary toxicity study and an OECD-compliant 90-day feeding study. LNT is not genotoxic and has a no observed adverse effect level (NOAEL) of at least 4000 mg/kg/day (Phipps et al., 2018). Similar results were reported Parschat et al. (2020) for the mixture of HMOs containing 23.79% of Chr. Hansen A/S-manufactured LNT.

As summarized in GRN 923 (pages 28 and 29), the LNT ingredient used by Phipps et al. (2018) and the subject of this GRAS Determination are both manufactured by fermentation using genetically engineered strains of *E. coli* and contain similar amounts of LNT (77 vs 84.7% (average LNT content; see Table 2), respectively). They also have comparable carbohydrate by-products and other impurities controlled by product specifications, such as protein, ash, and moisture. Because Chr. Hansen A/S's LNT ingredient is qualitatively comparable and quantitatively similar to the LNT ingredient manufactured by Glycom A/S and tested by Phipps et al. (2018), the toxicity studies published by Phipps et al. are pivotal to supporting the safe use of Chr. Hansen A/S's LNT ingredient in foods. Thus, based on the results reported by Phipps et al. (2018), adverse effects resulting from the ingestion of Chr. Hansen A/S's LNT per the intended uses and use levels are not expected.

D. TOLERANCE STUDY IN NEONATAL PIGLETS

As summarized in GRN 923, the safety and tolerance of Chr. Hansen A/S's LNT have also been evaluated in the neonatal piglet, which is an appropriate model for understanding the tolerance of food ingredients in infants (Litten-Brown et al., 2010). In a good laboratory practice (GLP)-compliant study, a mixture of HMOs containing 2'-FL, 3-FL, LNT, 3'-SL, and 6'-SL was administered to neonatal piglets for 21 days. Since the filing of GRN 923, this study was published by Hanlon (2020). Because the subject of this GRAS determination was used in the mixture used by Hanlon (2020), the summary in GRN 923 is incorporated by reference (see pages 44 – 76 of GRN 923).

Briefly, thirty-six experimentally naïve domestic two-day-old Yorkshire crossbred piglets were assigned to one of three treatment groups (n=12/group). The treatment groups received either a control diet, a diet containing 5.75 g/L of HMO MIX 1, or a diet containing 8.0 g/L HMO MIX 1. The control diet was Land O'Lakes Specialty Milk Replacer and was used as the base diet for both HMO Mix 1 test diets. HMO MIX 1 was obtained from Chr. Hansen A/S (Rheinbreitbach, Germany) and contained 49.1% 2'-FL, 10.4% 3-FL, 19.9% LNT, 3.5% 3'-SL, and 4.2 % 6'-SL on a dry weight basis. The endpoints that were evaluated included mortality, clinical observations, body weight, feed consumption, feed efficiency, compound consumption,

clinical pathology parameters (hematology, coagulation, clinical chemistry, and urinalysis), gross necropsy findings, organ weights, and histopathologic examinations. Except for one male piglet in the 8.0 g/L dosing group, which was euthanized on day 7 for humane reasons, all of the remaining animals survived until the scheduled study termination on day 22. The clinical and veterinary observations of the male piglet in the 8.0 g/L dosing group that was euthanized included yellow discolored feces, thin body condition, unkempt appearance, generalized muscle wasting, and lateral recumbency. Additionally, *E. coli* was detected in a fecal culture of the one male piglet that was euthanized. Based on the presence of *E. coli* in the feces and the constellation of observations, the unscheduled death/euthanasia of the one male in the 8.0 g/L treatment group was determined to be due to an underlying infection that was distributed evenly between the animals in all dosing groups, not HMO Mix 1-related, and did not affect the validity of the results. The clinical pathology values and macroscopic and microscopic findings in the remaining animals did not reveal a relationship to the HMO Mix 1 treatment and, although increased cecum weights in males and females at ≥ 5.75 g/L, increased colon weights in males at ≥ 5.75 g/L, and decreased rectum weights in males and females at 8.0 g/L were observed, these changes were considered not adverse as there were no microscopic correlates. Together these results indicate that daily dietary administration of HMO Mix 1 to neonatal piglets for 3 weeks at concentrations up to 8.0 g/L with calculated intakes of 3.6 and 3.7 g/kg/bw of HMO Mix 1 (0.72 and 0.74 g LNT/kg bw) in males and females, respectively, was well-tolerated, did not produce adverse effects on growth and development.

E. CLINICAL STUDIES

Additional support for the safe use of LNT in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas at the intended use level is based on results of numerous clinical studies that evaluated the safety and tolerance of HMOs, such as 2'-FL, the LNT isoform lacto-*N*-neotetraose (LNnT), 3'-SL, and 6'-SL, as well as other non-digestible carbohydrates in infants, adults, sensitive populations consuming enteral tube feeding formulas and oral electrolytes solutions. In general, HMOs are well tolerated in infants up to 1 g/day, adults up to 20 g/day, and non-digestible carbohydrates are well tolerated in enteral tube feeding formulas up to 63 g/day and oral electrolyte solutions up to 50 g/L.

1. Clinical Studies with HMOs in Infants and Adults

Lacto-*N*-tetraose is a non-digestible HMO that is GRAS for use in infant formula and selected conventional foods (GRN 833, 2019; GRN 923, 2021). Although no clinical studies have been conducted with LNT specifically, numerous clinical studies have evaluated the tolerability of other HMOs, such as 2'-FL, LNnT, 3'-SL and 6'-SL in infants and adults, Storm et al.(2019), Marriage et al. (2015), Goehring et al. (2016), Puccio et al. (2017), Nowak-Wegrzyn et al. (2019), Kajzer et al. (2016), Alliet et al. (2016), Steenhout et al. (2016), Meli et al. (2014),

Simeoni et al. (2016), Cooper et al. (2016), Radke et al. (2017), Elison et al. (2016), Rasko et al. (2000), Parente et al. (2003), Gurung et al. (2018), Riechmann et al. (2020), Iribarren et al. (2020), Palsson et al. (2020), and Ryan et al. (2021). Except for the studies conducted by Riechmann et al. (2020), Iribarren et al. (2020), Palsson et al. (2020), and Ryan et al. (2021), all of these studies have been extensively summarized in previous GRAS Notifications (GRN 546, 2015; GRN 571, 2015; GRN 571 Supplement, 2019; GRN 650, 2016; GRN 659, 2016; GRN 735, 2018; GRN 749, 2018; GRN 766, 2018; GRN 815, 2019; GRN 852, 2019; GRN 880, 2020; GRN 897, 2020; GRN 919, 2020; GRN 925, 2021). Therefore, their summaries are incorporated by reference and the studies are briefly summarized in tabular format along with the new studies conducted by Riechmann et al. (2020), Iribarren et al. (2020), Palsson et al. (2020), and Ryan et al. (2021) (Table 6 and 7).

In infants, Storm et al. (2019), Marriage et al. (2015), Goehring et al., (2016), Puccio et al. (2017), Nowak-Wegrzym et al. (2019), and Riechmann et al. (2020) administered up to 1.0 g 2'-FL/L and 0.5 g LNnT/L (equivalent to approximately 1.0 g 2'-FL/day and 0.5 g LNnT/day, assuming that infants consume one liter of formula/day) and reported that both HMOs were well-tolerated and had no adverse effect on growth and development (Table 6). Meli et al. (2014), Simeoni et al. (2016), Cooper et al. (2016), and Radke et al. (2017) reported similar effects when they administered a mixture of oligosaccharides containing 3'-SL, galactooligosaccharides and 6'-SL up to a total of 10 g oligosaccharides/L (equivalent to approximately 10 g total oligosaccharides /day, assuming infants consume one liter of formula/day), although the levels of 3'-SL and 6'-SL ingested in the studies were not provided in the publications (Table 6). Importantly, none of the studies reported serious adverse events related to the ingestion of the HMOs and the most common effects were occasional flatulence, abdominal distress, diarrhea, and loose stools, which are not unexpected considering what is known to occur following the ingestion of diets containing high amounts of non-digestible carbohydrates (Eldridge et al., 2019).

In adults, Elison et al. (2016), Iribarren et al. (2020), Palsson et al. (2020), and Ryan et al. (2021) showed that the ingestion of up to 20 g/day of either 2'-FL, LNnT, or a combination of 2'-FL and LNnT in healthy adults and adults with inflammatory bowel disease (IBS), ulcerative colitis, Crohn's disease, or celiac disease was well tolerated, and as expected, the most common complaints were flatulence, abdominal distress, and abdominal pain. Similar results were also reported by Rasko et al. (2000), Parente et al. (2003), and Gurung et al. (2018) when the subjects ingested 20 g 3'-SL/day (Table 7).

Taken together, although no clinical studies have been conducted with LNT, the publicly available studies conducted with 2'-FL, LNnT, 3'-SL, and 6'-SL provide corroborative clinical evidence showing that long-lasting, irreversible adverse effects resulting from the ingestion of HMOs, including LNT, are not expected.

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
<i>2'-Fucosyllactose and/or Lacto-N-neotetraose</i>					
Riechmann et al., 2020	Non-randomized, open-label, prospective study Healthy term infants 7 days to 2 months old	Group 1: Formula-fed infants (n=82) Group 2: Infants consuming formula and human milk; the formula contained 1.0g/L of 2'-FL, 0.5 g LNnT, and Lactobacillus reuteri (n=62) Group 3: Breast-fed infants (n=63)	8 weeks	<ul style="list-style-type: none"> Sixteen subjects dropped out of Group 1 (six were excluded due to protocol deviations, three dropped out due adverse events (AEs), and seven were lost to follow-up). Fourteen subjects dropped out of Group 2 (eight were excluded due to protocol deviations, 3 dropped out due to adverse events, and three were lost to follow-up). Eighteen subjects dropped out of Group 3 (11 were excluded due to protocol deviations, one dropped out due to adverse events, and 6 were lost to follow-up). There were no significant differences between any of the groups for any of the anthropometric measures. Composite Infant Gastrointestinal Symptom Questionnaire (IGSQ) scores demonstrated low gastrointestinal distress in all feeding groups at all time points and there were no significant differences among feeding groups at baseline, 4, or 8 weeks. <ul style="list-style-type: none"> There were no significant differences among the groups in the gassiness, fussiness, crying or spitting-up/vomiting domains of the IGSQ. For the stooling domain, Group 2 were significantly different than Group 3 at baseline and 8 weeks. A total of 49 subjects experienced 58 adverse events over the course of the study. There were 19 AEs in Group 1, 21 in Group 2, and 18 AEs in Group 3. The incidence was generally low and not significantly different among the groups <ul style="list-style-type: none"> Three subjects experienced potentially product-related AEs, including two instances of cow's milk intolerance (one in Group 1 and one in Group 2) and one instance of irritability in Group 1. Six serious adverse events occurred (four in Group 1 and 2 in Group 2), all of which were bronchiolitis. All were considered unrelated to the study feeding. 	Not previously summarized
Nowak-Wegrzyn et al., 2019	Double-blind, placebo-controlled food challenges	Treatment #1: Whey-based extensively hydrolyzed formula Treatment #2: Whey-based extensively hydrolyzed	Not applicable	<ul style="list-style-type: none"> Sixty-four children completed at least one DBPCFC. Three children were excluded due to protocol deviations (n = 61). There was one allergic reaction to the Test, and one to the Control formula. Sixty-one of the 64 subjects completed the open-label home challenge phase with the Test formula 	GRN 919, page 33

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
	Children with cow milk protein allergy	formula containing 1.0 g/L 2'-FL and 0.5 g/L LNnT		<ul style="list-style-type: none"> ○ One subject vomited on Day 1 of the home challenge but completed the home challenge without further problems. ○ One patient developed diarrhea on the last day of the challenge, which the site investigator attributed to gastroenteritis. ○ No significant gastrointestinal symptoms (flatulence, abnormal stool frequency/consistency, increased spitting-up, or vomiting) were reported. ● No serious adverse events occurred during the entire study. 	
Storm et al., 2019	Randomized, placebo-controlled double-blind study Healthy term infants 14 days old ±5 days.	<p>Group 1: Formula containing <i>Bifidobacterium animalis</i> ssp <i>lactis</i> Bb12 (n=40)</p> <p>Group 2: Formula containing <i>Bifidobacterium animalis</i> ssp <i>lactis</i> Bb12 + 0.25 g/L 2'-FL (n=38)</p>	6 weeks	<ul style="list-style-type: none"> ● In the 2'-FL-treated group, one subject was lost to follow-up, one caregiver wished to withdraw, three subjects withdrew due to adverse events (AEs), and three subjects did not comply with feeding only the study formula. ● In the control group, one subject was lost to follow-up, one caregiver wished to withdraw, three subjects withdrew due to adverse events, and two subjects did not comply with feeding only the study formula. ● Infant gastrointestinal symptom questionnaire scores were similar in both groups at baseline and after 6 weeks of treatment. ● Stool frequency and consistency did not differ between the groups over the course of treatment. ● Significantly more stools were reported to be difficult to pass in the control group than in the test group (p<0.05), however, the number of infants with stools reported as difficult to pass was not different between the groups. ● Crying, fussing duration, vomiting frequency, and the proportion of babies reported to have any spit up over the 2-day diary period were similar between the two groups. ● Among the babies whose caregivers reported spit-up, significantly more were reported to have spit up >5 times/day in the 2'-FL group compared to the control group. ● There were no serious AEs and the AEs were equally distributed among the two groups. ● There were significantly more subjects that experienced infections and infestations in the control group than in the 2'-FL-treated group (n=9 vs n=3; p=5). ● There were no effects of the 2'-FL-containing formula on anthropometric measures (body weight and lengths, and weight-for-age and length-for-age). 	GRN 571 supplement, page 21

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
Puccio et al., 2017	Prospective, randomized, placebo-controlled study Healthy, term infants 0 to 14 days old	Group 1: Formula (n=87) Group 2: Formula with 1.0 g/L 2'-FL and 0.5 g/L LNnT (n=88)	6 months (after 6 months, all infants were switched to a non-HMO containing formula)	<ul style="list-style-type: none"> Twenty infants in control and 24 infants in the HMO containing formula withdrew before the primary outcome assessment at 4-months. The dropout rate was comparable between groups. The most common reason for discontinuation was an adverse event (n=11 in control; n=12 in test). Other reasons for discontinuation before 4 months included parent/guardian request (n=3 in control; n=6 in test); lost to follow-up/missing (n=5 in control; n=6 in test); and other (n=1 in control; n=40 in test). There was no difference in weight gain, mean weight-for-age, length-for-age, head circumference-for-age, and BMI-for-age z scores between the groups. Parent-reported infant behavioral patterns including restlessness/irritability and colic were similar in the HMO and control groups except for softer stool (P=0.021) and fewer nighttime wake-ups (P = 0.036) in the test group at 2 months. Infants receiving the HMO-containing formula had significantly fewer parental reports (P = 0.004 – 0.047) of bronchitis through 4 (2.3% vs 12.6%), 6 (6.8% vs 21.8%), and 12 months (10.2% vs 27.6%); lower respiratory tract infection (adverse event cluster) through 12 months (19.3% vs 34.5%); antipyretics use through 4 months (15.9% vs 29.9%); and antibiotics use through 6 (34.1% vs 49.4%) and 12 months (42.0% vs 60.9%) compared to the infants receiving the control formula. 	GRN 650, page 38
Goehring et al., 2016	Prospective, randomized, placebo-controlled study Healthy, term infants 5 days old	Group 1: Formula with GOS (n=39) Group 2: Formula with GOS + 0.2 g/L 2'-FL (n=37) Group 3: Formula with GOS + 1.0 g/L 2'-FL (n=37) Group 4: human milk (HM)(n=42)	16 weeks	<ul style="list-style-type: none"> Note: This is a sub-study of the clinical study conducted by Marriage et al., 2015. The objective was to investigate the effects of feeding formulas supplemented with HMO 2'-FL on biomarkers of immune cell function. Circulating plasma concentrations of inflammatory cytokines IL-1a, IL-1b, IL-6, and TNF-a and anti-inflammatory IL-1ra were significantly higher (82%, 72%, 76%, 58%, and 58%, respectively) in the group fed formula compared to the group receiving human milk (p ≤ 0.05). Both the groups receiving the formulas containing 2'-FL exhibited profiles that were significantly different from the formula group and not different from the human milk group or each other. There were no differences in plasma cytokines IFN-a2, IFN-g, IL-10, IP-10, or RANTES between any of the groups. 	GRN 735, page 62

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
Marriage et al., 2015	Prospective, randomized, placebo-controlled study Healthy, term infants 5 days old	Group 1: Formula with GOS (n=101) Group 2: Formula with GOS + 0.2 g/L 2'-FL (n=104) Group 3: Formula with GOS + 1.0 g/L 2'-FL (n=109) Group 4: human milk (HM)(n=106)	17 weeks	<ul style="list-style-type: none"> • 338 infants completed the study (84 in the control group, 81 in the group receiving the formula containing 0.2 g/L 2'-FL, 83 in the group receiving the formula containing 1.0 g/L 2'-FL, and 90 in the HM group); 304 of whom completed the study on the assigned feeding or HM (79 in the control group, 70 in the group receiving the formula containing 0.2 g/L 2'-FL, 72 in the group receiving the formula containing 1.0 g/L 2'-FL, and 83 in the HM group). The number of premature terminations was not statistically significant among the formula-fed groups. • Although the HM group gained significantly more weight than the group receiving 0.2 g/L 2'-FL from day 14 to 28 and the group receiving 1.0 g/L 2'-FL than the HM group from day 84 to 119, there were no significant differences (sex-specific or sex- combined) in mean weight, length, or head circumference among feeding groups during the study, and no significant differences among feeding groups in mean gains in these measures from day 14 to 119. • The mean number of stools/day was significantly higher for the HM group compared to all groups receiving the formulas for the three days before the study visits at day 28, 42, and 84. The mean number of stools/day was also significantly higher for the HM group compared to the control formula group for the three days before the study visits at day 119. • Although spitting-up or vomiting was significantly higher in the formula-fed groups compared to the HM group from enrollment to day 28, there were no differences after day 28. • Although the mean rank stool consistency was significantly higher for the group receiving 2'-FL from enrollment to day 28 and was significantly higher in the formula-treated groups than the HM group for the remainder of the study, there was no difference among the formula-treated groups over the course of the study. • There were no significant differences in the overall percentage of subjects experiencing adverse events or serious adverse events in the formula-treated groups. • The control formula and the 1 g/L 2'-FL groups had significantly more subjects with reported adverse events in the "infections and infestations" category compared with the 0.2 g/L group (p<0.05), but the types of adverse events were similar (upper respiratory tract symptoms; otitis media, viral infections, and oral candidiasis. The control formula-treated group also had a significantly higher percentage of subjects with eczema (p<0.05) 	GRN 650, page 37

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
Kajzer et al., 2016 (abstract)	Prospective, randomized, double-blind, placebo-controlled study Healthy term infants 0 and 8 days of age.	Group 1: Formula (n=42) Group 2: Formula with 0.2 g/L 2'-FL and 2 g/L scFOS (n=46) Group 3: human milk (HM)(n=43)	5 weeks	<ul style="list-style-type: none"> • Thirty-six (86%) subjects in the group receiving formula, 41 (89%) in the group receiving oligosaccharides and 42 (98%) in the group receiving human milk completed the study. • There was no difference in the mean rank stool consistency among the groups. • The average number of stools per day for the human milk group was significantly higher in the human milk group than both formula-fed groups. • There were no differences among groups for the average volume of study formula intake, number of study formula feedings/day, anthropometric data or percent feeding with spit-up/vomit. • Safety endpoints not reported. 	GRN 571, page 21
Alliet et al., 2016 (abstract)	Randomized, placebo controlled, study Healthy term infants 0-14 days old	Group 1: Cow's milk-based infant formula (n=87) Group 2: Cow's milk-based infant formula w/ 1.0 g/L 2'-FL and 0.5 g/L LNnT (n=88) Group 3: Human milk	3 months	<ul style="list-style-type: none"> • 2'FL and LNnT shift the stool microbiota towards that observed in breastfed infants. • Safety endpoints not reported. 	GRN 815, page 55
Steenhout et al., 2016 (abstract)	Randomized, placebo controlled, study Healthy term infants 0-14 days old	Group 1: Cow's milk-based infant formula (n=87) Group 2: Cow's milk-based infant formula w/ 1.0 g/L 2'-FL and 0.5 g/L LNnT (n=88) Group 3: Human milk	3 months	<ul style="list-style-type: none"> • 2'FL and LNnT shift the stool microbiota towards that observed in breastfed infants. • Safety endpoints not reported. 	GRN 735, page 62

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
<i>3'-Sialyllactose and 6'-Sialyllactose</i>					
Radke et al., 2017	Multicenter, randomized placebo-controlled, double-blind study Healthy term infants 0-14 days old	Group 1: Control formula; (n=207) Group 2: Test formula containing 5.8 ± 1.0 g BMOs*/100 g powdered formula (8 g/L in the reconstituted formula) and 1x10 ⁷ cfu/g <i>B. lactis</i> CNCM I-3446; (n=206) Group 3: Breastfed reference group; (n=63) *BMOs were generated from whey permeate and contained galactooligosaccharides and milk oligosaccharides, such as 3'- and 6'- sialyllactose; the concentrations of 3'- and 6'- sialyllactose are not known	6 months Follow-up at 12 months, no test formula 6-12 months	<ul style="list-style-type: none"> A total of 58 infants (27 in each of the Test and the Control groups and four in the Breast-fed group) were excluded from the ITT analyses because they dropped out before the 1-mo visit. The population that completed the entire study duration was 150 infants in the test formula group, 157 in the control formula group, and 49 in the breastfed group. The proportion of infants with AEs related to infections was comparable among the formula groups. No significant difference in diarrhea or febrile infections incidence among the groups at 6 and 12 months. Test formula was well tolerated and no difference in anthropometric measures were observed among the groups. The test formula group showed similar gut microbiota patterns, fecal IgA, and stool pH to breastfed infants and was significantly different than the control formula group. 	GRN 766, pages 62-64
Simeoni et al., 2016	Randomized, placebo-controlled, double-blind study Healthy 5-day old, term infants	Group 1: Standard formula; (n=37) Group 2: Standard formula plus 5.7±1.0 g/100 g bovine milk oligosaccharides (BMOs*; 8.0 g/L reconstituted formula) and 1x10 ⁷ cfu/g of <i>B. lactis</i> CNCM I-3446; (n=39) Group 3: Human milk; (n=37)	12 weeks	<ul style="list-style-type: none"> No difference in compliance or tolerability was observed among the three groups. <ul style="list-style-type: none"> 10 infants discontinued in the human milk/breastfed group (5 withdrew voluntarily and 5 for other reasons) 7 infants discontinued in the standard formula group (2 withdrew due to GI symptoms, 4 withdrew voluntarily, and 2 were lost to follow-up) 7 infants discontinued in the standard formula with the BMOS and <i>B. lactis</i> CNCM I-3446 group (3 withdrew due to GI symptoms, 2 withdrew voluntarily, and 3 were lost to follow-up) There were no differences in anthropometric measures among the three groups. 	GRN 766, pages 62-64

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
		<p>*BMOs were generated from whey permeate and contained galactooligosaccharides and milk oligosaccharides, such as 3'- and 6'- sialyllactose; the concentrations of 3'- and 6'- sialyllactose are not known</p>		<ul style="list-style-type: none"> • There were no differences in the standard formula and standard formula with BMOS and <i>B. lactis</i> CNCM I-3446 groups in 'spitting up', vomiting, crying, colic, flatulence and irritability. • Infants from the standard formula with BMOS and <i>B. lactis</i> CNCM I-3446 group, but not the standard formula group, showed a proportion of yellowish versus greenish stools equivalent to the breast-fed infants. • Infants in the standard formula with BMOS and <i>B. lactis</i> CNCM I-3446 group showed more liquid stools than infants in the standard formula group; liquid stools were the dominant observation in the breast-fed infants. 	
<p>Cooper et al., 2016</p>	<p>Multicenter, randomized, placebo-controlled, double-blind study</p> <p>Healthy term infants born to HIV+ mothers</p>	<p>Group 1: Cesarean-delivered infants consuming standard formula; (n=101)</p> <p>Group 2: Cesarean-delivered infants and standard formula containing 5.8 ± 1.0 g BMOs*/100 g powder formula (8 g/L in the reconstituted formula) and 1x10⁷ cfu/g <i>B. lactis</i> CNCM I-3446; (n=92)</p> <p>Group 3: Vaginally delivered infants and standard formula; (n=113)</p> <p>Group 4: Vaginally delivered infants standard formula containing 5.8 ± 1.0 g BMOs/100 g powder formula (equivalent to 8 g/L in the reconstituted formula) and 1x10⁷ cfu/g <i>B. lactis</i> CNCM I-3446; (n=115)</p>	<p>4 months</p>	<ul style="list-style-type: none"> • Four hundred and thirty infants were randomized into the study. <ul style="list-style-type: none"> ○ Nine (2.1%) infants were lost to follow-up after randomization but before starting the study formulas. ○ Eight infants were found to be HIV infected, seven at the 4-week visit (v2) and one became positive at 6 months (v5). ○ Of the eight that were HIV infected, three infants died and one discontinued the study. ○ Over the course of the study, there were a total of 55, 57, 47, and 55 discontinuations in the vaginal starter formula containing BMOs and <i>B. lactis</i> CNCM I-3446, vaginal group starter formula, cesarean starter formula containing BMOs and <i>B. lactis</i> CNCM I-3446, and cesarean starter formula groups, respectively. ○ There were no significant differences in tolerability and adverse events between the groups in both delivery methods. • Test formula supplemented with BMOS lowered fecal pH and improved fecal microbiota counts in both delivery methods. 	<p>GRN 766, pages 62-64</p>

Table 6. Clinical Studies with Human Milk Oligosaccharides and Infants

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
		*BMOs were generated from whey permeate and contained galactooligosaccharides and milk oligosaccharides, such as 3'- and 6'- sialyllactose; the concentrations of 3'- and 6'- sialyllactose are not known			
Meli et al., 2014	Randomized, double-blind, single-center study Healthy term infants (<14 days old)	Group 1: Standard formula; (n=84) Group 2: Standard formula plus 10 g bovine milk oligosaccharides (BMOs*/L); (n= 99) Group 3: Standard formula plus 10 g BMOs/L, 2×10^7 cfu/g <i>Bifidobacterium longum</i> ATCC BAA-999 (B1999), and 2×10^7 cfu/g <i>Lactobacillus rhamnosus</i> CGMCC 1.3724 (LPR); (n=98) Group 4: Human milk; (n=39) *BMOs were generated from whey permeate and contained galactooligosaccharides and milk oligosaccharides, such as 3'- and 6'- sialyllactose; the concentrations of 3'- and 6'- sialyllactose are not known	4 months	<ul style="list-style-type: none"> • 90 infants from formula groups and 18 infants from breastfed groups withdrew <ul style="list-style-type: none"> ○ Higher rates of discontinuations were observed in the BMOS-supplemented formula groups (36.4% in Group 2; 34.7% in Group 3) compared with the standard formula-treated group (23.8%), although the differences did not reach statistical significance. ○ GI symptoms (i.e., regurgitation, vomiting, diarrhea, constipation, and abdominal pain characterized by prolonged crying) were the most common reason for study discontinuation in all three formula groups: 14.3% of infants in the standard formula-treated group, 17.2% in Group 2 and 13.3% in the Group 3 discontinued due to GI symptoms. • Weight gain and length and head circumference showed no significant differences between standard and BMOS-containing formula groups • BMOS groups had more frequent and less hard stools compared to control • No significant differences were observed between the standard and BMOS containing formula-treated groups in caregivers' reports of flatulence, vomiting, spitting up, crying, fussing, and colic. 	GRN 766, pages 62-64

Table 7. Clinical Studies with Human Milk Oligosaccharides and Adults					
Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
<i>2'-Fucosyllactose and/or Lacto-N-neotetraose</i>					
Ryan et al., 2021	Open-label, single arm study Adults (21 – 75 years old) with a BMI of 19-40 kg/m ² and with previously diagnosed inflammatory bowel disease (IBS), ulcerative colitis, Crohn's disease, or celiac disease	Group 1: 4 g of 2'-FL in combination with micronutrients, macronutrients, amino acids, and isomalto-oligosaccharide (n=20)	6 weeks	<ul style="list-style-type: none"> • Twelve subjects completed the study. • Eight subjects withdrew from the study <ul style="list-style-type: none"> ○ Two dropped out/declined to participate ○ Three dropped out due to non-serious adverse events. They reported worsening of pre-existing gastrointestinal symptoms, gastrointestinal upset, and a non-study-related viral infection • Three were lost to follow-up. 	Not previously reviewed
Palsson et al., 2020	Open-label, single arm study Adult male and female patients (18 and older) with IBS	Group 1: 5 g of 2'-FL/LNnT (4:1 ratio) (n=317)	12 weeks	<ul style="list-style-type: none"> • Thirteen subjects were discontinued after completing the baseline survey because they did not start the intervention. Therefore, 273 patients completed the study. <ul style="list-style-type: none"> ○ Eight subjects withdrew due to an adverse event. ○ Four subjects withdrew consent. ○ Nineteen subjects were lost to follow-up. • The authors reported that there were no incidents causing safety concerns and the patients generally reported that the intervention was well-tolerated <ul style="list-style-type: none"> ○ Forty-seven patients reported a total of 87 adverse events (AEs) in the study ○ Sixty-one of the AEs were related to the gastrointestinal tract. ○ The most common side effect was passing gas, followed by abdominal distension and pain. • One serious AE occurred (hospitalization due to colitis) but was determined to be unrelated to the intervention by the study's medical safety officer. 	Not previously reviewed

Table 7. Clinical Studies with Human Milk Oligosaccharides and Adults

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
Iribarren et al., 2020	Parallel, double-blind, randomized, placebo-controlled study Adult male and female patients (18 – 64 years old) with inflammatory bowel syndrome (IBS).	Group 1: Placebo (n=21) Group 2: 5 g 2'-FL/LNnT (4:1 ratio) (n=20) Group 3: 10 g 2'-FL/LNnT (4:1 ratio) (n=20)	4 weeks of treatment followed by a 4-week washout	<ul style="list-style-type: none"> • Group 1: one patient discontinued intervention due to worsening of symptoms during the treatment period; one patient was lost to follow-up during the washout period. • Group 2: no patients left the study • Group 3: one patient discontinued intervention due to worsening of symptoms during the treatment period; one patient was lost to follow-up during the washout period. • There were no differences in overall gastrointestinal symptom severity among the groups at week four or week eight. • None of the treatments aggravated the IBS symptoms. • There were no significant differences among the groups in the individual domains of the Gastrointestinal Symptom Rating Scales (abdominal pain, bloating, constipation, diarrhea, and satiety). • Within the groups: <ul style="list-style-type: none"> ○ There was a decrease in the severity of bloating and diarrhea in Group 1 at week 4. ○ In Group 2 and 3, there was a decrease in bloating and abdominal pain at week 8, respectively. • There were no differences between groups or within the groups at week 4 or 8 regarding IBS symptom severity. 	Not previously reviewed

Table 7. Clinical Studies with Human Milk Oligosaccharides and Adults

Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
Elison et al., 2016	Randomized, placebo-controlled double-blind study Healthy male and female adults ages 18 to 60 years.	Group 1: 2g glucose (n=10) Group 2: 5 g 2'-FL (n=10) Group 3: 10 g 2'-FL (n=10) Group 4: 20 g 2'-FL (n=10) Group 5: 5 g LNnT (n=10) Group 6: 10 g LNnT (n=10) Group 7: 20 g LNnT (n=10) Group 8: 3.3 g 2'-FL; 1.7 g LNnT (n=10) Group 9: 6.7 g 2'-FL; 3.4 g LNnT (n=10) Group 10: 13.3 g 2'-FL; 6.7 g LNnT (n=10)	1-2 week run-in period followed by a 2 week treatment period	<ul style="list-style-type: none"> • All subjects were compliant and completed the study according to the protocol without any dropouts. • Fifty-six adverse events were reported by forty-four subjects. <ul style="list-style-type: none"> ○ All were judged as 'mild', and all subjects tolerated the investigational products throughout the trial period. ○ Adverse events were usually reported as a complex of multiple symptoms such as flatulence, bloating and constipation, and were primarily reported at the end of the 2-week intervention. ○ Most adverse events were reported by subjects taking the highest doses of 2'FL and LNnT. Gas/flatulence was the most common adverse event reported, followed by stomach pain, diarrhea/loose stools and rumbling, but at lower frequencies. • No significant difference in bowel movement was observed compared to Group 1. • No change in clinical significance in any physical parameter including pulse rate and blood pressure was found during the 2-week intervention. • There was no difference in clinical chemistry or hematology among the groups at the end of the 2-week intervention period 	GRN 735, page 61

Table 7. Clinical Studies with Human Milk Oligosaccharides and Adults					
Reference	Study Design and Population	Groups (Numbers of Subjects)	Duration	Safety Parameters	GRN Reference
<i>3'-Sialyllactose</i>					
Gurung et al., 2018	Randomized, double-blind, placebo-controlled study Adults with <i>H. pylori</i> infection	Group 1: Placebo (n=17) Group 1: 12 g/day 3'-SL (n=24)	4 weeks	<ul style="list-style-type: none"> There were no significant differences between pre- and post-dose gastrointestinal tolerance and clinical chemistry (serum biochemistry, hematology, and urine analysis) outcomes. Pre- and post-dose urea breath test values were not significantly different within or between the 3'-SL and placebo groups. Compliance and adverse events were similar between the groups. 	GRN 880, pages 35,36
Parente et al., 2003	Randomized, double-blind, placebo-controlled study Adults with <i>H. pylori</i> infection (dyspepsia)	Group 1: Placebo (n=21) Group 2: 10 g/day 3'-SL sodium salt (n=17) Group 3: 20 g/day 3'-SL sodium salt (n=22)	4 weeks	<ul style="list-style-type: none"> Five patients were excluded from analysis due to protocol violation. Adverse events recorded in 6 patients were halitosis, asthenia, epigastric pain, and headache. One patient dropped out due to headache associated with epigastric pain. No serious adverse events were observed. <i>H. pylori</i> colonization documented by the ¹³C-Urea Breath Test (UBT) decreased significantly (<i>p</i>-value not provided) in both treatment groups and placebo but was most likely due to regression toward mean effect. 	GRN 766, pages 64-67
Rasko et al., 2000	Randomized, double-blind, placebo-controlled study Adults with <i>H. pylori</i> infection	Group 1: Placebo (n=6) Group 2: 4g 3'-SL (n=6) Group 3: 8g 3'-SL (n=7) Group 4: 20g 3'-SL (n=7)	56 days for Control and Groups 1 and 2 28 days for Group 3	<ul style="list-style-type: none"> Oral supplementation of 3'-SL did not change Lewis antigen expression of <i>H. pylori</i> strains isolated from human gastric mucosa. No adverse effects on safety or tolerance were reported. 	GRN 766, pages 64-67

2. Clinical Studies with Other Non-digestible Carbohydrates and Enteral Tube Feeding Formulas

Enteral tube feeding is indicated in any patient that has a functioning and accessible gastrointestinal tract and cannot meet their nutritional requirements by consuming food orally (reviewed in Wireko and Bowling, 2010). Enteral tube feeding is administered either as a bolus or continuously via nasogastric tubes, nasojejunal tubes, or gastrostomy and can be associated with issues with the tubes and their insertion, as well as adverse effects in the patient, such as diarrhea, constipation, nausea, and vomiting/aspiration/reflux, bloating, refeeding syndrome and various electrolyte disturbances (<https://gi.org/topics/enteral-and-parenteral-nutrition/>; accessed on February 11, 2021). As a result, enteral tube feeding is generally administered and managed in a medical setting. Importantly, the purpose of using non-digestible carbohydrates in enteral tube feeding formulas is to help alleviate alterations in bowel function and maintain the healthy balance of the microbiota.

Although no clinical studies have been conducted with enteral tube feeding formulas containing LNT, published clinical studies administering other non-digestible, poorly absorbed carbohydrates in enteral tube feeding formulas are relevant to understanding the tolerance of LNT as a non-digestible carbohydrate in enteral tube feeding formulas. As summarized in an amendment to GRN 897 to support the safe use of another HMO, 2'-FL, in enteral formulas, numerous published clinical studies have administered non-digestible carbohydrates, such as partially hydrolyzed guar gum (PHGG), galactomannan, fructooligosaccharides (from short-chain FOS to long-chain inulin), galactooligosaccharides (GOS), and GOS/FOS blends in enteral formulas to infants, children, healthy adults, bed-ridden elderly adults, and patients hospitalized for a variety of serious medical conditions (Akatsu et al., 2016; Alam et al., 2000; Alam et al., 2005; Armanian et al., 2016; Fussell et al., 1996; Garleb et al., 1996; Homann et al., 1994; Homann et al., 2004; Karakan et al., 2007; Khoshoo et al., 2010; Lampe et al., 1992; Meier et al., 1993; Modi et al., 2010; Nakao et al., 2002; Peters and Davidson, 1996; Rushdi et al., 2004; Simakachorn et al., 2011; Spapen et al., 2001; van den Berg et al., 2015; Zheng et al., 2006). Because these studies are extensively summarized in an amendment to GRN 897 their summaries are incorporated by reference and briefly summarized in tabular format below (Table 8). Collectively these studies show that the use of non-digestible carbohydrates in enteral tube feeding formulas at levels up to 63 g/day is well-tolerated.

The Institute of Medicine has also evaluated the potential adverse effects associated with overconsumption of non-digestible carbohydrates such as PHGG, FOS, and GOS, and concluded that although occasional adverse gastrointestinal symptoms can occur (flatulence, abdominal distress, and diarrhea), serious chronic adverse effects have not been observed. Additionally, due to the bulky nature of these substances, excess consumption is likely to be self-limiting and tolerable upper limit (UL) was not established (Eldridge et al., 2019).

Taken together, these data indicate that the risk of adverse effects from the judicious use of non-digestible carbohydrates, such as LNT, in enteral formulas intended for patients with serious medical conditions is generally low and within the GRAS standard of reasonable certainty of no harm.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
<i>Partially Hydrolyzed Guar Gum (PHGG)</i>				
Lampe et al., 1992	Prospective, randomized, placebo-controlled, double-blind, crossover study 11 healthy men	<ol style="list-style-type: none"> 1. Self-selected diet 2. Enteral formula containing no added fiber (maltodextrin) 3. Enteral formula containing 15 g PHGG/day 4. Enteral formula containing 15 g soy polysaccharide 	18 days with a 10 day - washout between each diet period	<ul style="list-style-type: none"> • 12 subjects completed the study; one man did not comply with the diet protocol and his data were excluded from the analyses. No other adverse events were reported. • Compared to the enteral diet with no fiber, fecal wet and dry weights, frequency, stool weight, fecal consistency, fecal moisture, and fecal pH were not statistically different, whereas mean transit time and fecal nitrogen were significantly increased in the PHGG-treated group. • Compared to the enteral diet with no fiber, fecal wet and dry weights, fecal nitrogen, frequency, stool weight, fecal consistency, and fecal pH were not statistically different, whereas mean transit time was significantly decreased and fecal moisture was significantly increased in the soy polysaccharide-treated group. • Colonic fluid acetate, propionate, butyrate and total short chain fatty acids were not significantly different between the PHGG- and no fiber-treated groups • The authors concluded that “despite significant differences in mean transit time, few differences in other parameters of bowel function were observed when healthy subjects consumed enteral formula diets containing 0 g of fiber and 15 g of total dietary fiber as modified guar and soy.”
Meier et al., 1993	Randomized, placebo-controlled crossover study 12 healthy men	<ol style="list-style-type: none"> 1. Standardized normal diet 2. Liquid formula diet 3. Liquid formula diet supplemented with PHGG; intake 42 g PHGG/day 	7 days with a 7 day washout between each diet	<ul style="list-style-type: none"> • Significantly increased colonic but not orocecal transit time compared with either a self-selected diet or the enteral formula without fiber. • PHGG did not affect on stool consistency or frequency.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
Homann et al., 1994	Prospective, randomized, double-blind, placebo-controlled trial 100 hospital patients (30 receiving total enteral nutrition and 70 receiving enteral supplementation)	1. Standard diet 2. Standard diet with 20 g PHGG/L of formula; intake of TPN patients = 24 g PHGG/day; intake of enteral supplementation patients = 20 g PHGG/day	Total enteral nutrition was given for a minimum of 5 days	<ul style="list-style-type: none"> • Patient receiving either total or supplemental enteral nutrition had reduced incidence of diarrhea, but increased flatulence when receiving the standard diet with PHGG compared to those receiving the standard diet alone. • In the patients receiving total enteral nutrition, four patients on the standard total enteral diet, but no patients on the standard diet with PHGG discontinued due to diarrhea. • In the supplemental feeding groups, four patients receiving the standard diet vs. two receiving the standard diet with PHGG discontinued gastrointestinal side effects. • The authors, therefore, reported that: <ul style="list-style-type: none"> ○ The total number of patient with gastrointestinal side effects that resulted in discontinuation of the enteral feeding dropped from eight to two in the standard diet vs the standard diet with PHGG ○ The total number of GI-side effects was not different in the two groups (17 in each group).
Fussell et al., 1996 (Abstract)	Prospective, randomized, double-blind, placebo-controlled study 57 tube-fed adults in 5 diagnostic categories: abdominal surgery/trauma, cerebral trauma, head/neck surgery, multiple fractures, and vascular surgery	1. Fiber free tube feeding formula 2. Fiber free tube feeding formula w/14 g PHGG/L of formula	5-14 days	<ul style="list-style-type: none"> • Forty-four patients completed the protocol. • There was no effect of the fiber on daily diarrhea, nor on albumin, transthyretin, or flatulence. • The PHGG was generally well tolerated.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
Peters and Davidson, 1996	Prospective, randomized, double-blind cross-over study 12 enterally fed patients with Type 1 diabetes	<ol style="list-style-type: none"> 1. Formula containing 29% fat, 55% carbohydrate, and PHGG 2. Formula containing 40% fat, 44% carbohydrate, and PHGG 3. Formula containing 50% fat, 33% carbohydrate, and soy polysaccharide 4. Ensure (53% carbohydrate and no fiber) 	1 day with a week in between treatments	<ul style="list-style-type: none"> • The 2 formulas containing PHGG (concentration not specified) were not effective in attenuating the postprandial glucose response. • No adverse effects were reported.
Spapen et al., 2001	Prospective, randomized, double-blind, placebo-controlled study 25 ICU patients (13 M, 12 F; mean age = 68.5±13.1 years) with severe sepsis and septic shock fed enterally	<ol style="list-style-type: none"> 1. Control formula 2. Formula containing 22 g PHGG/L of formula 	At least 6 days	<ul style="list-style-type: none"> • The group receiving PHGG supplementation exhibited a significantly reduced frequency of diarrhea and a reduction in the number of days with diarrhea • PHGG supplementation had no significant effect on sepsis-related mortality (1 death in the test group, 4 in the control) or duration of stay in the intensive care unit. • The authors concluded: <ul style="list-style-type: none"> ○ “Fiber treatment was well-tolerated” ○ “Total enteral nutrition supplemented with soluble fiber is beneficial in reducing the incidence of diarrhea in tube-fed full-resuscitated and mechanically ventilated septic patients.”
Homann et al., 2004	Prospective, randomized, double-blind, placebo-controlled trial 100 medical and surgical patients (50 patients per group); 30 patients received total enteral nutrition and 70 patients received 1000 ml/day supplemental enteral nutrition	<ol style="list-style-type: none"> 1. Standard diet 2. Standard diet with 20 g PHGG/L of formula; intake of TPN patients = 24 g PHGG/day; intake of enteral supplementation patients = 20 g PHGG/day 	Total enteral nutrition was given for a minimum of 5 days	<ul style="list-style-type: none"> • The PHGG-supplemented formula significantly reduced the number of patients with diarrhea (6 vs. 15 on the fiber-free formula) and the number of days patients suffered from diarrhea (10.2 vs. 40.6 days). • The number of patients experiencing GI side effects was the same in both groups (n = 17 per group), although flatulence was reported in more patients in the PHGG group. • Enteral nutrition was discontinued due to GI side effects in 4 patients on the control/standard diet, but no patients on the PHGG-supplemented diet.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
Rushdi et al., 2004	Prospective, randomized, double-blind, controlled study 30 IBS patients (11 M, 9 F; aged 28-73 years with mean age = 57/5±13/8 years) on enteral nutrition with 3 or more liquid stools/day	1. Standard fiber-free feed 2. Enteral feed enriched with 222 g PHGG/L (22 to 37 g PHGG/day)	4 days	<ul style="list-style-type: none"> • 20 patients completed the protocol (n=10/group); the ten patients that did not complete the protocol because they switched to parenteral nutrition or oral diet, death, or leaving the ICU before completing the study. • Supplementation with PHGG significantly reduced the number of liquid stools. • There were no differences in the incidence or severity of gastrointestinal symptoms between the two groups. • The authors discussed tolerance issues extensively: “Throughout the course of this clinical trial, in the fiber- enriched feed group, only two patients complained of flatulence (20%). On the other hand, in the control group, four patients complained of flatulence (40%), two patients got vomiting (20%) and one case of constipation (10%) was reported. However, no statistical significance was found between both groups as regards incidence or severity of gastrointestinal symptoms. None of these symptoms was severe enough to necessitate therapeutic intervention.”
<i>Galactomannan</i>				
Nakao et al., 2002	Open-label study 20 elderly bed-ridden males and females (10 M, 10 F, mean age = 79.3±5.1 years) receiving enteral feeding	A semi-digested formula containing galactomannan 7 g galactomannan/day during the first week; the dose was increased 7 g/day each week until they received 28 g galactomannan/ day for the fourth week	4 weeks	<ul style="list-style-type: none"> • No adverse effects were reported. • Serum diamine oxidase activity significantly increased following the treatment with the semi-digested formula containing galactomannan. • The water content of the feces decreased, and the frequency of normal stools increased with the semi-digested formula containing galactomannan. • The frequency of bowel movements, the number of aerobic bacteria, and the pH of feces decreased, while fecal SCFA, especially acetic and propionic acids, increased with the semi-digested formula containing galactomannan. • All effects reversed after termination of the galactomannan supplementation. • There was no change in counts of total bacteria or anaerobes and no change in body weight, total serum protein, prealbumin, transferrin, retinol-binding protein, total cholesterol, triacylglycerol, iron, copper, or zinc.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
<i>Fructooligosaccharides</i>				
Karakan et al., 2007	Randomized, double-blind, placebo-controlled study 30 patients aged 46.1±14.0 years with severe acute pancreatitis requiring stoppage of oral feeding for 48 hr	1. Diet 2. Diet containing 0.7 g/soluble fiber and 0.8g/100 g insoluble fiber (24 g/day)	2 days	<ul style="list-style-type: none"> Both enteral feeding solutions were well tolerated with no reported adverse effects. The median duration of enteral feeding and the hospital stay was significantly shorter in the group receiving the fiber-containing diet. The fiber-containing diet also significantly improved the pancreatitis severity scores. The authors concluded that fiber supplementation in severe AP improves hospital stay, duration of nutrition therapy, acute phase response and overall complications compared to standard EN therapy.
Khoshoo et al., 2010	Randomized, double-blind crossover study 14 children aged 1-15 years receiving 75- 100% of calories via feeding tube and were candidates for receiving a peptide-based enteral formula based on documented gastrointestinal dysfunction	1. Formula 2. Formula with 3.5 g FOS/L (approximately 3.5 g FOS/ day)	2 weeks with a 5-day washout period between treatment periods	<ul style="list-style-type: none"> There were nine patients with neurological disorders; 3 patients with inflammatory bowel disease; and 2 patients with short bowel syndrome There were no withdrawals. There was no significant difference in the daily number of bowel movements between children receiving either the fiber or control formulas when evaluating the three diagnoses groups combined or the short bowel syndrome group alone. The children with neurological impairments had more frequent bowel movements when fed the control formula than when fed fiber formula whereas the inflammatory bowel disease group had more daily bowel movements when fed the fiber-containing formula Stools were in the “mushy” category when the participants consumed the fiber containing formula Children with neurological impairment had a significantly lower proportion of stools (P<0.05) characterized as hard nuts and a significantly lower proportion of stools. In the inflammatory bowel disease group, stool frequency was higher with the fiber formula, but there was no change in consistency. There was no difference in the occurrence of vomiting between the two treatments in any of the groups For nine children with a neurological disorder, the mean grade of flatulence/gas was significantly less (P<0.05) when participants consumed the fiber formula whereas there was no difference in flatulence in the other groups.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
				<ul style="list-style-type: none"> • There were no differences in abdominal pain or weight gain among the different groups. • The authors concluded, “This study showed that a peptide-based formula containing fiber was as well-tolerated as a fiber-free formula in a small population of children with gastrointestinal impairments.”
Garleb et al., 1996	Randomized, double-blind, controlled study 27 healthy male college students (n=9/treatment group)	<ol style="list-style-type: none"> 1. Formula 2. Formula with 5 g scFOS/L (approx. 15 g scFOS/day) 3. Formula with 10 g scFOS/L (approx. 30 g scFOS/day) 	14 days	<ul style="list-style-type: none"> • One subject dropped out of the study after one day due to intolerance to the liquid product. The subject was replaced with an alternate. • There were no differences in body weight or deviations from the normal range of blood chemistry values among the three treatment groups. • Although there were no differences in propionate or butyrate, fecal pH, or fecal percent dry matter, fecal acetate, isobutyrate, and isovalerate concentrations were higher among students ingesting scFOS. • Consumption of scFOS also increased fecal bifidobacteria. • Complaints of nausea, cramping, distension, vomiting, diarrhea, and regurgitation were similar across all groups and were present on fewer than 5% of participant-days. • Flatus was reported more frequently by those consuming 30 g scFOS/day, but most complaints occurred during the first 4 days. • The authors concluded that “these results indicate that scFOS does not compromise serum chemistry profiles, is well tolerated particularly at an intake of 15 g/d and would serve as a bifidogenic factor when incorporated into a liquid enteral product.”
Simakachorn et al., 2011	Randomized, double-blind, placebo-controlled study 94 critically ill children age 1-3 years under mechanical ventilation and enteral feeding (n=47/groups)	<ol style="list-style-type: none"> 1. Control formula 2. Test formula with 2.6 g/L of oligo-fructose/inulin and 2.8 g/L of acacia gum in combination with 2 strains of live microorganisms 	7 days of enteral feeding followed by 14 days of oral feeding	<ul style="list-style-type: none"> • 6 children withdrew from the test formula group; 8 children withdrew from the control formula group. One child withdrew consent in the test formula group, 5 children withdrew consent in the control formula group. • One child was lost to follow-up in the test formula group (moved to another hospital) and one child was lost to follow-up in the control formula group (no reason given). Four children discontinued the intervention in the test formula group due to death whereas two children discontinued the intervention in the control formula group due to death.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
				<ul style="list-style-type: none"> • There were no significant differences in adverse events between the two groups and no reported secondary infections during the ICU stay. • Abdominal distension, vomiting, and stool frequency were also unaffected by the fiber. • The authors concluded that the experimental enteral formula is safe and well-tolerated by children in intensive care receiving enteral nutrition.
Majid et al., 2014	Randomized, double-blind, placebo-controlled study 47 adults in the intensive care unit	<ol style="list-style-type: none"> 1. Control formula containing soy polysaccharides, resistant starch, Arabic gum, cellulose, inulin, and oligofructose (0.7 g/100 ml soluble fiber and 0.8 g/100 ml insoluble fiber, equivalent to 6.75 g/day)); n=23 2. Formula containing soy polysaccharides, resistant starch, Arabic gum, cellulose, inulin, and oligofructose (0.7 g/100 ml soluble fiber and 0.8 g/100 ml insoluble fiber; equivalent to 6.75 g/day) with and additional 7 g oligofructose/inulin; n=24 	A minimum of 3 days	<ul style="list-style-type: none"> • 12 patients discontinued the study before the intervention (7 in the placebo group and 5 in the oligofructose/inulin group) • 6 patients discontinued the intervention in the control formula group (1 patient transferred to an oral diet and five transferred to palliative care) vs 7 patients discontinued in the oligofructose/inulin group (5 transferred to palliative care and 2 were discharged to another hospital) • There was no significant difference in short-chain fatty acid concentrations at baseline or follow-up between the two groups. • Fecal pH was similar in the two groups at baseline and at follow-up. • There were no significant differences in fecal frequency or the daily fecal score between the two groups. • There was no difference between the two groups in the mean number of days of diarrhea or in the number of patients experiencing diarrhea on either one or two or more consecutive days.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
<i>Galactooligosaccharides or GOS/FOS</i>				
Modi et al., 2010	Prospective, randomized, double-blind, placebo-controlled, multi-center study 160 preterm infants (gestational age <33 weeks) receiving enteral feeding	1. Standard formula 2. Test formula with 8 g/L of scGOS/lc FOS in a 9:1 ratio	~8 weeks or until discharge	<ul style="list-style-type: none"> 83 infants received the standard formula; 77 infants received the test formula containing GOS/FOS. The parents of two and four infants withdrew consent in the standard and test formula groups, respectively. One infant in the standard formula group died before reaching the primary outcome and two infants in the test formula group died before reaching the primary outcome. One infant in the standard formula treated group was discharged before reaching the primary outcome. Six adverse events were reported by one infant, five of which were not considered related to the trial. There were three cases of necrotizing enterocolitis (one in the standard formula group vs 2 in the test formula group). Nineteen infants develop at least one episode of a blood stream infection (10 in the standard formula group vs 9 in the test formula group). There was no overall difference in tolerance between control and test formula, but the addition of scGOS/lc FOS to formula improved tolerance for the most immature infants. There were no differences in gains in weight, length, or head circumference; in stooling frequency, stool characteristics, or fecal microbiota; or in GI signs or water balance (based on concentrations of serum sodium and creatinine). The authors concluded that scGOS/lc FOS supplementation is safe.
Akatsu et al., 2016	Prospective, randomized, double-blind, placebo-controlled study 36 elderly individuals	1. Oral feeding (n=13) 2. Enteral formula (n=11) Enteral formula w/ GOS and bifidogenic growth stimulator (BGS; 2-amino-3-carboxy-1,4-naphtho-quinone) (n=12) Products were delivered via percutaneous endoscopic gastrostomy	10 weeks	<ul style="list-style-type: none"> No adverse effects were reported.

Table 8. Clinical Studies of Non-digestible Carbohydrates Administered Via Enteral Feeding¹				
Citation	Study Design	Treatments	Duration	Safety-Related Findings
Armanian et al., 2016	Prospective, randomized, double-blind, placebo-controlled study 25 hyper-bilirubinemic preterm neonates who had reached 30 ml/kg bw/day enteral feeding volume	1. Distilled water 2. A supplement containing scGOS/lc FOS in a 9:1 ratio *The supplement was initially administered by 0.5 g/kg/day and then increased to 1 g/kg/day and 1.5 g/kg/day	1 week	<ul style="list-style-type: none"> No adverse effects were reported. Stool frequency was significantly increased in the scGOS/lc FOS-treated group. The authors concluded that oligosaccharides increase stool frequency, improve feeding tolerance and reduce bilirubin level in preterm neonates and therefore can be efficacious for the management of neonatal hyperbilirubinemia.
Van den Berg et al., 2015	Prospective, randomized, double-blind, placebo-controlled study to determine the effect of combined short-chain galacto-oligosaccharides (scGOS), long-chain fructo-oligosaccharides (lcFOS) and pectin-derived acidic oligosaccharides (pAOS) on antibody concentrations after pneumococcal conjugate vaccination in very preterm infants. 113 infants with a gestational age of <32 weeks or birth-weight <1500 g	1. Placebo/maltodextrin (n=58) 2. scGOS/lc FOS/ pectin-derived acidic oligosaccharides(pAOS) (n=55)	4 weeks	<ul style="list-style-type: none"> Nine infants died in the placebo-treated group whereas six infants died in the scGOS/lc FOS/pAOS-treated group. Adverse events were not reported. The authors concluded “Short-term supplementation of scGOS/lcFOS/pAOS during day 3–30 of life decreased the pneumococcal vaccine antibody response after the primary series of PCV7 at 5 months in preterm infants to levels which are similar in term infants from a Dutch population study. However, after the booster vaccination at 12 months, this effect of the scGOS/lcFOS/pAOS on the PCV response had disappeared.”

¹Incorporated by reference from the amendment to GRN 897.

3. Clinical Studies with Other Non-digestible Carbohydrates and Oral Electrolyte Solutions

a. Background

Oral electrolyte solutions (OESs) are liquid products that facilitate rapid and effective rehydration. OESs contain, at a minimum, a digestible carbohydrate such as dextrose and sodium in water to facilitate water absorption from the lumen of the gastrointestinal tract. Specifically, dextrose absorption facilitates sodium ion absorption, which thereby raises the concentration of sodium ions in the blood stream, pulling water from the lumen of the gastrointestinal tract into the blood stream. Importantly, this is all accomplished through a balance between the amount of carbohydrate and the electrolytes in the OES. Additionally, although sodium absorption improves as the dextrose concentration of the oral fluid is increased up to about 2.5% w/w, higher concentrations of dextrose can increase the osmotic load in the gut, pulling water out of the blood stream, further exacerbating dehydration. Simple sugars such as dextrose and fructose have also been shown to be more effective than larger, more complex carbohydrates in facilitating electrolyte absorption and many oligosaccharides are not stable in acidic mediums such as OESs. As a result, conventional OESs generally do not include oligosaccharides or polysaccharides (Patent 10,695,358, date issued June 30, 2020 Abbott Laboratories).

Importantly, non-digestible carbohydrates, such as LNT, GOS, FOS and LNnT stimulate the growth or activity, or both, of Bifidobacterium in the gastrointestinal tract (reviewed in Gibson and Roberfroid, 1995). Non-digestible carbohydrates are also fermented by the colonic bacteria to short-chain fatty acids (SCFA), which are rapidly absorbed in the colon and further stimulating fluid and sodium absorption (reviewed in Binder et al., 2014). Thus, OESs supplemented with non-digestible carbohydrates, such as LNT, may facilitate rehydration, as well as maintenance of the microbiota.

b. Use of Non-Digestible Carbohydrates in Acute Diarrhea and As an Ingredient in Oral Electrolyte Solutions

The safety and tolerance of numerous non-absorbable carbohydrates (GOS, FOS, xylooligosaccharides (XOS)) have been extensively reviewed and been the subject of numerous GRAS Notices (GRNs 44, 172, 233, 236, 246, 285, 286, 334, 343, 370, 458, 484, 495, 518, 537, 569, 605, 620, 623, 671, 674, 717, 721, 729, 779, 797, 816, 818, 896); human milk oligosaccharides have also been extensively reviewed and the subjects of numerous GRAS Notices (2'-FL: GRNs 546, 571, 650,735, 749, 815, 852, 859, 897; 3-FL: GRN 925; 3'-SL and 6'-SL: GRNs 766, 880, 881, 921, 922; LNT: GRN 923; LNnT: GRNs 919, 895).

During diarrhea, pathogenic bacteria may either grow and colonize the gastrointestinal (GI) tract and then invade the host tissues or, alternatively, they may secrete toxins which may disrupt the function of the intestinal mucosa, causing nausea, vomiting, and diarrhea. Oli et al., (1998) showed that in a pig model, adding fructo-oligosaccharides (FOS) to an OES accelerated the recovery of lactobacilli and reduced bacterial counts of Enterobacteriaceae. Brunser et al. (2006) studied the effect of FOS on the intestinal microbiota during treatment with amoxicillin and reported an increase in bifidobacteria in patients receiving FOSs after seven days of antibiotic treatment compared to a control group. These authors reported that the effect of FOS on the occurrence of antibiotic-related diarrhea episodes was not significant. Vaisman et al. (2010) investigated the effect of a mixture of long-chain FOS, GOS, and acidic oligosaccharides on the number and consistency of stools and on immune system biomarkers in 104 supplemented and non-supplemented subjects (aged 9–24 months) with acute diarrhea. No treatment-related adverse effects were reported. Additionally, studies of OESs supplemented with non-digestible carbohydrates and/or sources of non-digestible carbohydrates, such as guar gum, FOS, XOS, and high amylose maize starch, indicate that non-digestible carbohydrates do not exacerbate acute diarrhea (Table 9; Alam et al., 2015; Passariello et al., 2011; Vandenplas et al., 2011; Raghupathy et al., 2006; Hoekstra et al., 2004; Alam et al., 2000). Therefore, based on the weight of the evidence, adverse effects resulting from the addition of LNT to OESs are not expected.

c. Lack of Impact of LNT on Osmolarity

The WHO current standard OES osmolarity is 245 mOsm/L; Pedialyte® from Abbott is 250 mOsm/L (Ofei et al., 2019). Despite common perceptions that sport drinks can be used for dehydration, liquid products such as sports beverages and juices are hyperosmolar (330–730 mOsm/L) and inappropriate as rehydration solutions for diarrhea and dehydration because they increase fluid losses and worsen the diarrheal disease. It is critical that the addition of any ingredient to an OES not impact the osmolarity. The addition of 0.6 g/L of LNT to OES, such as Pedialyte®, is calculated on the basis of molar weight to add 0.85 mOsm/L. Thus, the addition of 0.6 g/L of LNT will not impact the osmolarity of the solution.

Table 9. Studies of Oral Electrolyte Solutions (OES) with Added Nondigestible Carbohydrate			
Reference	Trial Design	Test Article	Results
Alam et al., 2015	Randomized, double-blind placebo controlled clinical trial of 126 malnourished children (male and female) (weight for length/weight for age <3 Z-score with or without pedal edema), aged 6-36 months with acute diarrhea	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) Group 2: Standard hypotonic ORS with 15 g/L partially hydrolyzed guar gum 	<ul style="list-style-type: none"> The mean duration of diarrhea was significantly shorter in children in Group 2 compared to Group 1. Adverse events/tolerance related to test article not reported by authors.
Passariello et al., 2011	Single-blind, prospective, controlled trial including children (age range, 3-36 months) with acute diarrhea	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) Group 2: hypotonic ORS with zinc, 0.35 g/L fructooligosaccharides and 0.35 g/L xylooligosaccharides 	<ul style="list-style-type: none"> Resolution of diarrhea at 72 hours, number of daily outputs at 24, 48, and 72 hours was statistically significantly improved in Group 2 compared to Group 1. Total ORS intake in the first 24 hours of rehydration therapy was statistically significantly lower in Group 1 than Group 2. No adverse events related to the use of the ORS were observed in the study groups.
Vandenplas et al., 2011	Randomized, prospective, double-blind placebo-controlled trial in children between 3 and 186 months (males and females) with acute diarrhea	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) Group 2: Standard hypotonic ORS with a symbiotic blend (<i>Streptococcus thermophilus</i>, <i>Lactobacillus rhamnosus</i>, <i>Lactobacillus acidophilus</i>, <i>Bifidobacterium lactis</i>, <i>Bifidobacterium infantis</i>, fructo-oligosaccharides). 	<ul style="list-style-type: none"> Children in Group 2 had significantly reduced duration of diarrhea compared with Group 1. Adverse events/tolerance related to test article not reported by authors.
Raghupathy et al., 2006	Randomized, double-blind, placebo-controlled study including boys aged 6 months to 3 years with acute diarrhea with clinically detectable dehydration	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) (311 mOsm/kg) Group 2: Standard hypotonic ORS with 50 g/L high-amylose maize starch 	<ul style="list-style-type: none"> Statistically significant shortened duration of diarrhea in Group 2 compared to Group 1. Before the start of this study high-amylose maize starch, ORS was administered orally to 6 children with acute diarrhea and found to be well tolerated. It did not induce vomiting or significant increase in diarrhea.

Table 9. Studies of Oral Electrolyte Solutions (OES) with Added Nondigestible Carbohydrate			
Reference	Trial Design	Test Article	Results
Hoekstra et al., 2004	Randomized, double-blind, placebo-controlled multicenter study including boys aged 1 to 36 months with acute diarrhea	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) Group 2: Standard hypotonic ORS with a mixture of non-digestible carbohydrates (soy polysaccharide 25%, alpha-cellulose 9%, gum arabic 19%, fructooligosaccharides 18.5%, inulin 21.5%, resistant starch 7%) 	<ul style="list-style-type: none"> No significant differences in mean 48 hours stool volume or duration of diarrhea in Group 2 compared to Group 1. No significant adverse effects, as compared to ORS with placebo, were noted.
Alam et al., 2000	Double-blind, randomized, placebo controlled clinical trial of 150 male children aged 4 to 18 months who had acute diarrhea	<ul style="list-style-type: none"> Group 1: Standard hypotonic oral rehydration solution (ORS) Group 2: Standard hypotonic ORS with 15 g/L partially hydrolyzed guar gum 	<ul style="list-style-type: none"> Children in Group 2 had significantly reduced duration of diarrhea compared with Group 1. Adverse events/tolerance related to test article not reported by authors.

F. ALLERGENICITY

The allergenicity of Chr. Hansen A/S's LNT ingredient was extensively reviewed in GRN 923. Therefore, the allergenicity summary in GRN 923 is incorporated by reference (see page 77 of GRN 923). Allergic reactions resulting from the exposure to Chr. Hansen A/S's LNT product are not expected based on the following:

- LNT is a component of human milk;
- Allergic reactions to HMOs have not been reported;
- Genetically engineered strains of *E. coli* BL21(DE3) are safely used in the production of food and pharmaceutical ingredients;
- Cross-reactivity of the engineered proteins in *JBT-LNT* with known allergens is not expected based on the results of FASTA amino acid alignments with the AllergenOnline Database maintained by the University of Nebraska – Lincoln;
- The protein content of Chr. Hansen A/S's LNT is controlled with a specification of ≤ 0.01 % protein.

G. REGULATORY APPROVALS AROUND THE WORLD

Lacto-*N*-tetraose is GRAS in the United States for use in cow's milk-based term infant formula at levels up to 0.8 g/L and selected conventional foods and beverages at levels ranging from 1 to 20 g/kg (GRN 833, 2019). LNT is also a Novel Food in the European Union for use in infant and follow-on formulas at 0.8 and 0.6 g/L, respectively, and in selected conventional foods up to 10 g/kg (Commission Implementing Regulation (EU) 2020/484). Following their review of the LNT Novel Food application submitted by Glycom A/S, the European Food Safety Authority (EFSA) agreed that the NOAEL for Glycom's LNT is 4,000 mg/kg bw/day, which was the highest dose tested in the 90-day oral toxicity study conducted by Phipps et al. (2018). EFSA opined that the intake of LNT at the proposed use levels is unlikely to exceed the intake level of naturally occurring LNT in breastfed infants on a body weight basis and that the intake of other carbohydrates structurally related to LNT is not a safety concern. Moreover, they concluded that LNT is safe for use in infant and follow-on formulas at 0.8 and 0.6 g/L, respectively, and in selected conventional foods up to 10 g/kg (EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA) et al., 2019).

VII. SUPPORTING DATA AND INFORMATION

A. REFERENCES

All information included in the following list of references is generally available.

Akatsu, H., Nagafuchi, S., Kurihara, R., Okuda, K., Kanetsaka, T., Ogawa, N., Kanematsu, T., Takasugi, S., Yamaji, T., Takami, M., Yamamoto, T., Ohara, H., and Maruyama, M. (2016). Enhanced vaccination effect against influenza by prebiotics in elderly patients receiving enteral nutrition. *Geriatr Gerontol Int* 16, 205–213.

Alam NH, Ashraf H, Kamruzzaman M, et al. (2015). Efficacy of partially hydrolyzed guar gum (PHGG) supplemented modified oral rehydration solution in the treatment of severely malnourished children with watery diarrhoea: a randomised double-blind controlled trial. *J Health Popul Nutr* 34, 3.

Alam, N.H., Meier, R., Sarker, S.A., Bardhan, P.K., Schneider, H., and Gyr, N. (2005). Partially hydrolysed guar gum supplemented comminuted chicken diet in persistent diarrhoea: a randomised controlled trial. *Arch Dis Child* 90, 195–199.

Alam NH, Meier R, Schneider H, et al. (2000). Partially hydrolyzed guar gum-supplemented oral rehydration solution in the treatment of acute diarrhea in children. *J Pediatr Gastroenterol Nutr* 31, 503-507.

Alliet, P., Puccio, G., Janssens, E., Cajozzo, C., Corsello, G., Berger, B., Sperisen, P., Martin, F.-P., Sprenger, N., and Steenhout, P. (2016). Term infant formula supplemented with human milk oligosaccharides (2' fucosyllactose and lacto-neotetraose) shifts stool microbiota and metabolic signatures closer to that of breastfed infants. *Journal of Pediatric Gastroenterology and Nutrition* 63, S55.

Armanian, A.M., Barekatin, B., Hoseinzadeh, M., and Salehimehr, N. (2016). Prebiotics for the management of hyperbilirubinemia in preterm neonates. *J Matern Fetal Neonatal Med* 29, 3009–3013.

Asakuma, S., Hatakeyama, E., Urashima, T., Yoshida, E., Katayama, T., Yamamoto, K., Kumagai, H., Ashida, H., Hirose, J., and Kitaoka, M. (2011). Physiology of consumption of human milk oligosaccharides by infant gut-associated bifidobacteria. *J Biol Chem* 286, 34583–34592.

Asakuma, S., Urashima, T., Akahori, M., Obayashi, H., Nakamura, T., Kimura, K., Watanabe, Y., Arai, I., and Sanai, Y. (2008). Variation of major neutral oligosaccharides levels in human colostrum. *European Journal of Clinical Nutrition* 62, 488–494.

Bao, Y., Chen, C., and Newburg, D.S. (2013). Quantification of neutral human milk oligosaccharides by graphitic carbon high-performance liquid chromatography with tandem mass spectrometry. *Analytical Biochemistry* 433, 28–35.

Binder HJ, Brown I, Ramakrishna BS, Young GP. (2014). Oral rehydration therapy in the second decade of the twenty-first century. *Curr Gastroenterol Rep* 16, 376.

Brunser O, Gotteland M, Cruchet S, Figueroa G, Garrido D, Steenhout P. (2006). Effect of a milk formula with prebiotics on the intestinal microbiota of infants after an antibiotic treatment. *Pediatr Res* 59, 451-456.

CDC (Centers for Disease Control). (2006). Analytical and Reporting Guidelines: The National Health and Nutrition Examination Survey (NHANES). National Center for Health Statistics, Centers for Disease Control and Prevention; Hyattsville, Maryland. Available from: http://www.cdc.gov/nchs/data/nhanes/nhanes_03_04/nhanes_analytic_guidelines_dec_2005.pdf

CDC (2018). NHANES 2015-2016 Dietary Data. Available from: <https://www.cdc.gov/nchs/nhanes/search/datapage.aspx?Component=Dietary&CycleBeginYear=2015>

Chaturvedi, P., Warren, C.D., Altaye, M., Morrow, A.L., Ruiz-Palacios, G., Pickering, L.K., and Newburg, D.S. (2001). Fucosylated human milk oligosaccharides vary between individuals and over the course of lactation. *Glycobiology* 11, 365–372.

Chaturvedi, P., Warren, C.D., Ruiz-Palacios, G.M., Pickering, L.K., and Newburg, D.S. (1997). Milk oligosaccharide profiles by reversed-phase HPLC of their perbenzoylated derivatives. *Anal Biochem* 251, 89–97.

Commission Implementing Regulation (EU) 2020/484 of 2 April 2020 authorising the placing on the market of lacto-*N*-tetraose as a novel food under Regulation (EU) 2015/2283 of the European Parliament and of the Council and amending Commission Implementing Regulation (EU) 2017/2470.

Cooper, P., Bolton, K.D., Velaphi, S., de Groot, N., Emady-Azar, S., Pecquet, S., and Steenhout, P. (2016). Early Benefits of a Starter Formula Enriched in Prebiotics and Probiotics on the Gut Microbiota of Healthy Infants Born to HIV+ Mothers: A Randomized Double-Blind Controlled Trial. *Clin Med Insights Pediatr* 10, 119–130.

Coppa, G.V., Pierani, P., Zampini, L., Carloni, I., Carlucci, A., and Gabrielli, O. (1999). Oligosaccharides in human milk during different phases of lactation. *Acta Paediatr Suppl* 88, 89–94.

EFSA Panel on Nutrition, N.F.A.F.A.N.D.A., Turck, D., Castenmiller, J., De Henauw, S., Hirsch-Ernst, K.I., Kearney, J., Maciuk, A., Mangelsdorf, I., McArdle, H.J., Naska, A., Pelaez, C., Pentieva, K., Siani, A., Thies, F., Tsabouri, S., Vinceti, M., Cubadda, F., Engel, K.H., Frenzel, T., Heinonen, M., Marchelli, R., Neuhäuser-Berthold, M., Poulsen, M., Schlatter, J.R., van

Loveren, H., Colombo, P., and Knutsen, H.K. (2020). Safety of 3'-Sialyllactose (3'-SL) sodium salt as a novel food pursuant to Regulation (EU) 2015/2283. *EFSA Journal* 18.

EFSA Panel on Nutrition, Novel Foods and Food Allergens (NDA), Turck, D., Castenmiller, J., De Henauw, S., Hirsch-Ernst, K.I., Kearney, J., Maciuk, A., Mangelsdorf, I., McArdle, H.J., Naska, A., Pelaez, C., Pentieva, K., Siani, A., Thies, F., Tsabouri, S., Vinceti, M., Cubadda, F., Engel, K.H., Frenzel, T., Heinonen, M., Marchelli, R., Neuhäuser-Berthold, M., Poulsen, M., Sanz, Y., Schlatter, J.R., van Loveren, H., Colombo, P., and Knutsen, H.K. (2019). Safety of lacto-*N*-tetraose (LNT) as a novel food pursuant to Regulation (EU) 2015/2283. *EFSA Journal* 17.

EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA). Safety of lacto-*N*-neotetraose as a novel food ingredient pursuant to Regulation (EC) No 258/97. *EFSA Journal* 2015;13(7):4183.

Eldridge, A.L., Catellier, D.J., Hampton, J.C., Dwyer, J.T., and Bailey, R.L. (2019). Trends in Mean Nutrient Intakes of US Infants, Toddlers, and Young Children from 3 Feeding Infants and Toddlers Studies (FITS). *J Nutr* 149, 1230–1237.

Elison, E., Vigsnaes, L.K., Rindom Krogsgaard, L., Rasmussen, J., Sørensen, N., McConnell, B., Hennem, T., Sommer, M.O., and Bytzer, P. (2016). Oral supplementation of healthy adults with 2'-*O*-fucosyllactose and lacto-*N*-neotetraose is well tolerated and shifts the intestinal microbiota. *Br J Nutr* 116, 1356–1368.

Erney, R.M., Malone, W.T., Skelding, M.B., Marcon, A.A., Kleman-Leyer, K.M., O'Ryan, M.L., Ruiz-Palacios, G., Hilty, M.D., Pickering, L.K., and Prieto, P.A. (2000). Variability of human milk neutral oligosaccharides in a diverse population. *J Pediatr Gastroenterol Nutr* 30, 181–192.

Fussell, S.T., Garmhausen, L., and Koruda, M.J. (1996). The influence of guar gum on diarrhea in critically-ill tube-fed patients. *JPEN J Parenter Enteral Nutr* 20, 26S.

Gabrielli, O., Zampini, L., Galeazzi, T., Padella, L., Santoro, L., Peila, C., Giuliani, F., Bertino, E., Fabris, C., and Coppa, G.V. (2011). Preterm milk oligosaccharides during the first month of lactation. *Pediatrics* 128, e1520–31.

Galeotti, F., Coppa, G.V., Zampini, L., Maccari, F., Galeazzi, T., Padella, L., Santoro, L., Gabrielli, O., and Volpi, N. (2012). On-line high-performance liquid chromatography-fluorescence detection-electrospray ionization-mass spectrometry profiling of human milk oligosaccharides derivatized with 2-aminoacridone. *Anal Biochem* 430, 97–104.

Garleb, K.A., Snook, J.T., Marcon, M.J., Wolf, B.W., and Johnson, W.A. (1996). Effect of fructooligosaccharide containing enteral formulas on subjective tolerance factors, serum chemistry profiles, and faecal bifidobacteria in healthy adult male subjects. *Microbial Ecology in Health and Disease* 9, 279–285.

Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *J Nutr.* 1995 Jun;125(6):1401-12. doi: 10.1093/jn/125.6.1401. PMID: 7782892.

Goehring, K.C., Marriage, B.J., Oliver, J.S., Wilder, J.A., Barrett, E.G., and Buck, R.H. (2016). Similar to Those Who Are Breastfed, Infants Fed a Formula Containing 2'-Fucosyllactose Have Lower Inflammatory Cytokines in a Randomized Controlled Trial. *The Journal of Nutrition* 146, 2559–2566.

GRN 44 (2000). Fructooligosaccharide. GTC Nutrition Company.

GRN 172 (2005). Isomalto-oligosaccharide. BioNeutra, Inc.

GRN 233 (2009). Combination of galacto-oligosaccharides and polydextrose. Mead Johnson & Company.

GRN 236 (2008). Galacto-oligosaccharides. Friesland Foods Domo.

GRN 246 (2008). Isomalto-oligosaccharide mixture. BioNeutra, Inc.

GRN 285 (2009). Galacto-oligosaccharides. GTC Nutrition.

GRN 286 (2009). Galacto-oligosaccharides. GTC Nutrition.

GRN 334 (2010). Galacto-oligosaccharides. Yakult Pharmaceutical Industry Co., Ltd.

GRN 343 (2010). Wheat bran extract composed primarily of xylo- and arabinoxylo-oligosaccharides. Fugeia NV.

GRN 370 (2011). Xylooligosaccharide. Shangdong Longlive Bio-technology Co., Ltd.

GRN 458 (2014). Xylooligosaccharide. Shangdong Longlive Bio-technology Co., Ltd.

GRN 484 (2014). Galacto-oligosaccharides. Clasado, Inc.

GRN 495 (2014). Galacto-oligosaccharides. Clasado, Inc.

GRN 518 (2014). New Francisco Biotechnology Corporation. New Francisco Biotechnology Corporation.

GRN 537 (2014). Short-chain fructo-oligosaccharides. Ingredion Incorporated.

GRN 546 (2015). 2'-Fucosyllactose. Glycom A/S.

GRN 547 (2015). Lacto-*N*-neotetraose. Glycom A/S.

GRN 569 (2015). Galacto-oligosaccharides. New Francisco Biotechnology Corporation.

GRN 571 Supplement, (2019). 2'-Fucosyllactose. Jennewein Biotechnologie, GmbH.

GRN 571 (2015). 2'-Fucosyllactose. Jennewein Biotechnologie, GmbH.

- GRN 605 (2016). Fructo-oligosaccharides. Tata Chemicals Limited.
- GRN 620 (2016). Galacto-oligosaccharides. Nestle Nutrition.
- GRN 623 (2016). Fructooligosaccharides. Hank Tsai.
- GRN 650 (2016). 2'-Fucosyllactose. Glycom A/S.
- GRN 659 (2016). Lacto-*N*-neotetraose. Glycom A/S.
- GRN 671 (2017). Galacto-oligosaccharides (GOS). Vitalus Nutrition, Inc.
- GRN 674 (2017). Isomalto-oligosaccharides. BioNeutra North America Inc.
- GRN 717 (2018). Short-chain fructo-oligosaccharides. Galam, Ltd.
- GRN 721 (2017). Galacto-oligosaccharides. Vitalus Nutrition Inc.
- GRN 729 (2018). Galacto-oligosaccharides. Neo Cremar, Co., Ltd.
- GRN 735 (2018). 2'-Fucosyllactose. Glycosyn and FrieslandCampina.
- GRN 749 (2018). 2'-Fucosyllactose. Dupont Nutrition and Health.
- GRN 766 (2018). 3'-Sialyllactose. GeneChem, Inc.
- GRN 779 (2018). Isomalto-oligosaccharide mixture. Baolingbao Biology Co., Ltd.
- GRN 797 (2018). Fructooligosaccharides. New Francisco (Yunfu City) Biotechnology Corporation.
- GRN 815 (2019). 2'-Fucosyllactose and Difucosyllactose. Glycom A/S.
- GRN 816 (2019). Xylooligosaccharides from sugarcane. Prenexus Health, Inc.
- GRN 818 (2019). Isomalto-oligosaccharides. Ingredion Inc.
- GRN 833 (2019). Lacto-*N*-tetraose. Glycom A/S.
- GRN 852 (2019). 2'-Fucosyllactose. BASF SE.
- GRN 859 (2019). 2'-Fucosyllactose. Advanced Protein Technologies, Corp.
- GRN 880 (2020). 3'-Sialyllactose. Glycom A/S.
- GRN 881 (2020). 6'-Sialyllactose sodium salt. Glycom A/S.
- GRN 895 (Pending). Lacto-*N*-neotetraose. Glycom A/S.
- GRN 896 (Pending). Alpha-galacto-oligosaccharides. Olygose.
- GRN 897 (2020). 2'-Fucosyllactose. Dupont Nutrition and Health.
- GRN 919 (2020). Lacto-*N*-neotetraose. Jennewein Biotechnologie, GmbH.

GRN 921 (2020). 3'-Sialyllactose sodium salt. Jennewein Biotechnologie, GmbH.

GRN 922 (2021). Lacto-*N*-tetraose. Jennewein Biotechnologie, GmbH.

GRN 923 (2021). Lacto-*N*-tetraose. Jennewein Biotechnologie, GmbH.

GRN 925 (2021). 3'-Fucosyllactose. Jennewein Biotechnologie, GmbH.

Gurung, R.B., Woo, J., Cho, S.S., Hong, J.H., and Kim, D.H. (2018). Gastrointestinal Tolerance and Safety of 3'-Sialyllactose in Subjects Positive with *Helicobacter pylori*: A Pilot Study. *EC Nutrition* 13.9, 600–608.

Hanlon, P.R. (2020). A safety evaluation of mixed human milk oligosaccharides in neonatal farm piglets. *Toxicology Research and Application* 4, 239784732097125.

Hayes, A.W., and Kruger, C.L. (2014). Food Safety and Foodborne Toxicants. In Hayes' principles and methods of toxicology, eds. (Boca Raton: CRC Press), pp. 631.

Hoekstra JH, Szajewska H, Zikri MA, et al. (2004). Oral rehydration solution containing a mixture of non-digestible carbohydrates in the treatment of acute diarrhea: a multicenter randomized placebo controlled study on behalf of the ESPGHAN working group on intestinal infections. *J Pediatr Gastroenterol Nutr* 39, 239-245.

Homann, H.-H., Senkal, M., Kemen, M., and Lehnhardt, M. (2004). The beneficial effects of PHGG in enteral nutrition in medical and surgical patients. *Clinical Nutrition Supplements* 1, 59–62.

Homann, H.H., Kemen, M., Fuessenich, C., Senkal, M., and Zumtobel, V. (1994). Reduction in diarrhea incidence by soluble fiber in patients receiving total or supplemental enteral nutrition. *JPEN J Parenter Enteral Nutr* 18, 486–490.

Hong, Q., Ruhaak, L.R., Totten, S.M., Smilowitz, J.T., German, J.B., and Lebrilla, C.B. (2014). Label-free absolute quantitation of oligosaccharides using multiple reaction monitoring. *Anal Chem* 86, 2640–2647.

Iribarren C, Törnblom H, Aziz I, Magnusson MK, Sundin J, Vigsnaes LK, Amundsen ID, McConnell B, Seitzberg D, Öhman L, Simrén M. Human milk oligosaccharide supplementation in irritable bowel syndrome patients: A parallel, randomized, double-blind, placebo-controlled study. *Neurogastroenterol Motil.* 2020 Oct;32(10):e13920. doi: 10.1111/nmo.13920. Epub 2020 Jun 14. PMID: 32536023.

Kajzer, J., Oliver, J., and Marriage, B. (2016). Gastrointestinal tolerance of formula supplemented with oligosaccharides. *The FASEB Journal* 30, Abstract 671.4.

Karakan, T., Ergun, M., Dogan, I., Cindoruk, M., and Unal, S. (2007). Comparison of early enteral nutrition in severe acute pancreatitis with prebiotic fiber supplementation versus

standard enteral solution: a prospective randomized double-blind study. *World J Gastroenterol* 13, 2733–2737.

Khoshoo, V., Sun, S.S., and Storm, H. (2010). Tolerance of an enteral formula with insoluble and prebiotic fiber in children with compromised gastrointestinal function. *J Am Diet Assoc* 110, 1728–1733.

Lampe, J.W., Effertz, M.E., Larson, J.L., and Slavin, J.L. (1992). Gastrointestinal effects of modified guar gum and soy polysaccharide as part of an enteral formula diet. *JPEN J Parenter Enteral Nutr* 16, 538–544.

Leo, F., Asakuma, S., Fukuda, K., Senda, A., and Urashima, T. (2010). Determination of sialyl and neutral oligosaccharide levels in transition and mature milks of Samoan women, using anthranilic derivatization followed by reverse phase high performance liquid chromatography. *Biosci Biotechnol Biochem* 74, 298–303.

Litten-Brown, J.C., Corson, A.M., and Clarke, L. (2010). Porcine models for the metabolic syndrome, digestive and bone disorders: a general overview. *Animal* 4, 899–920.

Majid, H.A., Cole, J., Emery, P.W., and Whelan, K. (2014). Additional oligofructose/inulin does not increase faecal bifidobacteria in critically ill patients receiving enteral nutrition: a randomised controlled trial. *Clin Nutr* 33, 966–972.

Marriage, B.J., Buck, R.H., Goehring, K.C., Oliver, J.S., and Williams, J.A. (2015). Infants Fed a Lower Calorie Formula With 2'FL Show Growth and 2'FL Uptake Like Breast-Fed Infants. *J Pediatr Gastroenterol Nutr* 61, 649–658.

Meier, R., Beglinger, C., Schneider, H., Rowedder, A., and Gyr, K. (1993). Effect of a liquid diet with and without soluble fiber supplementation on intestinal transit and cholecystokinin release in volunteers. *JPEN J Parenter Enteral Nutr* 17, 231–235.

Meli, F., Puccio, G., Cajozzo, C., Ricottone, G.L., Pecquet, S., Sprenger, N., and Steenhout, P. (2014). Growth and safety evaluation of infant formulae containing oligosaccharides derived from bovine milk: a randomized, double-blind, noninferiority trial. *BMC Pediatrics* 14,

Modi, N., Uthaya, S., Fell, J., and Kulinskaya, E. (2010). A randomized, double-blind, controlled trial of the effect of prebiotic oligosaccharides on enteral tolerance in preterm infants (ISRCTN77444690). *Pediatr Res* 68, 440–445.

Nakao, M., Ogura, Y., Satake, S., Ito, I., Iguchi, A., Takagi, K., and Nabeshima, T. (2002). Usefulness of soluble dietary fiber for the treatment of diarrhea during enteral nutrition in elderly patients. *Nutrition* 18, 35–39.

Nakhla, T., Fu, D., Zopf, D., Brodsky, N.L., and Hurt, H. (1999). Neutral oligosaccharide content of preterm human milk. *Br J Nutr* 82, 361–367.

Nowak-Wegrzyn, Czerkies, Reyes, Collins, and Heine (2019). Confirmed Hypoallergenicity of a Novel Whey-Based Extensively Hydrolyzed Infant Formula Containing Two Human Milk Oligosaccharides. *Nutrients* 11, 1447.

Ofei SY, Fuchs GJ 3rd. (2019) Principles and Practice of Oral Rehydration. *Curr Gastroenterol Rep* 21, 67.

Oli MW, Petschow BW, Buddington RK. Evaluation of fructooligosaccharide supplementation of oral electrolyte solutions for treatment of diarrhea: recovery of the intestinal bacteria. *Dig Dis Sci.* 1998 Jan;43(1):138-47. doi: 10.1023/a:1018892524790. PMID: 9508515.

Palsson, O.S., Peery, A., Seitzberg, D., Amundsen, I.D., McConnell, B., and Simrén, M. (2020). Human Milk Oligosaccharides Support Normal Bowel Function and Improve Symptoms of Irritable Bowel Syndrome: A Multicenter, Open-Label Trial. *Clin Transl Gastroenterol* 11, e00276.

Parente, F., Cucino, C., Anderloni, A., Grandinetti, G., and Bianchi Porro, G. (2003). Treatment of *Helicobacter pylori* infection using a novel antiadhesion compound (3'sialyllactose sodium salt). A double blind, placebo-controlled clinical study. *Helicobacter* 8, 252–256.

Parschat, K., Oehme, A., Leuschner, J., Jennewein, S., and Parkot, J. (2020). A safety evaluation of mixed human milk oligosaccharides in rats. *Food Chem Toxicol* 136, 111118.

Passariello A, Terrin G, De Marco G, et al. (2011), Efficacy of a new hypotonic oral rehydration solution containing zinc and prebiotics in the treatment of childhood acute diarrhea: a randomized controlled trial. *J Pediatr* 158, 288-92.e1.

Peters, A.L., and Davidson, M.B. (1996). Addition of hydrolyzed guar to enteral feeding products in type I diabetic patients. *Diabetes Care* 19, 899–900.

Phipps, K.R., Baldwin, N., Lynch, B., Stannard, D.R., Šoltsová, A., Gilby, B., Mikš, M.H., and Röhrig, C.H. (2018). Preclinical safety evaluation of the human-identical milk oligosaccharide lacto-*N*-tetraose. *Regul Toxicol Pharmacol* 99, 260–273.

Puccio, G., Alliet, P., Cajozzo, C., Janssens, E., Corsello, G., Sprenger, N., Wernimont, S., Egli, D., Gosoni, L., and Steenhout, P. (2017). Effects of Infant Formula With Human Milk Oligosaccharides on Growth and Morbidity: A Randomized Multicenter Trial. *J Pediatr Gastroenterol Nutr* 64, 624–631.

Radke, M., Picaud, J.C., Loui, A., Cambonie, G., Faas, D., Lafeber, H.N., de Groot, N., Pecquet, S.S., Steenhout, P.G., and Hascoet, J.M. (2017). Starter formula enriched in prebiotics and probiotics ensures normal growth of infants and promotes gut health: a randomized clinical trial. *Pediatr Res* 81, 622–631.

Raghupathy P, Ramakrishna BS, Oommen SP, et al. (2006). Amylase-resistant starch as adjunct to oral rehydration therapy in children with diarrhea. *J Pediatr Gastroenterol Nutr* 42, 362-368.

Rasko, D.A., Wilson, T.J., Zopf, D., and Taylor, D.E. (2000). Lewis antigen expression and stability in *Helicobacter pylori* isolated from serial gastric biopsies. *J Infect Dis* 181, 1089–1095.

Riechmann Román E, Moreno Villares JM, Domínguez Ortega F, Carmona Martínez A, Picó Sirvent L, Santana Sandoval L, Casas Rivero J, Alshweki A, Cercamondi C, Dahbane S, Vidal Guevara ML. Real-world study in infants fed with an infant formula with two human milk oligosaccharides. *Nutr Hosp*. 2020 Aug 27;37(4):698-706. English. doi: 10.20960/nh.03084. PMID: 32698596.

Rushdi, T.A., Pichard, C., and Khater, Y.H. (2004). Control of diarrhea by fiber-enriched diet in ICU patients on enteral nutrition: a prospective randomized controlled trial. *Clin Nutr* 23, 1344–1352.

Ryan, J.J., Monteagudo-Mera, A., Contractor, N., and Gibson, G.R. (2021). Impact of 2'-Fucosyllactose on Gut Microbiota Composition in Adults with Chronic Gastrointestinal Conditions: Batch Culture Fermentation Model and Pilot Clinical Trial Findings. *Nutrients* 13, 938.

Scientific Committee on Food (1998). Opinion of the Scientific Committee on Food on the applicability of the ADI (Acceptable Daily Intake) for food additives to infants (expressed on 17/09/1998).

Simakachorn, N., Bibiloni, R., Yimyaem, P., Tongpenyai, Y., Varavithaya, W., Grathwohl, D., Reuteler, G., Maire, J.C., Blum, S., Steenhout, P., Benyacoub, J., and Schiffrin, E.J. (2011). Tolerance, safety, and effect on the faecal microbiota of an enteral formula supplemented with pre- and probiotics in critically ill children. *J Pediatr Gastroenterol Nutr* 53, 174–181.

Simeoni, U., Berger, B., Junick, J., Blaut, M., Pecquet, S., Rezzonico, E., Grathwohl, D., Sprenger, N., Brüssow, H., Study, T., Szajewska, H., Bartoli, J.M., Brevaut-Malaty, V., Borszewska-Kornacka, M., Feleszko, W., François, P., Gire, C., Leclaire, M., Maurin, J.M., Schmidt, S., Skórka, A., Squizzaro, C., and Verdote, J.J. (2016). Gut microbiota analysis reveals a marked shift to bifidobacteria by a starter infant formula containing a synbiotic of bovine milk-derived oligosaccharides and *Bifidobacterium animalis* subsp. *lactis* CNCM I-3446. *Environ Microbiol* 18, 2185–2195.

Spapen, H., Diltor, M., Van Malderen, C., Opendacker, G., Suys, E., and Huyghens, L. (2001). Soluble fiber reduces the incidence of diarrhea in septic patients receiving total enteral nutrition: a prospective, double-blind, randomized, and controlled trial. *Clin Nutr* 20, 301–305.

Steenhout, P., Sperisen, P., Martin, F., Sprenger, N., Wernimont, S., Pecquet, S., and Berger, B. (2016). Term Infant Formula Supplemented with Human Milk Oligosaccharides (2'-Fucosyllactose and Lacto-*N*-neotetraose) Shifts Stool Microbiota and Metabolic Signatures Closer to that of Breastfed Infants. *The FASEB Journal* 30, 275.7–275.7.

Storm, H.M., Shepard, J., Czerkies, L.M., Kineman, B., Cohen, S.S., Reichert, H., and Carvalho, R. (2019). 2'-Fucosyllactose Is Well Tolerated in a 100% Whey, Partially Hydrolyzed Infant Formula With *Bifidobacterium lactis*: A Randomized Controlled Trial. *Glob Pediatr Health* 6, 2333794X19833995.

Sumiyoshi, W., Urashima, T., Nakamura, T., Arai, I., Saito, T., Tsumura, N., Wang, B., Brand-Miller, J., Watanabe, Y., and Kimura, K. (2003). Determination of each neutral oligosaccharide in the milk of Japanese women during the course of lactation. *Br J Nutr* 89, 61–69.

Thurl, S., Henker, J., Siegel, M., Tovar, K., and Sawatzki, G. (1997). Detection of four human milk groups with respect to Lewis blood group dependent oligosaccharides. *Glycoconj J* 14, 795–799.

Thurl, S., Munzert, M., Boehm, G., Matthews, C., and Stahl, B. (2017). Systematic review of the concentrations of oligosaccharides in human milk. *Nutr Rev* 75, 920–933.

Thurl, S., Munzert, M., Henker, J., Boehm, G., Müller-Werner, B., Jelinek, J., and Stahl, B. (2010). Variation of human milk oligosaccharides in relation to milk groups and lactational periods. *British Journal of Nutrition* 104, 1261–1271.

Urashima, T., Asakuma, S., Leo, F., Fukuda, K., Messer, M., and Oftedal, O.T. (2012). The predominance of type I oligosaccharides is a feature specific to human breast milk. *Adv Nutr* 3, 473S–82S.

USDA 2012. What We Eat In America (WWEIA), NHANES: overview. Available from: <http://www.ars.usda.gov/Services/docs.htm?docid=13793#release>. Retrieved 29 January 2018.

USDA 2018. U.S. Department of Agriculture, Agricultural Research Service. 2018. USDA Food and Nutrient Database for Dietary Studies 2015-2016. Food Surveys Research Group Home Page, <http://www.ars.usda.gov/nea/bhnrc/fsrg>. Available at <https://www.ars.usda.gov/northeast-area/beltsville-md-bhnrc/beltsville-human-nutrition-research-center/food-surveys-research-group/docs/fndds-download-databases/>

US Patent 10,695,358. Oral rehydration composition with oligosaccharides. Abbott Laboratories, June 30, 2020.

Vaisman N, Press J, Leibovitz E, Boehm G, Barak V. (2010) Short-term effect of prebiotics administration on stool characteristics and serum cytokines dynamics in very young children with acute diarrhea. *Nutrients* 2, 683-692.

van den Berg, J.P., Westerbeek, E.A., van der Klis, F.R., Sanders, E.A., Berbers, G.A., and van Elburg, R.M. (2015). Response on Pneumococcal Vaccine in Preterm Infants After Neutral and Acidic Oligosaccharides Supplementation. *Pediatr Infect Dis J* 34, 976–982.

Van Niekerk, E., Autran, C.A., Nel, D.G., Kirsten, G.F., Blaauw, R., and Bode, L. (2014). Human Milk Oligosaccharides Differ between HIV-Infected and HIV-Uninfected Mothers and Are Related to Necrotizing Enterocolitis Incidence in Their Preterm Very-Low-Birth-Weight Infants. *The Journal of Nutrition* 144, 1227–1233.

Vandenplas Y, De Hert SG; PROBIOTICAL-study group. (2011). Randomised clinical trial: the synbiotic food supplement Probiotal vs. placebo for acute gastroenteritis in children. *Aliment Pharmacol Ther* 34, 862-867.

WHO (World Health Organization). (2020) World Health Organization Fact Sheet. Infant and young child feeding. <https://www.who.int/news-room/fact-sheets/detail/infant-and-young-child-feeding>.

Wireko, B.M., and Bowling, T. (2010). Enteral tube feeding. *Clin Med (Lond)* 10, 616–619.

B. EXPERT PANEL STATEMENT

We, the members of the Expert Panel, qualified by scientific training and experience to evaluate the safety of substances directly or indirectly added to food, have performed a comprehensive and critical review of available information and data on the safety and Generally Recognized As Safe (GRAS) status of Lacto-*N*-tetraose (LNT) in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas has been shown to be safe and GRAS, using scientific procedures, under the Federal Food, Drug, and Cosmetic Act (FFDCA), as described under 21 CFR §170.30(b). The safety of the intake of LNT toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas has been determined to be GRAS by demonstrating that the safety of this level of intake is generally recognized by experts qualified by both scientific training and experience to evaluate the safety of substances directly added to food and is based on generally available and accepted information.

The use of LNT as an ingredient for the intended use in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas has been determined to be safe through scientific procedures set forth under 21 CFR §170.30(b) based on the following:

1. The subject of this GRAS Determination is a spray-dried, powdered food ingredient that contains not less than 75% LNT dry weight. The remaining components include carbohydrate by-products, ash, and moisture.
 - a. LNT is a neutral, non-fucosylated oligosaccharide in human milk.
 - b. Published studies show that the amount of LNT in human milk ranges from 0.003 to 6.7 g/L, with means and medians ranging from 0.1 to 3.9 and 0.2 to 2.1 g/L, respectively.
 - c. Human milk oligosaccharides, including LNT, are resistant to the digestive enzymes in the gastrointestinal tract, poorly absorbed, and pass through the gastrointestinal tract where they are either fermented by the microbiota or excreted unchanged.
2. The subject of this GRAS Determination is also the subject of GRAS Notice 923, which received a “no questions” letter on February 2, 2021 for the use of LNT in non-exempt term infant formula.

- a. The subject of this GRAS Determination is manufactured using a genetically engineered strain of *Escherichia coli* BL21(DE3) by Chr. Hansen A/S in Food Safety System Certification (FSSC) 22000-, ISO 9001:2015-, GMP-, and/or International Featured Standards Food 6.1-compliant facilities. Chr. Hansen A/S is a Food Facility registered with FDA.
- b. The genetically engineered strain of *Escherichia coli* BL21(DE3) used by Chr. Hansen A/S is not toxigenic and not capable of DNA transfer to other organisms and has the same virulence profile as *E. coli* BL21(DE3).
- c. All raw materials, processing aids, and food contact substances are GRAS and/or conform to the specifications stated in 21 CFR and/or the Food Chemicals Codex (FCC).
- d. Process procedures and product specifications are in place to control the levels of carbohydrate by-products, as well as heavy metals, microbes, and production organism-derived DNA and possible endotoxin, ensuring a consistent, safe, food-grade finished ingredient.
- e. The available stability studies indicate a shelf-life of two years when stored from the date of production under ambient conditions.
- f. Use of the subject of this GRAS Determination in the intended selected conventional foods and enteral tube feeding formulas results in mean and 90th percentile estimated daily intakes (EDIs) of 1.02 and 2.33 g/day (0.015 and 0.035 g/kg bw/day) for consumers not less than 2 years old.
- g. Use of the subject of this GRAS Determination in selected conventional foods and enteral tube feeding formulas results in mean and 90th percentile cumulative estimated daily intakes (EDIs) of 0.463 and 1.05 g/day (0.007 and 0.016 g/kg bw/day) for consumers not less than 2 years old.
- h. Use of the subject of this GRAS Determination in oral electrolyte solutions results in an estimated daily intake of 0.8 – 1.6 g LNT (equivalent to 59.3 – 118.5 mg of LNT/kg bw/day assuming a 13.5 kg toddler and 11.4 – 22.9 mg of LNT/kg bw/day assuming a 70 kg adult). Because OESs are intended for short term use, intake of LNT from OESs will not impact the cumulative LNT intake resulting from the use of LNT in selected conventional foods and enteral tube feeding formulas.

3. Genotoxicology and rat subchronic toxicology studies published by Phipps et al. (2018) show that LNT is not genotoxic and has a no observed adverse effect level (NOAEL) of 4 g/kg bw/day, which was the highest dose tested.
4. The safety of exposure to Chr. Hansen A/S's LNT per its intended uses and intended use levels is supported by:
 - a. Data demonstrating the qualitative and quantitative similarities between the subject of this GRAS Notice and the LNT ingredient tested in the pivotal genotoxicology and subchronic toxicology studies conducted by Phipps et al. (2018);
 - b. The lack of genotoxicity and no observed adverse effect level (NOAEL) for LNT established in the 90-day subchronic dietary toxicology conducted by Phipps et al. (2018);
 - c. Published genotoxicology, 90-day subchronic dietary toxicology, and neonatal piglet studies with mixtures containing the subject of this GRAS Determination (Parschat et al., 2019; Hanlon, 2020);
 - d. Clinical data showing the ingestion of synthetic forms of HMOs is well tolerated in infants up to 1.0 g/day and adults up to 20 g/day;
 - e. Clinical data showing that the use of other non-digestible carbohydrates in infants, adults, enteral tube feeding products, and oral electrolyte solutions is well tolerated up to 63 g/day;
 - f. The GRAS status of the subject of this GRAS Determination for use in infant formula (GRN 923);
 - g. The GRAS status of LNT for use in selected conventional foods (GRN 833).

Therefore, LNT is safe and GRAS at the proposed level of addition to the intended toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas. Lacto-*N*-tetraose is, therefore, excluded from the definition of a food additive, and may be used in the U.S. without the promulgation of a food additive regulation by the FDA under 21 CFR.

Peter Pressman, MD, MS, FACN,
GRAS Expert Panel Member
Medicine Public Health & Nutrition
The Daedalus Foundation

Signature:



Date: May 18, 2021

A. Wallace Hayes, PhD, DABT, FATS, ERT
GRAS Expert Panel Member
Harvard School of Public Health

Signature:



Date: May 18, 2021

Thomas E. Sox, PhD, JD
GRAS Expert Panel Member
Principal, Pondview Consulting LLC

Signature:



Date: May 18, 2021

Claire Kruger, PhD, DABT
Scientific Advisor to the Panel

Signature:



Date: May 18, 2021

FDA USE ONLY

GRN NUMBER 001017	DATE OF RECEIPT Jun 8, 2021
ESTIMATED DAILY INTAKE	INTENDED USE FOR INTERNET
NAME FOR INTERNET	
KEYWORDS	

DEPARTMENT OF HEALTH AND HUMAN SERVICES
Food and Drug Administration
**GENERALLY RECOGNIZED AS SAFE
(GRAS) NOTICE** (Subpart E of Part 170)

Transmit completed form and attachments electronically via the Electronic Submission Gateway (*see Instructions*); OR Transmit completed form and attachments in paper format or on physical media to: Office of Food Additive Safety (*HFS-200*), Center for Food Safety and Applied Nutrition, Food and Drug Administration, 5001 Campus Drive, College Park, MD 20740-3835.

SECTION A – INTRODUCTORY INFORMATION ABOUT THE SUBMISSION

1. Type of Submission (*Check one*)
 New Amendment to GRN No. _____ Supplement to GRN No. _____

2. All electronic files included in this submission have been checked and found to be virus free. (*Check box to verify*)

3. Most recent presubmission meeting (*if any*) with FDA on the subject substance (*yyyy/mm/dd*): _____

4. For Amendments or Supplements: Is your amendment or supplement submitted in response to a communication from FDA? (*Check one*)
 Yes If yes, enter the date of communication (*yyyy/mm/dd*): _____
 No

SECTION B – INFORMATION ABOUT THE NOTIFIER

1a. Notifier	Name of Contact Person Kate Urbain	Position or Title Head of Regulatory Affairs North America	
	Organization (<i>if applicable</i>) Chr. Hansen A/S		
	Mailing Address (<i>number and street</i>) 9015 W Maple St.		
City West Allis	State or Province Wisconsin	Zip Code/Postal Code 53214	Country United States of America
Telephone Number 414-607-5819	Fax Number	E-Mail Address USKAUR@chr-hansen.com	
1b. Agent or Attorney (if applicable)	Name of Contact Person Dietrich B. Conze	Position or Title Managing Partner	
	Organization (<i>if applicable</i>) Spherix Consulting Group, Inc.		
	Mailing Address (<i>number and street</i>) 751 Rockville Pike, Unit 30-B		
City Rockville	State or Province Maryland	Zip Code/Postal Code 20852	Country United States of America
Telephone Number 240-367-6089	Fax Number	E-Mail Address dconze@spherixgroup.com	

SECTION C – GENERAL ADMINISTRATIVE INFORMATION

1. Name of notified substance, using an appropriately descriptive term

Lacto-N-tetraose (LNT)

2. Submission Format: *(Check appropriate box(es))*

- Electronic Submission Gateway Electronic files on physical media
 Paper
If applicable give number and type of physical media

3. For paper submissions only:

Number of volumes _____

Total number of pages _____

4. Does this submission incorporate any information in CFSAN's files? *(Check one)*

- Yes *(Proceed to Item 5)* No *(Proceed to Item 6)*

5. The submission incorporates information from a previous submission to FDA as indicated below *(Check all that apply)*

- a) GRAS Notice No. GRN 923
 b) GRAS Affirmation Petition No. GRP _____
 c) Food Additive Petition No. FAP _____
 d) Food Master File No. FMF _____
 e) Other or Additional *(describe or enter information as above)* GRN 546, 571, 650, 659, 735, 749, 766, 815, 852, 880, 897, 919, 925

6. Statutory basis for conclusions of GRAS status *(Check one)*

- Scientific procedures *(21 CFR 170.30(a) and (b))* Experience based on common use in food *(21 CFR 170.30(a) and (c))*

7. Does the submission (including information that you are incorporating) contain information that you view as trade secret or as confidential commercial or financial information? *(see 21 CFR 170.225(c)(8))*

- Yes *(Proceed to Item 8)*
 No *(Proceed to Section D)*

8. Have you designated information in your submission that you view as trade secret or as confidential commercial or financial information *(Check all that apply)*

- Yes, information is designated at the place where it occurs in the submission
 No

9. Have you attached a redacted copy of some or all of the submission? *(Check one)*

- Yes, a redacted copy of the complete submission
 Yes, a redacted copy of part(s) of the submission
 No

SECTION D – INTENDED USE

1. Describe the intended conditions of use of the notified substance, including the foods in which the substance will be used, the levels of use in such foods, and the purposes for which the substance will be used, including, when appropriate, a description of a subpopulation expected to consume the notified substance.

Chr. Hansen A/S intends to use LNT as an ingredient in toddler formulas, foods for infants and young children, meal replacements drinks for adults, non-carbonated drinks, bars, oral electrolyte solutions, and enteral tube feeding formulas.

2. Does the intended use of the notified substance include any use in product(s) subject to regulation by the Food Safety and Inspection Service (FSIS) of the U.S. Department of Agriculture?

(Check one)

- Yes No

3. If your submission contains trade secrets, do you authorize FDA to provide this information to the Food Safety and Inspection Service of the U.S. Department of Agriculture?

(Check one)

- Yes No, you ask us to exclude trade secrets from the information FDA will send to FSIS.

SECTION E – PARTS 2 -7 OF YOUR GRAS NOTICE

(check list to help ensure your submission is complete – PART 1 is addressed in other sections of this form)

- PART 2 of a GRAS notice: Identity, method of manufacture, specifications, and physical or technical effect (170.230).
- PART 3 of a GRAS notice: Dietary exposure (170.235).
- PART 4 of a GRAS notice: Self-limiting levels of use (170.240).
- PART 5 of a GRAS notice: Experience based on common use in foods before 1958 (170.245).
- PART 6 of a GRAS notice: Narrative (170.250).
- PART 7 of a GRAS notice: List of supporting data and information in your GRAS notice (170.255)

Other Information

Did you include any other information that you want FDA to consider in evaluating your GRAS notice?

Yes No

Did you include this other information in the list of attachments?

Yes No

SECTION F – SIGNATURE AND CERTIFICATION STATEMENTS

1. The undersigned is informing FDA that Chr. Hansen A/S
(name of notifier)

has concluded that the intended use(s) of Lacto-N-tetraose (LNT)
(name of notified substance)

described on this form, as discussed in the attached notice, is (are) not subject to the premarket approval requirements of the Federal Food, Drug, and Cosmetic Act based on your conclusion that the substance is generally recognized as safe recognized as safe under the conditions of its intended use in accordance with § 170.30.

2. Chr. Hansen A/S
(name of notifier) agrees to make the data and information that are the basis for the conclusion of GRAS status available to FDA if FDA asks to see them; agrees to allow FDA to review and copy these data and information during customary business hours at the following location if FDA asks to do so; agrees to send these data and information to FDA if FDA asks to do so.

9015 W Maple St, West Allis, WI 53214
(address of notifier or other location)

The notifying party certifies that this GRAS notice is a complete, representative, and balanced submission that includes unfavorable, as well as favorable information, pertinent to the evaluation of the safety and GRAS status of the use of the substance. The notifying party certifies that the information provided herein is accurate and complete to the best of his/her knowledge. Any knowing and willful misinterpretation is subject to criminal penalty pursuant to 18 U.S.C. 1001.

**3. Signature of Responsible Official,
Agent, or Attorney**

Dietrich B. Conze, PhD Digitally signed by Dietrich B. Conze, PhD
Date: 2021.06.04 11:22:50 -04'00'

Printed Name and Title

Dietrich B. Conze, PhD, Managing Partner

Date (mm/dd/yyyy)

06/04/2021

SECTION G – LIST OF ATTACHMENTS

List your attached files or documents containing your submission, forms, amendments or supplements, and other pertinent information. Clearly identify the attachment with appropriate descriptive file names (or titles for paper documents), preferably as suggested in the guidance associated with this form. Number your attachments consecutively. When submitting paper documents, enter the inclusive page numbers of each portion of the document below.

Attachment Number	Attachment Name	Folder Location (select from menu) (Page Number(s) for paper Copy Only)
	Chr Hansen LNT GRAS to FDA.pdf	Submission
	References	Submission

OMB Statement: Public reporting burden for this collection of information is estimated to average 170 hours per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to: Department of Health and Human Services, Food and Drug Administration, Office of Chief Information Officer, PRASStaff@fda.hhs.gov. (Please do NOT return the form to this address). An agency may not conduct or sponsor, and a person is not required to respond to, a collection of information unless it displays a currently valid OMB control number.