

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22

U.S. FOOD AND DRUG ADMINISTRATION
CENTER FOR DRUG EVALUATION AND RESEARCH
AND
JOHNS HOPKINS UNIVERSITY CERSI WORKSHOP

Addressing Challenges in the Design and
Analysis of Rare Disease Clinical Trials:
Considerations and Tools

Day 2

Wednesday, May 3, 2023

9:00 a.m. to 11:58 a.m.

1 **Meeting Roster**

2 **Frank Harrell, PhD**

3 Expert Biostatistics Advisor to CDER, FDA and
4 Professor of Biostatistics, Vanderbilt University

5
6 **Rima Izem, PhD**

7 Director of Statistical Methodology Group, Novartis
8 Pharma

9
10 **Kelley Kidwell, PhD**

11 Associate Professor & Associate Chair for Academic
12 Affairs in Biostatistics, School of Public Health,
13 University of Michigan

14
15 **J. Jack Lee, PhD, DDS**

16 Professor, Department of Biostatistics, University
17 of Texas MD Anderson Cancer Center

18
19 **Gregory Levin, PhD,**

20 Associate Director for Statistical Science and
21 Policy, Office of Biostatistics, Center for Drug
22 Evaluation and Research (CDER), FDA

1 **Dionne Price, PhD**

2 Deputy Director, Office of Biostatistics, Office of
3 Translational Sciences, CDER, FDA

4
5 **Karen Price, PhD**

6 Associate Vice President and Statistical Officer,
7 Statistical Innovation Center, Eli Lilly & Company

8
9 **Michael Rosenblum, PhD**

10 Professor of Biostatistics, Johns Hopkins Bloomberg
11 School of Public Health

12
13 **Noah Simon, PhD**

14 Associate Professor, Department of Biostatistics,
15 University of Washington

16
17 **Nigel Stallard, MSc, PhD**

18 Professor of Medical Statistics and Deputy Director
19 of Clinical Trials Unit, Warwick Medical School,
20 United Kingdom

21

22

1	C O N T E N T S	
2	AGENDA ITEM	PAGE
3	Design and Analysis Methods for Clinical	
4	Trials for Rare Diseases	
5	Welcome	
6	Dionne Price, PhD	6
7	Session 1: Adaptive Designs in	
8	Small Populations	
9	Moderator: Michael Rosenblum, PhD	7
10	Panelists	
11	SMART Design and Bayesian Methods for	
12	Rare Disease Trials	
13	Kelley Kidwell, PhD	8
14	Adaptive Enrichment Designs in the	
15	Rare Disease Setting	
16	Noah Simon, PhD	29
17	Clinical Trials in Rare Diseases:	
18	Should We Do Them Differently?	
19	Nigel Stallard, MSc, PhD	47
20	Summary of Presentations	
21	Gregory Levin, PhD	65
22	Q&A with Panelists	71

1	C O N T E N T S (continued)	
2	AGENDA ITEM	PAGE
3	Session 2: Analysis Methods in	
4	Small Populations	
5	Moderator: Michael Rosenblum, PhD	80
6	Panelists	
7	Bayesian Methods and Master Protocols in	
8	Rare Disease Drug Development	
9	Karen Price, PhD	81
10	Bayesian Information Borrowing for	
11	Efficient and Accurate Statistical	
12	Inference in Rare Diseases	
13	J. Jack Lee, PhD, DDS	100
14	Randomized and Non-Randomized Designs for	
15	Causal Inference with Longitudinal Data in	
16	Rare Disorders	
17	Rima Izem, PhD	113
18	Summary of Presentations	
19	Frank Harrell, PhD	132
20	Concluding Remarks	
21	Dionne Price, PhD	142
22		

P R O C E E D I N G S

(9:00 a.m.)

Welcome

DR. D. PRICE: Good morning, everyone, and welcome to day 2 of our workshop on Addressing Challenges in the Design and Analysis of Rare Disease Clinical Trials. Our focus yesterday was on quality and fit-for-purpose data for use in rare disease trials and how that data collected from patients could be used to inform drug development. Today, we will focus on design and analysis methodologies that may be useful in settings with small numbers of patients.

The first session will focus on adaptive designs and the second session will focus on analysis methods in small populations. We've assembled an experienced group of speakers and panelists who will share their insights and knowledge with us today.

Both sessions will be moderated by Dr. Michael Rosenblum. Dr. Rosenblum is Professor of Biostatistics at the Johns Hopkins Bloomberg

1 School of Public Health. His research is in causal
2 inference, the design and analysis of clinical
3 trials, and enhancing capacity in low- and
4 middle-income countries in statistical methods for
5 clinical trial design and analysis, and he is a
6 fellow of the American Statistical Association.

7 So, without further delay, I will turn it
8 over to Dr. Michael Rosenblum. Thank you.

9 **Session 1 - Michael Rosenblum**

10 DR. ROSENBLUM: Thank you, Dionne, and
11 welcome, everyone. Thank you for joining today. I
12 want to remind everyone, if you have questions,
13 please put them in the Q&A. We won't be
14 recognizing the raise-hand feature, so any
15 questions, please put them in the Q&A, and we'll
16 try to answer them.

17 The setup for this first session, as well as
18 the second, is 20-minute talks, three of them,
19 followed by 20 minutes of Q&A. And I want to cut
20 right to the chase and introduce the first of our
21 three outstanding speakers in this first session.

22 Kelley Kidwell is an Associate Professor and

1 Associate Chair for Academic Affairs and
2 Biostatistics at the University of Michigan School
3 of Public Health. She has multiple areas of
4 expertise, including sequential multiple
5 assignment, randomized trials, also called SMART
6 trials, and other novel trial designs. She's
7 collaborated in a wide variety of fields, including
8 oncology, mental health, and rare and chronic
9 diseases. We're very lucky to have Kelley Kidwell
10 speaking today, and I will hand it over to you.

11 **Presentation - Kelley Kidwell**

12 DR. KIDWELL: Thank you so much.

13 OPEN SLIDE SET. So yes, I'm so grateful to
14 be here. Thank you. I'm grateful to start the day
15 off. I'm going to talk about SMART Design and
16 Bayesian Methods for Rare Disease Trials. Next
17 slide please. As many of you already know the
18 challenges in rare disease research -- I won't
19 spend too long on this -- quite obviously, the
20 small number of patients make this very difficult
21 to come up with robust conclusions, and sometimes
22 when we're interested in especially binary

1 endpoints -- success, failure, response,
2 non-response -- there's even a smaller number of
3 those endpoint events.

4 Something else that is really notable is
5 that because of the small number of patients, it's
6 quite challenging to run separate dose-finding
7 trials, along with additional separate confirmatory
8 trials. So trying to think of designs in which
9 those can be combined is really important in this
10 area. Then it's also very difficult to meet those
11 standard frequentist benchmarks, like 80 percent
12 power and 5 percent two-sided type 1 error that we
13 generally think of being somewhat almost necessary
14 or expected in clinical trials. Next slide please.

15 There's a clear need for clinical trial
16 innovation, both on the design side and also in the
17 analytic aspect. In the design side, we want a
18 trial that is of minimal size but will still
19 provide robust evidence. We also would like the
20 design that we know it's not necessary that the
21 patients are going to do very well in the trial,
22 and it's more for the future benefit; however, in

1 order to get patients interested, recruited, and
2 retained in the trial, we'd like to have some sort
3 of benefit for participants. So maximize the
4 chance of receiving therapy and minimize the number
5 of individuals receiving placebo or standard of
6 care. It'd also be great for design to consider
7 more than one dose of treatment and confirm its
8 efficacy.

9 On the analytic side, we'd like to provide
10 estimates of the treatment effect with clinical
11 interpretability, and if possible, incorporate
12 external data such as natural history studies,
13 registries, previous trials, expert opinion, and
14 formalize the content of that.

15 The Complex Innovative Design program,
16 particularly thinking about Bayesian design and
17 analysis, has really kind of allowed us to think
18 more innovatively about clinical trial design,
19 especially in small samples in rare diseases. Next
20 slide please.

21 A lot of my work started about seven years
22 ago when we were discussing this trial in isolated

1 skin vasculitis, a rare disease. This is the first
2 snSMART. We call it the small sample SMART that's
3 in the field. The clinicaltrials.gov number is
4 here. It's called a randomized, multicenter trial
5 in isolated skin vasculitis, or ARAMIS for short,
6 and it looks like this.

7 The physicians were using multiple
8 treatments; however, they didn't know which one was
9 really better. So this is a comparative
10 effectiveness study where we have treatments A
11 versus B, versus C, and patients are followed for
12 some time in ARAMIS at 6 months, where we expect
13 the treatment to be effective. And if it's not
14 effective, then participants are re-randomized. So
15 those who didn't respond are re-randomized to the
16 other two treatments that they didn't first
17 receive. So if they got A, then they'd be
18 randomized to B or C. If they did respond, they
19 continue on that treatment.

20 The goal of this study, while there are two
21 stages, is actually to estimate the first stage
22 treatment response, so to say is it best to get A,

1 or B, or C, but we want to use data across those
2 two stages, so we've developed methods that are
3 appropriate for whether the outcome is either
4 binary like a response yes/no or continuous like a
5 score. Next slide please.

6 I want to describe this design a little bit
7 more. SMART comes from the term "sequential,
8 multiple assignment, randomized trial," and these
9 designs really came together in the last two or
10 three decades, two decades or so, in the early
11 2000s, really thinking about more in the phase 2
12 space for large samples, and to think about these
13 treatment sequences. We're kind of forgetting
14 about the original intent of SMART, but thinking
15 about using that design, a multistage design, where
16 the second stage of randomization depends upon
17 response to the initial treatment, and using it in
18 a small sample to get more information from a small
19 number of individuals.

20 This is a multistage design. Patients can
21 be re-randomized based on response to initial
22 treatments. You might want to think about it as

1 like a restricted crossover design. Crossover
2 designs are used quite often in small samples.
3 First, a patient gets A, and then they get B, or
4 they first get B, and then they get A. Here, we're
5 going to say, "Well, I don't want to give everybody
6 that second treatment because if they did well,
7 then they probably just want to stay on that
8 treatment, and that will help with retention.
9 Instead, if they didn't do well, then we can
10 re-randomize them." So the SMART design in this
11 aspect is more of this restricted crossover, but
12 we're going to be able to use both stages of data
13 in our analytic method to get a more efficient
14 treatment estimate.

15 Now, this design isn't appropriate for all
16 rare diseases. It's really only appropriate for
17 ones which are chronic and relatively stable over
18 the trial period, so the two trial stages. Next
19 slide please.

20 After we did some work on comparative
21 effectiveness, the three active treatments,
22 snSMART, we thought, you know what, this design

1 would actually be maybe more useful, considering
2 how we could register a drug for a small sample.
3 So the fact that there isn't often dose finding in
4 rare diseases, because we don't have enough
5 patients to go from a dose finding trial to an
6 additional confirmatory trial, we thought we could
7 actually do this in one trial.

8 We've been developing these snSMART designs
9 considering two dose levels and placebo. Here's a
10 design where all participants are randomized
11 between placebo low and high dose. They're
12 followed for some time in which we expect to see
13 the treatment effect, and then both placebo and
14 low-dose groups are re-randomized between low and
15 high dose. So you can see here that while there
16 are some patients that received placebo, at the end
17 of the two stages of the trial, all participants
18 received some dose of the trial. So this, again,
19 may help with recruitment and retention in the
20 trial.

21 For those on high dose, if the individuals
22 didn't respond, as long as that dose is safe, they

1 could stay on high dose, perhaps they need it
2 longer; whereas if they did respond, perhaps low
3 dose could be just as effective but maybe have less
4 side effects, so we could re-randomize individuals
5 between low and high dose. Next slide please.

6 We've modified the design slightly, which is
7 perhaps maybe more patient-centered. So all those
8 on placebo would be re-randomized as before, but
9 for those on low dose, if they didn't respond,
10 they'd get high dose, assuming it's safe, and if
11 they did respond, they could stay on low dose;
12 perhaps they don't want to change.

13 For those who don't respond on high dose,
14 perhaps it's not as accepted to continue to give
15 high dose; instead, they'd sort of be off trial and
16 given physician's choice, whereas those who did
17 respond, similarly to the last design, could be re-
18 randomized.

19 Again, the goal here is to use all of the
20 data from stages 1 and phase 2 to decide is a low
21 dose effective? Is it more effective than placebo?
22 Is high dose more effective than placebo? And

1 could you go forward with low dose as opposed to
2 high dose? Next slide please.

3 Actually, as we're developing these methods,
4 we noticed that some trials in the rare disease
5 world were using something very similar. This is
6 an actual trial. It's the SPITFIRE trial. The
7 clinicaltrials.gov number is here. It was a
8 two-phase, placebo-controlled study of two dose
9 levels of treatment in ambulatory boys with
10 Duchenne muscular dystrophy or DMD. You can see
11 that here, only the placebo group was re-randomized
12 in stage 2 to low or high dose. Again, this is
13 something that is useful for recruitment. Those
14 who got low or high dose, they continued their low
15 or high dose. Here, it was likely to see long-term
16 effects.

17 Now, in their primary analysis, they only
18 were planning to use that stage 1 data. Their
19 primary outcome was the change from baseline to
20 week 48 in the 6-minute walk distance, and that
21 secondary outcome was the NSAA score or ambulatory
22 score. You can see that this design is quite

1 similar to the snSMART design that I just showed
2 you; however, we use this in comparison to what if
3 they did an snSMART and actually use that
4 second-stage data. We could see more efficiency,
5 and I'm going to come back to that in a few slides.
6 Next slide please.

7 There are many advantages of this snSMART
8 with the dose levels; the fact that many
9 participants will receive a higher dose level in
10 stage 2, or at least some dose level, so this can
11 really help with engagement and retention, and also
12 recruitment. The design allows for both dose
13 finding and dose confirmation, which allows us to
14 find the dose effect and also register the dose
15 within one trial as opposed to requiring two. This
16 is really helpful as one of the most common reasons
17 for not being able to register a drug, being the
18 fact that the correct dose wasn't found.

19 The analysis can formally incorporate expert
20 opinion or external control data, external co-data,
21 from previous trials or registries, and this can
22 really help with the efficiency of the treatment

1 effect estimates but also decrease the number of
2 individuals on the placebo arm. Next slide please.
3 We've worked the analytic methods in the Bayesian
4 framework, and I'm not going to show you any
5 equations here, but I want to give you some
6 intuition for what's happening, and if you're
7 interested in the actual equations, I'm happy to
8 share those papers with you, so feel free to follow
9 up with me.

10 Our goal is to estimate the first stage
11 response rate, or the mean outcomes, of each
12 treatment by pooling together the data across both
13 stages. We want to provide credible intervals of
14 the effect, or the differences between the dose
15 levels and placebo, or the differences between the
16 treatments, and those are going to contain the true
17 effect with some particular probability. This is
18 actually the definition of a credible interval that
19 most people think is what a confidence interval is.

20 With this Bayesian framework, we must shift
21 our focus away from significance and p-values and
22 really focus on the estimate itself, and we can

1 formally incorporate expert opinion, historical
2 data, or external control data, which is going to
3 help increase our precision, and we can formally
4 test the effects of that as well. I know people
5 are very worried about Bayesian framework in terms
6 of the priors set, but you can rigorously test the
7 effect of these priors, the effect of that prior
8 information. Next slide.

9 The Bayesian framework is really set up such
10 that, actually, we think we don't know the true
11 population parameters, so we're trying to figure
12 out what they are. They're random; they can
13 change. So we take our best guess at these
14 response rates based on our current knowledge,
15 which might be expert opinion, registry of prior
16 trials, and that's this red curve. This is our
17 start of the trial idea of what we think the
18 estimates are going to be.

19 Then we collect data to observe the response
20 rates in the trial, so that's going to be the data
21 that we see, or that distribution of the effect
22 that we see in this blue curve, the likelihood.

1 Then we combine the prior information with our
2 likelihood, and we get this purple curve, the
3 purple distribution, which is now our updated
4 estimate of the response rates or our posterior
5 distribution. Next slide.

6 These prior distributions can be informed by
7 clinical investigators or historical data, and we
8 do want them to be slightly informative; however,
9 we don't want them to overtake the data in the
10 trial, so we only want them to be a few people's
11 worth of information. For example, say we plan on
12 having 60 individuals in our trial, our prior
13 distributions might be worth the strength of two or
14 three individuals, so not remotely going to take
15 over the amount of information coming in from the
16 trial.

17 We also can use what's called a mixture
18 approach, which says, well, we can have some
19 informative prior, but we can combine that with a
20 non-informative prior if we're worried about the
21 effect of that prior being overly informative; then
22 let's combine it with a non-informative prior and

1 see how that affects our results. And like I said,
2 we can really test the sensitivity of our results,
3 given different prior distributions, to make sure
4 that we're not being overly informative.

5 Then for our Bayesian analysis
6 approach -- we call it the Bayesian joint stage
7 model -- we model the first stage simply, then we
8 model the second stage data conditional on the
9 outcome, whether they responded or not. Basically
10 what we try to do is link the first stage response
11 rate to the second stage response rate via what we
12 call linkage parameters, and this induces some
13 within-patient correlation and says, well, we know
14 the second stage outcome isn't as good as the first
15 stage outcome, if we're estimating the first stage
16 outcome, because they already responded or didn't
17 respond. So we want to basically kind of discount
18 that data in some way, but we also want to be able
19 to use it because it's still very helpful in
20 getting these treatment effect estimates. Next
21 slide.

22 Obviously, if we're going to incorporate

1 external data, it's very important to have a
2 careful choice of this control data. There's some
3 some criteria, the Pocock criteria, to assess the
4 similarity between external controls and trial data
5 that include trying to make things similar across
6 inclusion/exclusion criteria, endpoint definition
7 control treatment, distribution of demographic
8 data, et cetera. And something important, as I've
9 mentioned already, or as I mentioned for the prior,
10 is that we also want the number of external control
11 patients, or really the effect of sample size of
12 this prior information or this external control
13 data, not to exceed the number of controls on our
14 trial. But what's really advantageous about
15 incorporating this data is that we can then lower
16 the number of participants on the placebo arm,
17 which makes the recruitment perhaps a bit easier
18 for an snSMART or trial that includes a placebo arm
19 in rare diseases. Next slide, thank you.

20 Our models do have some assumptions. Right
21 now, we don't incorporate patient or disease
22 characteristics or covariates into the model. We

1 often make the simplifying assumptions about the
2 parameters that link the first stage and second
3 stage outcomes together. We do assume that there's
4 some sort of washout period or no carryover effect
5 between the first and the second stage, so much
6 like a crossover trial.

7 Right now, we're only working with one
8 endpoint of interest, and we also assume no to low
9 missing data; however, we're working on extensions
10 to all of these assumptions. Also, once you use
11 our model, you can test many of these assumptions
12 and how appropriate they are in your settings in
13 various ways, and we provide some guidelines to
14 that. Next, please.

15 What we found is that our Bayesian joint
16 stage models provide estimates with very low to no
17 bias and are much more efficient or have lower
18 variance than other estimates. I have "double
19 whammy" on the page because, really, with the two-
20 stage design plus the Bayesian model, we have this
21 double-whammy effect in which we've got these more
22 efficient and robust estimates of the treatment

1 effect, and we can test our sensitivity to our
2 various assumptions. Next slide.

3 So going back to that SPITFIRE trial, we
4 simulated some of the data and reanalyzed the data,
5 incorporating some external control data from one
6 of the largest natural history studies in DMD,
7 CINRG. What you can see is the first row if we
8 just use the first stage of data, like a
9 traditional analytic approach, which they used for
10 the SPITFIRE analysis. The next two rows are using
11 methods which we've developed, the Bayesian joint
12 stage model and a robust meta-analytic combined
13 model, which I didn't go into, but it really
14 formally incorporates that external control data.

15 What you can see is that these estimates are
16 quite the same, 1.8 and 1.6, but the credible
17 intervals are smaller, or we have more efficient
18 estimates with these Bayesian joint stage models,
19 or the Robust MAC, because we've used both stages
20 worth of data, and we formally incorporated
21 external control data, so we can see the savings.
22 Next slide.

1 So you might be thinking, "Okay. If I'm
2 interested in running this, well, how do I size
3 it?" We have two sample size calculators out there
4 in Rshiny applet ready to go if you're interested
5 in three active comparators and you have a binary
6 outcome. We can provide the sample size such that
7 you have some amount of probability, say 80 percent
8 probability, for the 90 percent credible interval
9 of the difference between the best and second best
10 treatment to exclude zero. Here are some results,
11 for example, where you'd need 28 participants per
12 arm or just under 90 patients, depending upon the
13 treatment difference that you're interested in
14 seeing. That Rshiny applet is available online.
15 Next slide please.

16 We also have an Rshiny applet, which is soon
17 to be online, for the placebo high and low dose
18 trial, where you have continuous outcomes, and
19 here, we can really quantify the sample size
20 savings. If we look at the first row, scenario 1,
21 you can see that if you had a one-stage design and
22 a frequentist analysis, say like comparison of the

1 means, like a t-test or so, you would have
2 50 participants per arm.

3 Now, because we've formally incorporated the
4 external control data or have some prior
5 information, we save four patients with a Bayesian
6 analysis, so we need 46 per arm. However, if we
7 have a two-stage design with the snSMART and use
8 our Bayesian joint stage models, we only require
9 31 participants per arm. So you can see there's
10 this massive savings, 62 to 67 percent of savings
11 in our sample size by using the two-stage data and
12 the Bayesian joint stage model. So again, that
13 double-whammy effect, where we're seeing that we
14 can reduce the sample size from a one-stage design
15 by 15 to 60 percent. Next slide please.

16 All of our current methods, Bayesian, and
17 then we have similar frequentist methods, are
18 available in an R package called snSMART, which you
19 can download and use. We have a number of papers
20 already in the field, and there are two snSMARTs in
21 the field that I know of right now. There may be
22 one or more additional ones, but ARAMIS was sort of

1 our initial motivating. Then there is a more
2 recent one called MISTIC, which has just been
3 written about, the protocol paper, and is in the
4 field. Next slide please.

5 In summary, the snSMART design and Bayesian
6 joint stage models fit under this complex
7 innovative design program for comparative
8 effectiveness, but also for confirmatory
9 dose-finding and confirmatory drug comparisons.
10 But really, I'm not saying this is for all rare
11 diseases. This is for chronic stable rare
12 diseases, but this design has potential to aid in
13 recruitment and retention. The design and analysis
14 can both dose-find and dose-confirm that best dose
15 level. When we use a two-stage design and the
16 Bayesian framework, we're allowing for more
17 efficient, unbiased treatment effect estimates. We
18 have developed software to disseminate these
19 methods, and we hope that this design will aid in
20 identifying more effective treatments for rare
21 diseases. Next slide.

22 I just want to acknowledge some of the

1 people that helped with this work, some of those
2 pictured in this picture here, and also through
3 contracts with PCORI and also the FDA. I think
4 that's the end of it, so thank you so much, and I'd
5 be happy to take questions later or offline. My
6 email is kidwell@umich.edu.

7 DR. ROSENBLUM: Thank you, Dr. Kidwell.
8 That was a fantastic presentation. There are
9 multiple questions in the Q&A, and we'll come back
10 to them after all the speakers have gone, but you
11 kept your talk perfectly on time as well. Thank
12 you for that.

13 We'll turn next to Noah Simon. Noah is an
14 Associate Professor in the Department of
15 Biostatistics at the University of Washington. He
16 is an investigator for the Therapeutics Development
17 Network at Seattle's Children's Hospital. He has a
18 variety of research expertise areas, including
19 biomarker development; clinical trial design,
20 including adaptive clinical trial designs; and
21 machine learning. He primarily engages with trials
22 in oncology and cystic fibrosis. He's, as all the

1 speakers and panelists today, an outstanding
2 researcher, and I turn it over to you, Noah.

3 **Presentation - Noah Simon**

4 DR. SIMON: Perfect. Thank you so much,
5 Michael. It's a pleasure to be here.

6 I'm going to be talking about adaptive
7 enrichment designs, and we'll get into that in a
8 little bit. I think there are some real challenges
9 in employing those in rare disease settings, but I
10 also think some real opportunities. I like to work
11 on machine learning. We're not going to talk about
12 machine learning here. We're not going to be
13 talking about anything too fancy, I think, because
14 we're going to be very limited in the number of
15 people we can engage within these settings. Next
16 slide.

17 I wanted to give a shout out to a certain
18 Richard Simon, who I've chatted with a lot about
19 these ideas, and that's a fairly recent picture of
20 him. Next slide. I guess I want to say, normally,
21 I love to engage with questions during the talk,
22 but I'm probably not going to be able to do that

1 this time because we're a little time limited, so
2 I'm looking forward to engaging with questions
3 during the Q&A.

4 In many diseases, and in rare diseases a lot
5 as well, rather than engaging with general purpose
6 treatments that target maybe every person with the
7 disease, we often are engaging with very targeted
8 treatments that may only target a subset of people
9 with the disease. I guess something that we might
10 call one disease could actually be a heterogeneous
11 collection with a similar phenotype but different
12 mechanistic causes, and maybe slightly different
13 phenotypes that, again, are very similar.

14 So again, our new treatment might only
15 target one of those mechanistic causes, one type of
16 dysregulation, and often we have some idea of which
17 patients suffer from that dysfunction versus have a
18 disease caused by another, but we don't have a
19 perfect characterization of essentially which
20 subset of patients who have the disease will
21 benefit from our new treatment versus which subset
22 maybe have a slightly different dysregulation

1 before we start our phase 3 trial, before we start
2 our pivotal trial. The idea is when we have that
3 uncertainty, what do we do moving forward, and
4 we'll talk about how adaptive enrichment is one
5 class of designs we might use. Next slide.

6 So what might you do? I guess classically
7 you might say, "Well, we don't really know who's
8 going to benefit from new treatment, so let's just
9 enroll everyone with the disease," or you might
10 say, "Oh, here's our best guess, so rather than
11 just enrolling everyone, let's enroll our best
12 guess of the subset of people who we think will
13 benefit from the disease."

14 So the first I'm going to call an all-comers
15 design, and the second is what's called an
16 enrichment design. Generally, you need some
17 rationale for why you're not going to enroll a
18 subset of patients with the disease, but it's often
19 very hard, as you enter a phase 3 trial, to
20 identify exactly who should be enrolled. Next
21 slide.

22 An adaptive enrichment design ideally

1 provides a happy medium here. You start by
2 enrolling everyone and randomizing them to your new
3 treatment, maybe in standard of care in the case of
4 a randomized trial, and as people progress through
5 the trial, assuming you can measure at least some
6 short-term endpoint, you can start to
7 identify -- based on one or more covariates you
8 have measured, generally, at baseline -- who you
9 think will or is benefiting from treatment and who
10 is not, then you can modify your enrollment
11 criteria as you go, and maybe drop a strata of
12 patients for future enrollment based on what you're
13 learning.

14 As you can imagine, it's hard to learn a lot
15 during a trial, but maybe you can learn a small
16 amount that could still be really effective. So
17 these modifications will use outcomes and treatment
18 assignments from earlier patients, so you obviously
19 have to be careful about blinding, and bias, and
20 such, but there are ways to engage with that. Next
21 slide.

22 Before I jump into that, again, we said you

1 can have an all-comers design, you can have an
2 enrichment design, or maybe you can do this
3 adaptive enrichment design, where, on the fly as
4 the trial goes, you say, "Oh, this trial targets
5 people whose" -- in the oncology example -- "tumors
6 express this protein too much." But what does too
7 much mean?

8 Okay, we're starting to learn as the trial
9 goes along what a reasonable cutpoint for too much
10 versus too little is, and you sort of stop
11 enrolling patients who you think have too little,
12 and allow them to engage with other treatments,
13 again, as the trial progresses. You obviously have
14 to be very careful about statistical conclusions
15 there, but there are a number of frameworks for
16 engaging with that, which we will touch on as we
17 move forward.

18 In oncology -- and this is not a rare
19 disease example, but I want to give this because I
20 think it's a good archetypal example -- there are
21 exciting new immuno-oncology therapies that target
22 basically expression of this PD-1 or its ligand,

1 PD-L1. And again, we basically know that this
2 should work in tumor or tumor microenvironments
3 that have high expression of either PD-1 or its
4 ligand, and we have observed that. We've observed
5 the efficacy of treatment increases as that
6 expression increases, but it may be effective even
7 in relatively low expressors.

8 So the question is, what do you do when you
9 run a trial? It's become increasingly common to
10 engage with trials where, sort of on the fly, we
11 try to identify that cutpoint and initially allow
12 fairly low expressors to join the trial, and slowly
13 cut that off. Next slide please.

14 Another example where it wasn't used, but it
15 could have been used, and I would argue maybe
16 should have, is another cancer treatment.
17 Cetuximab is a treatment widely available and used.
18 It targets the epidermal growth factor receptor
19 protein on cancers, and the original pivotal trial
20 in colorectal cancer was an all-comers design and
21 didn't initially find significance for this drug
22 being effective at treating that cancer. But when

1 they looked back, did a retrospective analysis,
2 they actually identified it wasn't the amount of
3 expression of eGFR in the tumor that was most
4 important, which they hypothesized initially; it
5 was whether or not that was the pathway by which
6 the cancer was acting or not. So there was
7 actually this mutation, this KRAS mutation, and
8 what you needed was for this mutation not to happen
9 in order for cetuximab to work.

10 So again, we were able to look
11 retrospectively, reanalyzing very strong evidence
12 that treatment was effective in KRAS wild-type
13 patients. But maybe things could have been done
14 more cleanly. There was some chance we were going
15 to miss this, so maybe initially we could have
16 said, "Okay. Let's look at eGFR expression and
17 KRAS mutation status when we jump into this trial,
18 and think about modifying enrollment as the trial
19 moves on," based on who appeared to be benefiting
20 from new treatment over standard of care. Next
21 slide.

22 In all of these cases, there was a clear

1 molecular target, but before the trial was run, it
2 was hard to identify exactly who the right subgroup
3 of patients was, either because we weren't sure
4 what the cutpoint was or because we had maybe two
5 covariates that we thought could be involved in who
6 benefited from treatment.

7 So I would say these are prime choices for
8 adaptive enrichment. We don't want a zillion
9 features. We don't want someone to say, "Well, we
10 have expression of 20,000 genes measured on the
11 tumor, and we think any of them could be
12 informative," because you can't really learn that
13 during one trial and evaluate the efficacy of
14 treatment at the same time. You're asking for
15 trouble there, so you want a handful of features
16 that you think have a priori strong scientific
17 relevance, maybe just one, and you need to find the
18 cutpoint, which is pretty common. Next slide
19 please.

20 So we talked about some examples that are
21 not, maybe, so appropriate for a rare disease
22 forum. I also engage with cystic fibrosis, and

1 here I'm going to talk about a sort of example in
2 cystic fibrosis, where one could think about
3 applying these ideas that I think can map to a lot
4 of other problems.

5 Cystic fibrosis, as many of you probably
6 know, is a genetic disease that results from
7 dysfunction of the CFTR gene/protein, and there are
8 a lot of different mutations in the gene that can
9 cause various types of dysfunction that are all
10 termed "cystic fibrosis."

11 There are some of these mutations that are
12 very common, and there are some that are much more
13 rare. Sometimes they mechanistically might have
14 very similar effects, although maybe not identical
15 effects, and in terms of developing treatments,
16 often treatments are developed for those classes of
17 dysfunction, and for common mutations, we can also
18 identify, here's the dysfunction, and we can
19 actually evaluate the effective treatment in people
20 who have that mutation, that dysfunction. For rare
21 variants, it can be much harder. We maybe don't
22 have enough patients to actually just enroll

1 patients with that mutation.

2 So again, there's this new class of
3 treatments, these modulator therapies that provide
4 essentially replacement for those mechanistic
5 dysfunctions that we see, and these modulators have
6 been extremely successful. I mean, this is like a
7 real success story in medicine, in general, and in
8 the rare disease setting. Next slide please.

9 There's growing evidence that these
10 modulators -- and in particular this triple
11 combination therapy called "Trikafta," which is
12 essentially the most recently approved
13 modulator -- in addition to working for particular
14 common variants that were tested in the main
15 pivotal trial for approval, they also work in some
16 of these more rare mutations.

17 Given a rare mutation, you can actually run
18 an in vitro screen to look for how well, say,
19 Trikafta, that we believe it will work. You can
20 basically take cells that shouldn't express CFTR
21 protein. You can treat them so that they actually
22 do express a mutated version of CFTR protein, and

1 then you can treat that with Trikafta, and leave it
2 untreated, and then look at something like
3 conductance -- and conductance gets messed up
4 generally when you have that mutation -- and see if
5 conductance is fixed.

6 This is a screen by which we can say, okay,
7 does treatment work, not in a human, but sort of in
8 this in vitro very sick setting? You've got a
9 continuous measure of activity. There is strong
10 reason to believe that an increased value of that
11 measure should result in an increased clinical
12 outcome. So one question is, how much is enough
13 activity? What should a good cutpoint be?

14 I want to note I'm going to talk about
15 running trials here. There has been some label
16 expansion of Trikafta just based on this, where
17 additional trials didn't need to be run. It was
18 expanded to different mutations beyond the original
19 one, but one could imagine that in a scenario like
20 this, say before a pivotal trial had even been run,
21 one might want to employ an adaptive enrichment
22 design. Next slide please.

1 Again, is it a good fit for adaptive
2 enrichment? It's a potentially very effective
3 therapy but only for a subset of people.
4 Excellent. Are there good alternatives for people
5 to try? Maybe not great, so maybe that's a strike
6 against it. Adaptive enrichment is better when
7 there are good alternatives. Do we have a larger
8 potential pool of patients than we can likely
9 enroll? Are we in the case where we have
10 five patients with the disease in the enrollment
11 window, and we basically need to enroll all of
12 them, and cystic fibrosis is a relatively common
13 rare disease, so we're not.

14 Again, I would argue that it's a pretty good
15 candidate. I generated simulations roughly based
16 on values observed in the pivotal trial of
17 Trikafta. This is in people who had at least one
18 allele of this quite common variant, F508del, where
19 we hypothesized Trikafta should be effective. Next
20 slide please.

21 Here, again in my simulation, I generated a
22 Biomarker X. You can think of this as that

1 conductance measure as uniform between 0 and 1, and
2 I had a treatment effect -- in cystic fibrosis, the
3 outcome is this continuous measure of lung
4 function, which we measure at the beginning of the
5 trial and at the end. I'm going to evaluate
6 average improvement; that's commonly what's
7 evaluated, between the beginning and the end.

8 Here, for standard of care, regardless of
9 biomarker value, I'm imagining you have no average
10 improvement, where for Trikafta, in this case for
11 biomarker values here, I'm saying about 0.5, but I
12 vary this in the simulation setting. I'm imagining
13 you have an average improvement of something like
14 five points on this FEV predicted scale. I'm going
15 to vary, again, where the cutpoint is and the jump
16 just to show what gains we might get from using the
17 adaptive enrichment versus something like an
18 all-comers design here. Next slide please.

19 In these simulations, we have 60 patients
20 who we've randomized 30-30, the new treatment and
21 control, and I've run an adaptive enrichment design
22 where I have two blocks of 30 patients randomized

1 15-15. In the first block, I include everyone. In
2 the second block, I try to identify what that
3 cutpoint is and then restrict enrollment to only
4 those with the biomarker value above that cutpoint,
5 and I use a hypothesis test that combines the
6 p-values from block 1 and block 2, and I give a
7 citation for that at the end. Next slide.

8 What we see here is, to explain this, is we
9 have three panels. On the left, I'm imagining that
10 30 percent of people benefit from new treatment
11 over standard of care, so that cutpoint is actually
12 at 0.7. In the center, 50 percent of people should
13 benefit from new treatment over standard of care,
14 and on the right, 70 percent of people should
15 benefit from new treatment over standard of care.

16 Here, I have the actual effective treatment
17 in those who benefit, between 5 and 10, what the
18 average change is for those patients, and on the
19 Y-axis, I have the power ranging from 0 to 100, and
20 in the dotted line, I have the power of a simple,
21 non-adaptive design, where I just enroll everyone.
22 And here, I have the power of an adaptive

1 enrichment design, where I identify who to enroll
2 after that first block. Maybe I could have also
3 had an enrichment design where I just, a priori,
4 choose a cutpoint, and that probably would have
5 been a reasonable comparator as well.

6 Here, we basically just see, as one could
7 sort of expect here, that we can get a large
8 improvement, especially if only a small subset of
9 patients truly benefit from treatment. We can
10 identify those, and then in that second block of
11 the trial really only enroll those people. So
12 again, in all of these cases, it's a little bit of
13 a straw person scenario. It's set up such that
14 adaptive enrichment will do well, and in fact it
15 does great, but I think this is a pretty realistic
16 scenario we might see in practice. Next slide.

17 My takeaways here, we can have a large
18 improvement in power, and we're likely not
19 perfectly identifying that cutpoint, that
20 threshold, and we're not perfectly saying nobody
21 below this will benefit, but it gives us
22 statistical evidence to justify the use of

1 treatment in people with a large biomarker value.
2 A rare disease setting is a very hard setting to
3 identify whether a treatment is effective or not,
4 so for these very effective treatments, I think
5 identifying that the treatment does work in a
6 subset of patients is a good goal, although, again,
7 there's discussion to be had with the FDA. Next
8 slide please.

9 I'm going to jump into some discussion
10 points, which I will likely run out of time for in
11 a minute or two. But some points; I think we don't
12 want to let the perfect be the enemy of the
13 good, so I've used an adaptive enrichment design
14 formalism that doesn't prespecify subgroups and
15 then testing them. I think that's great if you
16 have a thousand patients. I think in the case that
17 you have 60 patients, you can't really do that.

18 So again, there is a hypothesis test that
19 we're running. We got a p-value, but there's a
20 little bit of subtlety there, and I'm happy to
21 answer questions about that during Q&A, online, or
22 it's in one of the papers I cite. And I added a

1 Yogi Berra quote that I think is definitely not a
2 real quote, but I kind of like it. Next slide
3 please. Again, I think statistics here is meant to
4 support decision making, but it's never going to
5 give us guarantees. Here, we're in the absence of
6 perfect info, so we can't make perfect decisions,
7 but hopefully we can make good ones. Next slide
8 please.

9 In this stereotyping example, it's possible
10 that no clinical trial is needed at all because
11 Trikafta is extremely effective, in general. We
12 have strong evidence that it will be effective in
13 these patients in vitro. In fact, again, there was
14 label expansion based on that without running a
15 trial.

16 That said, you can imagine a slightly
17 different world where we didn't originally have
18 that positive trial. We didn't originally have
19 that common variant that Trikafta was so effective
20 with, and instead, everything was a rare variant;
21 then maybe when we run that first trial, we have to
22 figure out where to threshold that assay. So we

1 have that problem from the beginning, and then
2 adaptive enrichment would be useful. Next slide
3 please.

4 I throw out the number 60 people that may be
5 a pipe dream in some settings. It may not be
6 totally appropriate. You can do this with fewer,
7 and maybe you're combining early- and later-phase
8 data. There is something called a seamless
9 phase 2/phase 3 that one could engage with there.

10 I'm going to cut it there. I have a little
11 bit more on the slides that mention maybe use of
12 registry data or historical data for a control arm
13 and some thoughts on specifics on what null
14 hypothesis is being tested. But again, those are
15 technical details. [Go through next several slides
16 and finish on last slide.] On my last slide, I
17 think I have citations, so there's some discussion
18 here and papers that I did with Richard Simon. So
19 anyhow, I look forward to answering questions.
20 Thank you all for your time.

21 DR. ROSENBLUM: Thank you very much, Noah,
22 for that outstanding presentation. I liked

1 especially the Yogi Berra pretend quote, but very
2 good. We will address questions at the end, and
3 we'll have at least one FDA panelist respond to
4 questions.

5 But at this point, we're honored to have
6 Dr. Nigel Stallard. He's a Professor of Medical
7 Statistics and Deputy Director of the Clinical
8 Trials Unit at Warwick Medical School in the UK.
9 He's an Editor in Chief of the Journal Statistics
10 in Medicine. He has a wide range of research
11 expertise, including statistical design and
12 analysis of clinical trials. In particular, he's
13 worked on optimal design for clinical trials and
14 rare diseases in small populations, and on
15 methodology for trials with interim analyses and
16 adaptations, such as treatment for subgroup
17 selection.

18 We're lucky to have Dr. Stallard here
19 presenting, and I will turn it over to you.

20 **Presentation - Nigel Stallard**

21 DR. STALLARD: Thank you, Michael, for the
22 introduction, and thank you for the opportunity to

1 speak. I was going to say this afternoon, but it's
2 the afternoon here but probably the morning for
3 most of you.

4 This talk does fit with the general theme
5 for today of design and analysis methods for
6 clinical trials for rare diseases, but although I
7 am interested in adaptive designs, I'm not
8 particularly going to be talking about adaptive
9 designs today. In fact, this will be less of a
10 talk about the methodological details or about a
11 particular approach, and more of an overview, both
12 an overview of current practice, and also thinking
13 about what we might do differently in terms of
14 methodology for rare disease clinical trials. I
15 guess I mean different in two ways, different to
16 what we're doing currently and also different to
17 what we might do in non-rare disease settings.

18 Next slide please.

19 Let's just start with some acknowledgements.
20 Most of the work that I'm talking about today has
21 really risen from two projects on clinical trials
22 in rare diseases or small populations. The first

1 is a project that was funded by the EU that I led,
2 called InSPiRe. It stands for innovative
3 methodology for small populations research, and the
4 second was a task force on small population
5 clinical trials, organized by IRDiRC, the
6 International Rare Disease Research Consortium. In
7 addition to acknowledging the EU funding, I should
8 acknowledge the input from a number of colleagues
9 on these two projects. I won't list them all, but
10 there were numerous comments, including Simon Day
11 and Tim Friede, whose names appear at the bottom
12 there. Nevertheless, I should say, of course, the
13 views that I'm expressing are my own and not
14 necessarily theirs or anyone else's. Next slide
15 please.

16 I want to step back from some of the details
17 of a particular trial design and really think about
18 what it is that we're trying to do when we do
19 clinical trials in rare diseases, so I'm going to
20 start off with these three quotes from our
21 regulatory guidance that many of you may have seen.

22 The first two come from the EMA guidance,

1 which, of course, applies in the European community
2 and also is now adopted by the UK. There are two
3 quotes here. The first says, "Patients with rare
4 conditions deserve the same quality, safety, and
5 efficacy in medicinal products as other patients,"
6 and then they go on from that to say, "Orphan
7 products should therefore be submitted to the
8 normal evaluation process." There's a similar
9 statement from the FDA and the U.S. saying "The
10 Orphan Drug Act does not create a statutory
11 standard different from common conditions."

12 The title of my talk asks should we be doing
13 clinical trials in rare diseases differently, and
14 that means differently to what we do in non-rare
15 disease settings, then these statements at face
16 value would suggest that the answer to that is no.
17 Nevertheless, of course, things are not quite as
18 simple as that, or we wouldn't be needing this
19 workshop today. So I want to start by looking at
20 what's actually being done in the world of rare
21 diseases. Next slide please.

22 This slide presents some work that was done

1 by a group in Liverpool, and what they did was they
2 went through the clinicaltrials.gov database,
3 looking at trials in rare diseases and non-rare
4 diseases, and comparing the recorded sample sizes
5 for those trials. So this slide is just a summary
6 of what they found. The first column gives the
7 ranges of trial sizes, naught to 50 patients, 51 to
8 100 patients, and so on; then the second and third
9 columns give the percentages of trials in each of
10 those size ranges, the first one for non-rare
11 diseases, and the second one for rare diseases.

12 So looking first at the non-rare diseases,
13 you can see many of the trials, even in non-rare
14 diseases, are quite small, with fewer than
15 50 patients, but there's quite a sizeable chunk,
16 around 30 percent of trials, with between 100 and
17 500 patients. I think that's slightly bimodal, if
18 you like distribution, and probably reflects a mix
19 of trial types. What they looked at was all of the
20 trials on clinicaltrials.gov. That's represented
21 by early phase and confirmatory late-phase trials,
22 so perhaps that's why we have a bunch of small

1 trials, and then a bunch of larger trials.

2 If we look at the rare diseases, then
3 perhaps, not surprisingly, we see something rather
4 different, and here it seems, as we probably would
5 have expected, the larger trials are much less
6 common. So here, the large majority of trials have
7 50 patients or fewer, and really, there's no
8 evidence of that second mode in the distribution,
9 if you like, of trials with more than 100 patients.
10 Next slide please.

11 Here's a summary of similar work, which
12 focuses particularly on rare diseases and splits up
13 for the phase 2 and phase 3 trials so we can look
14 at those separately, and also splits up diseases by
15 the disease prevalence because, of course, a rare
16 disease, which affects between 1 and 5 in 10,000
17 people, while still rare, is obviously very
18 different from one which affects perhaps one or two
19 in a million.

20 The left-hand panel shows phase 2 trials,
21 and the right-hand panel here shows phase 3 trials.
22 The sample sizes are on the Y-axis, and you can see

1 that this is on a logarithmic scale, and it does go
2 right down to 1 at the bottom there. In each of
3 the disease prevalence groups, we've got two sets
4 of bars. The first one is for actual trial sizes,
5 where the trial's been completed, and the second
6 one is for target trial sizes, where the trial is
7 still ongoing. Of course, quite often, the actual
8 trial sample size is rather smaller than the
9 target.

10 Looking first at phase 2 on the left here,
11 apart from the very, very rarest of the diseases,
12 the trial sample size doesn't seem to change that
13 much across the different prevalence groups, and
14 here, the typical sample sizes are just under 50,
15 so there may be kind of 30 or 50 patients in each
16 trial. So clearly that appears to be achievable
17 even in these rare diseases.

18 If we move across and look at the picture on
19 the right, here the story is rather different,
20 obviously, because these are phase 3 trials, these
21 are confirmatory trials. Trials typically in this
22 setting are rather larger, and it certainly looks

1 as we consider more rare diseases, the trials are
2 smaller than for the less rare of the rare
3 diseases, and we can see it's something of a trend
4 with larger trials on the right and smaller trials
5 on left within that panel. In the very rarest of
6 diseases, those affecting fewer than 1 in a
7 million, really, phase 3 just appears to be
8 completely absent in this picture. There are very,
9 very few trials which describe themselves as
10 phase 3 that were done in these very rare diseases.

11 Now, obviously the designation as phase 2 or
12 phase 3 was taken directly from entries on the
13 clinicaltrials.gov database, but it really appears
14 that, perhaps unsurprisingly, whatever the
15 regulations might say, trials are being done
16 differently in rare diseases to more common ones.
17 Trials in rare diseases are smaller, and for rarer
18 the disease, definitely [indiscernible] the trials.
19 And in particular, it looks like in phase 3, in the
20 very rarest of diseases, these might be skipped
21 altogether. Next slide please. But clearly we are
22 doing trials differently in rare to non-rare ones.

1 This is the well-known problem, and trial sample
2 sizes are small, often necessarily so, yet we want
3 as much high-quality data as possible for clinical
4 and regulatory decision making.

5 So how might we do them differently? How
6 might we do them better? I'm going to talk briefly
7 about three possible general approaches. The first
8 is to get more data. Assuming it's impossible to
9 increase the sample size, we nevertheless obtain
10 more data for our analyses and our decision making
11 somehow from the patients that we've got in our
12 trial.

13 Secondly, can we get more information for
14 our decision making from the same data? So use our
15 data as efficiently as possible. Then
16 thirdly -- and this isn't really talking about
17 doing trials differently, but more about the way in
18 which we use the information, and using that
19 differently -- can we consider changing the level
20 of evidence required for our decision making? Next
21 slide please.

22 So first of all, let's think about how we

1 might try to get more data, and first from within
2 the trial itself. So without increasing the sample
3 size, this means collecting more data from the same
4 patients, and sometimes we can do this by thinking
5 about the data that we collect. So we can ensure,
6 for example, that we collect continuous endpoints
7 where we can, rather than binary endpoints, and
8 certainly not dichotomize a continuous one into a
9 binary response. We can make sure we collect
10 relevant baseline covariate data, and we can
11 collect longitudinal data where we can and have a
12 long-term follow-up, if possible, and we can
13 collect secondary endpoints that can be used to aid
14 our decision making.

15 Now sometimes there are good clinical
16 reasons why we shouldn't do these things, but,
17 generally, if we can, then if we can collect and
18 use more data, then that can only be good and can
19 only help us in our decision making. Next slide
20 please.

21 So those are pretty much no-brainers, I
22 think, but slightly more controversial and

1 certainly requiring some more careful thought is
2 the use of additional data from outside of the
3 trials, and this may be from historical control
4 data, so taking the control data from previous
5 trials in the same population and using that either
6 to completely replace the control group in the
7 current trial or perhaps more likely to augment it
8 so we can use a smaller control group in our trial.

9 There's been quite a lot of recent
10 statistical work or methods which enable us to do
11 this. In particular, there's been a lot of work on
12 what are called dynamic borrowing methods, where
13 external control data were weighted according to
14 their concordance or discordance with the observed
15 data; so the idea being that if the historical
16 controls look similar to the current controls, then
17 we should include them, and if they look rather
18 different, then they could be ignored.

19 I think there's lots of nice work there, and
20 it's a really nice idea. The challenge is, if you
21 think about the decision as essentially being a
22 hypothesis test and about the type 1 error rate for

1 that hypothesis testing, the probability of a false
2 positive result, taken over all the possible sets
3 of data from the trial, with the historical control
4 data considered fixed, then by including that
5 additional control data, you can inflate the
6 conventional type 1 error rate. Now, there are
7 ways around that. You can make the tests more
8 stringent to bring the type 1 error rate down
9 again, but generally that removes the very
10 advantage of including the external data at all.

11 Now, obviously, hypothesis testing is only
12 one way to think about the decision making process,
13 so how much you worry about type 1 error rates
14 really depends on what it is that you think you're
15 doing in the trial. But if you are going to use
16 historical control data in this way, then I think
17 it's important to at least consider this. Maybe
18 it's something that could be discussed in the
19 discussion session later on.

20 Extending the use of external data further,
21 there was quite a lot of talk in the session
22 yesterday, of course, about using real-world

1 evidence with a whole range of methods possible,
2 including the generation of synthetic controls or
3 even conducted so-called in-silico trials, using
4 models based on real-world data. Again, I think
5 this probably comes with many of the challenges of
6 historical controls, and perhaps more. It does
7 have real potential, perhaps particularly for
8 exploratory work, or for planning trials, or using
9 alongside trials to interpret the results when the
10 samples are small, but I think it's something,
11 again, that needs to be very carefully thought
12 about. Next slide please.

13 So having collected that data, we obviously
14 want to make the best use of it possible, which
15 really makes sure that our analyses are as
16 efficient as possible. We're not going to say very
17 much about this because I know it's the theme of
18 some of the later talks today.

19 We also want to make sure, of course, that
20 our designs are as efficient as possible, which
21 we've heard in the last couple of talks; so using
22 methods like group-sequential approaches to stop as

1 soon as we have sufficient evidence to reach a
2 conclusion; or adapting our design to remain
3 efficient as we learn about treatment effects, or
4 about use of parameters, or about patient
5 heterogeneity; or going back to something more
6 similar to what we heard in the first talk, designs
7 which allow us to have patients receiving multiple
8 treatments, so they act as their own controls and,
9 obviously, there are a number of methods enabling
10 us to do things like this. Next slide please.

11 The final thing I want to talk about is
12 really thinking about what level of evidence is
13 required for decision making. Of course, when we
14 conventionally have a confirmatory trial in a
15 non-rare disease setting, then the focus is
16 primarily on type 1 error rates.

17 Now, I know the p-value isn't everything,
18 but in practice, much of our demonstration of
19 efficacy comes down to whether or not we have a
20 conventionally statistically significant result.
21 So when we plan our study, we typically fix the
22 type 1 error rate, and then find the sample size to

1 give us a certain power to detect a specified
2 effect. And when we analyze our study, we
3 typically control the type 1 error rate, and if the
4 trial is smaller than we'd hoped, then we accept it
5 can be underpowered.

6 But why are we really thinking about error
7 rates so much at all? Of course, the answer is
8 because we're worried about the consequences of an
9 incorrect result. In particular, our focus
10 specifically on the type 1 error rate is because
11 we're usually more concerned about protecting
12 ourselves and the patient population against false
13 positives and against false negatives. So maybe it
14 makes sense to focus explicitly on those
15 consequences and to think about the decisions that
16 we might make either in terms of patient outcomes
17 or in terms of health economic benefits to make
18 those decisions as well as we can. Next slide
19 please.

20 The health economic approach of value of
21 information analysis allows us to do this.
22 Essentially, in this setting we have a trade-off

1 between larger and smaller trials. If we do a
2 larger trial, then that will obviously give us more
3 information. It will allow us to better control
4 our error rates, and we can maintain our alpha as a
5 conventional level and ensure good power. But the
6 consequence of that is that the trial will take
7 longer, of course, than a smaller one, and that's a
8 particular challenge in a rare disease because it
9 might be a lot longer. And the longer the trial
10 goes on for, the longer it will be before the
11 patients benefit from the results of that trial.

12 So we're trading off getting better evidence
13 to make a good decision against having more people
14 more quickly benefiting from that decision, and
15 that balance changes as the number of potential
16 patients in the population changes. So it's
17 different for a rare disease than what it will be
18 for a more common one. And as the population gets
19 smaller, so it becomes optimal to use a smaller
20 size trial, and to achieve this, to use larger type
21 error rates than conventionally used.

22 As we see, this might be happening anyway.

1 Trials are smaller, and some of the methods used
2 probably don't control the type 1 error rates at
3 conventional levels. So maybe we should consider
4 this more explicitly to enable us to formalize
5 these things rather than making these rather ad hoc
6 decisions. Next slide please.

7 The final slide, I'll just end with another
8 quote from the EMA guideline here that says, "No
9 methods exist relevant to small studies that are
10 not also applicable to larger studies." This is
11 kind of true, of course. In many respects, the
12 methodology for clinical trials in rare diseases is
13 not that different to that for other diseases, but
14 in rare diseases in small populations, the
15 challenge is often greater as we seek to base
16 decisions on small samples.

17 So we somehow need to be more efficient,
18 faster, and smarter, so we need to make sure that
19 we do consider all information as much as we can,
20 and that our designs are as efficient as possible,
21 and that our decision making reflects the patient
22 populations to try to ensure that they benefit from

1 the clinical trials as much as they can. Thank
2 you.

3 DR. ROSENBLUM: Thank you, Dr. Stallard, for
4 the excellent talk, and I want to thank all three
5 speakers, Kelley Kidwell, Noah Simon, and Nigel
6 Stallard, for being crystal clear in presentations
7 of somewhat complicated topics. You've done very
8 well on that, and I appreciate it.

9 We'll turn next to panelist, Greg Levin.
10 Greg will give some feedback, based on the talks,
11 and then we'll go to answer questions with the
12 remaining time.

13 Dr. Levin is the Associate Director for
14 Statistical Science and Policy in the Office of
15 Biostatistics in the FDA's Center for Drug
16 Evaluation and Research, CDER. He has experience
17 supporting drug review across a wide range of
18 therapeutic areas and has represented CDER on
19 several policy and guidance working groups,
20 including efforts related to adaptive design,
21 master protocols, benefit-risk, and the evaluation
22 of effectiveness, and it's my pleasure to turn it

1 over to Greg.

2 **Summary of Presentations - Gregory Levin**

3 DR. LEVIN: Thanks, Michael, for the
4 introduction, and I really appreciate the
5 opportunity to participate today. I really enjoyed
6 the three talks, and thought I would share a few
7 brief thoughts on each one, and then make a few
8 additional remarks related to adaptive designs in
9 rare disease settings.

10 First, Dr. Kidwell gave an interesting talk
11 about SMART designs. I like the ability to
12 incorporate dose finding, which is often
13 inappropriately skipped in rare disease settings.
14 I also think there can be value when incorporating
15 a second stage with re-randomizations, and this
16 will often be better than simply rolling all
17 subjects over into an open-label and controlled
18 extension phase, which is something we often see.
19 The re-randomization can provide data on relevant
20 secondary clinical questions, things like she
21 discussed, for example, to evaluate whether there's
22 benefit in uptitrating to a high dose in subjects

1 you don't approve on a low dose.

2 That being said, I also think there are some
3 limitations to consider. Separate analyses of the
4 second stage to answer secondary questions, that's
5 always a good thing, but if the primary efficacy
6 analysis seeks to formally incorporate the
7 second-stage data, then we need to rely on fairly
8 strong assumptions about the stability of the
9 disease, and the lack of confounding, and the lack
10 of carryover effects.

11 The potential borrowing of external control
12 data that was discussed also adds additional risks
13 and complexities. These may be able to be
14 adequately addressed with careful approaches and in
15 certain settings, but there are challenges there.
16 I think the recent FDA guidance on
17 externally-controlled trials gets into a lot of
18 that.

19 In the second talk, Dr. Simon gave a nice
20 summary of adaptive enrichment designs, and I think
21 such an approach can definitely have value in
22 settings where there is a targeted treatment with a

1 fairly well understood mechanism of action that's
2 thought to be more likely to benefit a certain
3 subpopulation of the disease. I agree with the
4 recommendation that there should ideally be one
5 feature with strong a priori evidence considered
6 for adaptive enrichment.

7 Unfortunately, such evidence can be hard to
8 come by in the rare disease setting, and if it's
9 not available, it may be better to enroll and
10 obtain data on the full population, and then just
11 explore the relationship between the biomarker and
12 the treatment effect.

13 One additional comment I would make is that
14 the approach may not be as advantageous if the true
15 relationship between the biomarker and the
16 treatment effect is, say, more continuous in nature
17 rather than a step function, and this might be a
18 bit more realistic. And finally, I'd also be
19 interested in learning a bit more about approaches
20 to not just control type 1 error but to provide
21 reliable estimates of treatment effects in the
22 targeted subpopulation.

1 In terms of the third talk, I very much
2 agree with Dr. Stallard's recommendations to
3 maximize efficiency with simple approaches that are
4 often underutilized, things like analyzing
5 continuous endpoints and using all of the
6 information rather than dichotomizing; adjusting
7 for prognostic baseline covariates; and collecting
8 longitudinal data.

9 With respect to longitudinal data, I've seen
10 multiple trials in rare disease settings where a
11 short-term control period was carried out. They
12 didn't see evidence of a treatment effect, and then
13 there were post hoc hypotheses that the controlled
14 period was not long enough for the treatment effect
15 to manifest. I know this is challenging, but if
16 there's uncertainty at the design stage about the
17 duration of treatment that may be necessary to
18 provide benefit, having a longer controlled period
19 is really critical to increase the chances of
20 establishing benefit and making the effect of a
21 treatment available to patients.

22 I also think something like the information

1 approach to sample size determination that
2 Dr. Stallard discussed can be valuable, and I do
3 want to note that the 2019 FDA Effectiveness Draft
4 Guidance states that in certain settings where
5 flexibility is warranted, like in ultra rare
6 disease where sample sizes are limited, a higher
7 significance level than the typical 0.05 standard
8 may be acceptable if it's prespecified, justified,
9 and agreed upon with the agency. But we definitely
10 have more work to do on a framework, and factors to
11 consider, and trade-offs to consider in making such
12 determinations.

13 Finally, I'll just add one comment of my
14 own. The type of adaptive design in the rare
15 disease setting that I think is often overlooked
16 and should be considered nearly all the time, is
17 actually the simplest one, and that's the use of a
18 group-sequential design with multiple interim
19 analyses to potentially stop the trial early for
20 efficacy. Trials in rare diseases are often
21 planned with tremendous uncertainty about the
22 potential effect size, and the choice of the

1 alternative hypothesis that determines the sample
2 size is often pretty arbitrary and overly
3 optimistic, and the trial with a single analysis on
4 a fixed number of subjects may be substantially
5 underpowered to detect smaller effects, but effects
6 that would still be plausible and still be
7 clinically meaningful.

8 So to address this, a trial can be designed
9 that has interim analyses with prespecified
10 stopping rules, and this can lead to a high
11 probability of stopping for efficacy with smaller
12 sample sizes if the treatment truly has a large
13 effect, but it can also allow the trial to proceed
14 to larger sample sizes and ensure reasonable power
15 if the treatment truly has a more moderate effect
16 size. Such an approach can also incorporate
17 futility stopping so that more resources are
18 invested only if the interim results are promising.
19 So that's something I think should definitely be
20 considered more often. But I'll stop there, and
21 thanks again for the opportunity to share some
22 thoughts.

Q&A

1
2 DR. ROSENBLUM: Thank you, Dr. Levin. That
3 answered many of the questions that I had and some
4 of the participants had put into the Q&A and have
5 submitted beforehand.

6 First, I'd like to thank all the speakers
7 and panelist, Dr. Levin. We have 10 minutes, if
8 I'm not mistaken, to answer questions that have
9 been submitted, so if all the speakers open your
10 video. Great. Fantastic. I'll throw out
11 questions, but feel free to jump in, anyone, with
12 answers.

13 One of the questions that came up most
14 often -- and this is in more than 100 submitted
15 questions prior to the workshop -- is when are
16 single-arm studies appropriate for regulatory
17 decision making, specifically approval in rare
18 diseases? Actually, one of the live attendees
19 submitted this as well.

20 It's a tough question. That points towards
21 Greg, and it's, in some ways, not a fair question
22 because it's hard to say anything with generality.

1 But nonetheless, I still wanted to toss that your
2 way, but feel free to decline, of course, but that
3 does seem to be on many people's minds, at least
4 based on the questions that were submitted.

5 DR. LEVIN: Thanks, Michael. Yes, it's a
6 difficult question. I guess I would point people
7 towards the recent FDA draft guidance on
8 externally-controlled trials, which typically have
9 a single investigational arm, and then there is
10 some sort of comparison, whether it's to
11 patient-level external data or whether you're in a
12 setting, where there's understanding of the disease
13 process such that there's knowledge that the
14 outcome would not improve in the absence of an
15 effective treatment, and whether that's an oncology
16 situation where it's known that the tumor would not
17 shrink, and tumor shrinkage by a certain magnitude
18 would be considered meaningful.

19 I think it's a very complex set of factors,
20 that I would point to that guidance and some of the
21 considerations there about both the settings, maybe
22 more fit for purpose, and also some of the

1 approaches that would be necessary to try to
2 mitigate, to the extent possible, the biases that
3 can be introduced with non-randomized comparisons.

4 DR. ROSENBLUM: Thank you. That's a very
5 clear response. I haven't read that draft
6 guidance, so I'm going to read it myself after
7 this.

8 Any thoughts from the three speakers? I
9 mean, it's hard to comment on regulatory --

10 DR. SIMON: I can't comment on the
11 regulatory.

12 DR. ROSENBLUM: Just on your thoughts, in
13 general, on the evidence produced by them.

14 DR. SIMON: From what I've seen, one needs
15 extremely good historical data, and you need to be
16 very, very careful to make sure that things are
17 appropriately matched or that statistical methods
18 are in place to account for the fact that there may
19 not be a perfect match. I love statistical
20 methods, and I am very cautious when they say, "Oh,
21 just do an analytic correction for the fact that
22 you don't have matching populations." That rings

1 alarm bells.

2 I also want to say the perfect is the enemy
3 of the good here, I think, and it's an incredibly
4 challenging problem. I think sometimes we need to
5 be a little bit more aggressive, and noting that
6 there are type 1 and type 2 errors that we have to
7 be concerned with.

8 DR. ROSENBLUM: Those are great points.

9 Dr. Stallard or Dr. Kidwell, any thoughts?
10 You don't need to.

11 DR. STALLARD: Not really. I don't think
12 anything about what Noah just said. I think it is
13 something which you need to be very sure that you
14 really understand what's going on from your
15 historical or external data and have very good
16 confidence that that really is comparable.

17 DR. ROSENBLUM: Yes. That's quite
18 reasonable as well.

19 Good. Thank you. We'll move on to the next
20 question. The second most common question that
21 came up was about sample size, choosing one sample
22 size, given limited information. I liked what

1 Dr. Levin proposed about doing group-sequential
2 design to try to handle the large uncertainty about
3 effect size, and also I liked Dr. Stallard's
4 approach for potentially inflating alpha and
5 looking at decision theoretic criteria to decide on
6 the study design and sample size.

7 Dr. Kidwell, do you have software for sample
8 size selection? Do you want to talk a little bit
9 about that?

10 DR. KIDWELL: Sure.

11 DR. ROSENBLUM: It's an opportunity, and any
12 others who want to talk about the sample size
13 question, that also I think is on many people's
14 minds.

15 DR. KIDWELL: Sure. Yes, sample size, we
16 all have that question when we think about clinical
17 trials, and especially in rare diseases when we
18 know that the numbers are so limited. We have
19 Rshiny applets that are available via if you search
20 my name, I think. I guess one is in the draft
21 stage. It should be out very soon. They ask for
22 information that we think statisticians, along with

1 clinicians, could easily, or somewhat putting their
2 heads together, come up with and put in there, any
3 try to provide some guidelines; mainly, expected
4 treatment effects.

5 We ask some information about linking the
6 data from stage 1 to stage 2, and then what's the
7 probability, sort of asking about power and type 1
8 error, but since we're in a Bayesian framework, we
9 phrase it slightly differently. But it's very
10 similar in terms of what power do you want, and how
11 much are you willing to allow for an incorrect
12 conclusion.

13 So we hope that that helps facilitate the
14 use of these snSMART designs, as it can be
15 intimidating, if you're not used to the Bayesian
16 framework, to think about sample size and/or how to
17 use these. So we've created both, on the front
18 end, design software, and then also an R package on
19 the backend to actually analyze the data to help
20 with the whole process.

21 DR. ROSENBLUM: That's great. Thank you.

22 Any other thoughts on the sample size

1 question?

2 DR. STALLARD: First of all, really a great
3 report in what Gregory said about the use of
4 group-sequential designs here and allowing stopping
5 as early as possible. I think there's also a
6 question in the Q&A specifically about using
7 Bayesian methods in group-sequential design
8 whenever that's possible. And the answer is yes;
9 there are quite well-established Bayesian
10 sequential analysis methods only if they're
11 stopping in terms of the amount of posterior
12 information or controlling frequentist type error
13 rates using a Bayesian stopping rule. So yes, if
14 you want to be Bayesian, you can also still have
15 rules which allow you to stop early.

16 DR. ROSENBLUM: Great. Thank you.

17 DR. SIMON: I'm a naïve guy, so I'm going to
18 make a kind of naïve comment here. I think if you
19 have a blockbuster, you don't need a big sample
20 size, and if you have something that's pretty
21 incremental, at least my feeling -- from engaging
22 with some of this stuff, and other people may feel

1 totally differently -- you may never have the
2 sample size in a very rare disease if it's a really
3 small improvement to identify, where in oncology,
4 you could make those incremental advances because
5 you can enroll a lot of people in your trial.

6 Again, I definitely agree with Greg's
7 statement. If you have a blockbuster, you can run
8 a group-sequential trial and maybe identify it very
9 quickly, and/or maybe say, "Ooh. We thought it was
10 a blockbuster, but it's not." I know, of course,
11 everyone wants a treatment that's a blockbuster,
12 and we don't always have it.

13 I like adaptive enrichment because maybe
14 it's a blockbuster in some subset of people. Now,
15 you don't want to fish for that subset; you want to
16 maybe be able to identify it beforehand, but I
17 think there's obviously value in very carefully
18 identifying what the sample size should be, but it
19 seems like you do need a lot of flexibility there,
20 and maybe to go in noting that you're likely going
21 to only find blockbusters in some of these cases.

22 DR. LEVIN: This is Greg. I just want to

1 follow up. I agree with Noah. Unfortunately,
2 drugs that have very small incremental
3 improvements, it's probably going to be challenging
4 to identify treatment effects in rare disease
5 settings.

6 I do think there's kind of a middle category
7 of non-blockbusters that have moderate effects. A
8 lot of times I see hypotheses going into trials,
9 but it is a blockbuster. There's always the
10 optimism that it's a blockbuster, and that leads to
11 a fixed sample size that will be large enough for
12 power for a blockbuster effect size, but would be
13 substantially underpowered for a more moderate but
14 very meaningful and still plausible effect size.
15 And that's where I think allowing the trial to go
16 to a larger sample size, if results are promising
17 but not persuasive, that an interim analysis can be
18 valuable, perhaps in combination with something
19 like an enrichment approach, if there's a targeted
20 subpopulation that's realistic and there's evidence
21 to support.

22 DR. ROSENBLUM: Fantastic.

1 We'll stop here, and I want to thank, again,
2 all of the speakers and panelists. I learned a
3 lot. As a participant, if you enjoyed these
4 presentations, which I'm sure you did, please come
5 back in 10 minutes. We have our second session of
6 the day, where the focus is on analysis methods for
7 small populations.

8 Thank you to all the speakers, and we'll
9 return in 10 minutes.

10 (Whereupon, at 10:26 a.m., a recess was
11 taken, and workshop resumed at 10:36 a.m.)

12 **Session 2 - Michael Rosenblum**

13 DR. ROSENBLUM: Welcome back, everyone, from
14 the break. We'll now start Session 2 of today's
15 symposium. The focus is on Analysis Methods in
16 Small Populations, and we'll be discussing
17 different analysis methods and also differences in
18 the target of inference, also called the estimand,
19 the assumptions required when you have small
20 populations, and also performance of different
21 analysis methods.

22 We'll have three panelists and a discussant,

1 the same structure as earlier this morning. Each
2 panelist will be for 20 minutes, followed by some
3 discussions of feedback by the panelists, and then
4 we'll have Q&A, and please put all your questions
5 in the Q&A. Let's get started.

6 We're honored to have Dr. Karen Price as our
7 first presenter. Karen Price is Associate Vice
8 President and Statistical Officer at the
9 Statistical Innovation Center, which focuses on
10 innovative design and analysis of clinical trials.
11 This is at Eli Lilly and Company. She has a wide
12 range of research interests and expertise,
13 including Bayesian methods, innovative clinical
14 trial design and analysis, and quantitative
15 decision making, and I'm excited to pass it to you,
16 Dr. Price.

17 **Presentation - Karen Price**

18 DR. K. PRICE: Thank you so much, and it
19 really is an honor to be here to give this
20 presentation. It's been such a great series of
21 sessions in all of the presentations. I've learned
22 a lot, and I'm just honored to be here.

1 What I'm going to do is give an overview of
2 Bayesian approaches and master protocols, if time
3 on the master protocol front. I think it's great.
4 A lot of what I'm going to talk about, especially
5 up front, was touched on in the earlier session, so
6 I think, really building on one another, there may
7 be points that I will just re-emphasize and/or go
8 into in just a little bit more detail.

9 On the next slide, before I get going, I
10 wanted to give some acknowledgements to several of
11 my colleagues who have been involved in much of
12 this work over the years and helped provide slides
13 and/or were involved in the design in some of the
14 trials I'll talk about.

15 So in the next slide, what I will do is give
16 a super quick overview of rare and pediatric
17 diseases just to really touch on and motivate the
18 Bayesian framework, very much in line with how
19 Dr. Kidwell did earlier. Then I want to go into
20 motivating a Bayesian framework, but probably spend
21 the majority of the time on the third item,
22 showcasing some examples of Bayesian applications,

1 and then if time permits, conclude and touch on
2 master protocols.

3 From the next slide, again not needed really
4 for this audience, but on these next two slides,
5 obviously the key here and what is motivating the
6 use of a Bayesian approach in this instance is that
7 we are dealing with low prevalence of these
8 diseases. On the next slide, I think it's
9 important, of course, to note that it's also a very
10 vulnerable group of patients. Many of these are
11 children, and of course we have a very low number
12 of approved treatments for many of these diseases.

13 So we're definitely motivated. It's
14 incumbent on us to use all that we can from this
15 data to really squeeze everything that we can from
16 this really valuable data, whether it's external or
17 internal to an ongoing trial, so that we can make
18 better decisions and get these compounds to these
19 patients as quickly as possible.

20 So with that motivation, I wanted to talk
21 about, then, the Bayesian framework, so maybe a
22 couple of slides. I like to motivate Bayesian

1 thinking in a couple of different ways. I think
2 there's been a lot that's been discussed about the
3 fact that humans do struggle with prediction and
4 uncertainty. We do have a tendency to over-index
5 information, and we really need in place
6 quantitative approaches to help us frame the
7 available data, especially when it's diverse and
8 coming from multiple sources.

9 We can think about a Bayesian approach, as
10 is depicted on the right side, and you saw a
11 similar thing, again, in Dr. Kidwell's discussion
12 earlier, that this is very much about a continual
13 learning process. This is a very natural way of
14 thinking. We do this all of the time in our
15 decisions throughout the day as we utilize our
16 previous experience, but when we move into business
17 and scientific decisions, the information we have
18 is in the form of data, so we need a platform to
19 synthesize that information. Bayesian methods will
20 provide that so we have what we knew, and that's
21 the prior distribution; what we see, and that's our
22 likelihood; and then what we now know is our

1 posterior distribution, and that's going to
2 continually be updated.

3 On the next slide, we can get a little bit
4 more motivation on this as we talked about, and
5 you'll hear throughout my discussion today it's
6 very much about iterating upon the science and
7 allowing a continual learning process. Today's
8 posterior is tomorrow's prior, and when the facts
9 change, I change my mind. So this is, again, a
10 very natural way of thinking and updating the
11 science. It provides rigorous integration of what
12 we know already within the analysis of new data so
13 that we can shed light on what we don't know.

14 I think one of the things I really wanted to
15 emphasize here is the transparent nature of a
16 Bayesian approach. Whenever we look at data and
17 we're analyzing it, we do bring to the table our
18 previous experience, and that is weighing into how
19 we are thinking about that specific decision. But
20 a lot of that then is done without awareness of
21 others who are also viewing that data, so a
22 Bayesian approach can allow for that transparent

1 understanding of how other data sources are being
2 brought to bear in various decisions, and can
3 improve and allow for more efficient decision
4 making.

5 So it's a very transparent approach. It
6 does allow for straightforward statements of
7 probability and uncertainty, so I'll talk in a
8 little bit about, again, a Bayesian interpretation
9 is very straightforward and typically what people
10 would, I think, prefer, and wish that they could
11 interpret a p-value that way. A Bayesian design
12 can help reduce sample sizes or study durations.
13 Additionally, there is a tremendous flexibility
14 through hierarchical modeling, and now with
15 computational conveniences, we can fit a wide range
16 of models and synthesize information in ways that
17 we couldn't historically.

18 So with that, on the next slide, as I've
19 mentioned already, I think bringing a Bayesian
20 approach to bear here is very important in this
21 rare disease setting, where we do have the small
22 sample sizes, limited data, and few treatments,

1 that a Bayesian approach is going to allow us do
2 more borrowing of information and increase
3 precision, so again, it's providing that mechanism
4 for us.

5 With that motivation, I wanted to move into
6 the next slide. One more slide. Going into a
7 little more detail. Again, some of this you heard
8 in the earlier session, so let me maybe just talk
9 about it in a little bit more detail, and I wanted
10 to show some examples because I think it's
11 important leaving here understanding that it isn't
12 a scary thing, that it isn't a black box, but you
13 can have an understanding of how does this
14 borrowing impact inferences.

15 Borrowing approaches, as you heard, there
16 are two main types, a static type of borrowing as
17 well as dynamic borrowing. Some examples of static
18 borrowing would include pooling, single-arm trials,
19 and also power priors. Dynamic borrowing examples
20 include hierarchical modeling, mixture priors,
21 commensurate priors. There is an appeal, as was
22 noted as well in the earlier session, to dynamic

1 borrowing, at least bringing that to bear, as it
2 can borrow more when the current data are more
3 similar to the historical data and can help protect
4 against over-borrowing and some of the errors that
5 were discussed in the earlier session.

6 On the next slide, as we're going to look at
7 a couple of borrowing approaches, thinking about
8 what some data sources are, I always like to
9 mention expert and caregiver opinion in the rare
10 disease setting. These are diseases that people
11 devote their lives to understanding and live with
12 day in and day out, so there's just a tremendous
13 wealth of knowledge that can be brought to bear.
14 In a moment, I'll touch on that just a little bit
15 more, but certainly a really valuable use of
16 information that we should bring to bear when we're
17 thinking about trial design.

18 Natural history studies, summary level data,
19 individual patient data, PK/PD, preclinical, many
20 of these were discussed yesterday, so really what
21 we're talking about today is maybe ways to better
22 utilize this type of information. Of course, any

1 time we would then be looking to use the Bayesian
2 approach and bring historical data to bear, we need
3 to be thinking about its relevance, similar
4 indications, patient population, the relevance of
5 the endpoints, and so forth.

6 On the next slide, as I mentioned, I did
7 want to touch on the role of opinion or expert
8 knowledge. Just so you're aware, if you're not
9 yet, there is large literature on this topic in
10 eliciting beliefs about endpoints; there is a lot
11 available. There are formal, well-tested protocols
12 for eliciting distributions about belief, and all
13 of the methods we would talk about then can be
14 applied such that one can downweight that
15 information when thinking about it relative to
16 maybe formal data that would be coming in.

17 We've used it quite a bit at Lily in a
18 variety of settings. Oftentimes it's to help us in
19 the design of a trial, to be thinking about maybe
20 relationships between endpoints, or doses, or
21 populations. It has also been used to inform about
22 the relevance of historical information. So in a

1 case where maybe we want to borrow some adult data
2 in a pediatric setting, partnering with a patient
3 has been used and published to help in thinking
4 about how much to borrow.

5 There are examples available. MYPAN is a
6 rare inflammatory disease in children, where prior
7 elicitation was used. Again, I think there are a
8 lot of unexpected benefits. I've heard others talk
9 about the unexpected benefits on elicitation that
10 may not be fully used and could help with us as
11 we're thinking about designing trials, and really
12 setting things up to be as efficient as possible.

13 With that, on the next slide, just some
14 general comments about borrowing, and then I'll
15 show an example. Of course, there are some
16 questions about how much to borrow and things to
17 think about, and what data is eligible to be
18 included we need to understand. I won't go into
19 detail about simulating operating characteristics,
20 but that's a really important element. Considering
21 things such as prior effective sample size or the
22 prior probability of success is important, and of

1 course, understanding prior to posterior
2 sensitivity. We may borrow different amounts for
3 different treatments based on the medical need and
4 so forth.

5 Just one thing I'd like to mention is
6 sometimes there's a feeling that borrowing is
7 brought to bear to help favor a sponsor, but in
8 fact it's really just getting at what is the most
9 likely effect here, and getting the best estimate
10 of the true event we're interested in, so to dampen
11 it but, again, seeking to get at get what that true
12 effect is.

13 Next slide. What I wanted to do next is go
14 into an example. This is a hypothetical example.
15 I presented this elsewhere, but just to show what
16 happens on the backend when we analyze data in this
17 way, looking at a static and a dynamic prior.
18 Suppose that we have previous data on a control
19 group -- and it could be a trial, it could be a set
20 of trials, whatever -- and somehow we've
21 synthesized this information. We have 120 subjects
22 with 72 responses, so the historical rate is

1 60 percent. What we're going to look at is keeping
2 that historical rate constant, and then what we're
3 going to do is say, suppose we have a future trial
4 looking at 70 for the control and 140 in the new
5 treatment?

6 Next slide. What I want to show here is
7 just looking at what those priors look like for the
8 power prior versus the mixture prior. On the left
9 hand is a power prior. The power prior amount of
10 borrowing is governed by this value a_0 . The a_0 can
11 range from 0 to 1, and you can see as you go across
12 and sort of down, as a_0 goes from 0 to 1, you have
13 the non-informative prior up to, really, the whole
14 of the prior at a_0 equals 1 or borrowing the entire
15 prior.

16 Then on the right-hand side, we have mixture
17 priors. This is just an example of, again, that
18 same prior, where here the p is governing the
19 amount of borrowing from the informative part of
20 it, so we can compare. Just taking the blue, for
21 example, you can compare a_0 equals 0.25, and you can
22 see that, versus on the mixture side with p equals

1 0.25, you can see the difference in the weighting
2 of the non-informative portion of this mixture
3 prior is represented there in the graph.

4 All I wanted to highlight in the next couple
5 of slides is how does this affect what we observe
6 on the backend. This is showing for a power prior,
7 so suppose on the left-hand side, we observed
8 20 out of 70 in the control arm. What will happen
9 is you can see the prior is the blue, which on this
10 slide is the rightmost distribution. The
11 likelihood is the red, so then the purple lands in
12 between, pushing forward, in this case, the
13 inference to be closer to that prior and
14 downweighting that observed data. On the
15 right-hand side, you then see suppose the observed
16 data is higher than the prior, and, of course, the
17 posterior then would end up in between.

18 Then we can contrast that, on the next
19 slide, with the mixture approach. Here, very
20 similar observed cases, and the purple you can see
21 then on the left-hand side is now shifting more,
22 and it's coming more towards the likelihood, and

1 you can see on the right-hand side, again, similar.
2 So in this case, where there's mismatch, the
3 posterior is going towards that data. So that's
4 the interpretation of results or what results look
5 like with a couple of different priors.

6 I just wanted to, then, touch on one more
7 example here on the next slide, again, to say how
8 this works and to emphasize that it's really quite
9 straightforward and, again, intuitive. Here's an
10 example where we have an open-label study. It
11 could include an active reference arm, but what
12 we're really interested in is analyzing and looking
13 at that test treatment.

14 Suppose a control isn't feasible or it's
15 unethical. What we would use here is maybe this
16 historical information to establish what a
17 meaningful response would be, and then we need to
18 set up our decision rule. In this case, suppose
19 that we have utilized this historical information
20 and we've determined that 57 percent is the
21 effective interest and that 80 percent is the
22 probability threshold. Then all we're doing is

1 we're just setting up a decision rule that says the
2 probability that that response rate is greater than
3 0.57 needs to be greater than 0.8.

4 On the next slide, we can apply this.
5 Suppose we have a study with 30 patients in
6 juvenile idiopathic arthritis. In this case, we're
7 just showing we aren't using a formerly informative
8 prior on the treatment arm, but to say, okay, we've
9 observed now 20 out of 30 responders at week 24 in
10 this trial, so our observed response rate is 0.67,
11 roughly, and then on the next slide, all that we
12 need to do -- and it may not come over in this
13 form. I'm not sure if it builds.

14 Do you mind to go to the next to sort of
15 build that out? Okay. It does build.

16 So basically, all we're trying to do is look
17 at that effective interest, calculate the area
18 under the curve to the right of 0.57, and that's
19 85 percent. On the next slide then, applying that
20 decision rule, as success criterion has been met,
21 so that's how that simply would work.

22 The final thing in this part, on the next

1 slide, I wanted to touch on is predictive
2 probabilities. Again, we talked a lot about this
3 being a continual learning process, and in
4 particular focusing on using information that may
5 be external to the trial. Obviously, we can use
6 information within the trial and make decisions as
7 the trial progresses.

8 Again, this was touched on in the earlier
9 session, but basically the idea here is just
10 showing an example where suppose we have a final
11 sample size of 100, and we're going to declare
12 success if the probability of a response being
13 greater than 0.5 is greater than 96 percent. Then
14 this is just showing how those predictive
15 probabilities of being successful at N equal 100
16 progress, so we can see early on that predictive
17 probability is 54, and then as we progress, it
18 becomes very small. We would establish a decision
19 rule early about what predicted probability
20 would be -- if we saw that, then we would terminate
21 the trial to begin allowing for that continual
22 learning.

1 I know I just have about another minute or
2 two left, so let's go ahead and jump real quick
3 then. I wanted to touch maybe on this next slide,
4 and then I can close here.

5 Master protocols are a great way to look at
6 multiple indications or multiple drugs, and can be
7 very, very efficient in many settings, but in
8 particular, in the rare disease setting. Bayesian
9 methods will often feature heavily in these and, of
10 course, it also is allowing for more consistent
11 data, getting back to these data are then collected
12 under very similar circumstances, same timing,
13 similar inclusion/exclusion, and so forth.

14 This is an example trial that we did in
15 oncology at Lilly. We had two rare oncology tumor
16 types, and randomized patients to the drug or
17 control in these two indications. So at the end of
18 the day, we were able to bring in real-world
19 information and prior elicitation as well, and use
20 a dynamic model to analyze this data, being able to
21 more efficiently use the information between these
22 two tumor types.

1 The final on the next slide, this is an
2 example of a platform type of trial. This is in
3 pediatric IBD. Patients are randomized to a drug,
4 and then they follow a similar schedule of
5 activities. Again, in these cases where there are
6 very few patients, we want to have these trials
7 together so we're getting same endpoint, same time
8 points, and able to really use that information
9 together.

10 With that, I would like to conclude on the
11 next slide just to say Bayesian design analysis can
12 help facilitate rigorous incorporation of the
13 relevant data, especially in settings with
14 potentially limited sample size. Again, as I
15 mentioned, it's prespecified, it's transparent, and
16 needs to be studied via simulation to really
17 understand, before the trial is conducted, how this
18 will play out. It can result in an increase of
19 power. It, in some cases, can maintain low type 1
20 error, although it will often be inflated, but
21 there are methods to help mitigate against that,
22 and very much around efficient continual learning.

1 Designs such as master protocols can help
2 enhance this learning. In all of these cases, the
3 collaboration between the sponsor or regulatory is
4 very important to statisticians, others, and
5 bringing that patient experience to bear is
6 critical. We just need to continue to have
7 experience with these designs and analyses to
8 continue advancement.

9 DR. ROSENBLUM: Great. Thank you,
10 Dr. Price, for the excellent presentation.

11 Our next speaker is Dr. Jack Lee, Professor
12 in the Department of Biostatistics at the
13 University of Texas, MD Anderson Cancer Center.
14 He's also the Kennedy Foundation Chair in cancer
15 research there. His areas of expertise include
16 design and analysis of clinical trials, Bayesian
17 adaptive designs, statistical computation and
18 graphics, drug combination studies, and biomarker
19 identification and validation.

20 I just met Dr. Lee at the previous ENAR in
21 person for the first time, and I'll pass it off to
22 Dr. Lee.

Presentation - Jack Lee

1
2 DR. LEE: Thank you so much for that kind
3 introduction, and I also thank you for the
4 opportunity to share with you the Bayesian Adaptive
5 Design and Information Borrowing for Efficient and
6 Accurate Statistical Inference in Rare Diseases.

7 Next slide please. I will talk about the
8 statistical challenges and solutions in drug
9 development for rare diseases. I will give a brief
10 overview of the Bayesian statistical inference,
11 then give some examples of the clinical trial
12 design and analysis, and finally give concluding
13 remarks.

14 Next. As shown here in this dart-throwing
15 example, in the top-left panel, the result is not
16 accurate nor precise. In the top-right panel, the
17 result is accurate but not precise. In the
18 bottom-left panel, the result is precise but not
19 accurate. What we want to be is in the
20 bottom-right panel, where the result is both
21 accurate and precise. So we need to have methods
22 that can reduce bias and also increase the

1 efficiency.

2 Next slide. In drug development, we are
3 facing many challenges, shown on the left, and we
4 propose some solutions on the right. For example,
5 we know that randomized controlled trials are gold
6 standards, but then it requires a large sample
7 size, and hence not feasible in rare diseases. So
8 what's the solution? The solution is to have novel
9 adaptive designs.

10 For example, we can take all-comers with
11 adaptive randomization to put more patients in
12 better performing arms. We also need to implement
13 more frequent interim analyses. Lastly, we need to
14 make the study enrollment and conduct easier. It's
15 been discussed that single-arm trials are subject
16 to bias because there's no comparators, and it's
17 hard to make robust inference.

18 So how can we do better? We can borrow
19 information from the concurrent control or
20 historical control. As discussed yesterday,
21 nowadays we have many good registries and EMR data,
22 and these are large sample sizes that often times

1 comes from heterogeneous groups with mixed data
2 quality. So how can we better use real-world data
3 to turn real-world evidence? We need to have a
4 clever way to form synthetic controls, and we can
5 do propensity score matching and network
6 meta-analysis.

7 Next. This slide just shows you a simple
8 example of how statistical inference can be made.
9 Let's assume we want to estimate the unknown
10 response rate for a new drug and we conduct a
11 phase 2 trial with 30 patients. At the end, we
12 have 14 responses and 16 no responses, so the point
13 estimate of the response rate is 0.467. The bottom
14 left-hand panel shows how the posterior probability
15 of response rate is updated. The gray line in the
16 background shows how the previous step was done.
17 And at the end, we have the red line showing the
18 posterior probability of the response rate. As it
19 can be seen, as the trial moves along, the
20 distribution picks up. Why? It's because we have
21 more data; therefore, we have a more precise
22 estimate. On the bottom-right panel, it shows the

1 frequentist 90 percent confidence interval.

2 Next. With the Bayesian posterior
3 probability, we can do many things. For example,
4 we can calculate what's the probability of a
5 response greater than 0.3, 0.5, or 0.6, as being
6 shown in the figure with the area highlighted in
7 red. In the bottom panel, you can also see that we
8 can calculate the probability that the null
9 hypothesis is true or probability of the
10 alternative hypothesis is true. With that, we can
11 calculate the odds and compare the posterior odds
12 over the prior odds to form the base factor. In
13 this case, after we observe data, we can conclude
14 that the alternative hypothesis is 17 times
15 stronger than the null hypothesis, based on the
16 data.

17 Next. In the Bayesian inference, all
18 information pertinent to the parameter of interest
19 is contained in the posterior distribution, so in
20 our case, we start with data prior, and then we can
21 model the unknown parameters with statistical
22 distributions, and then we can properly address

1 various levels of uncertainty. In this talk, I
2 want to emphasize two things; that the Bayesian
3 method allows us to use all available
4 information -- prior, current, and even predict
5 into the future -- to make decisions, and we can
6 use information within and outside of the trial via
7 dynamic borrowing to increase the study efficiency
8 and also allow more frequent monitoring and
9 decision making.

10 You can incorporate subjective utility in
11 the decision making; for example, like the toxicity
12 and efficacy trade-off, which I'm not going to talk
13 about in my talk. But we need to be aware if the
14 data and models are compatible, then where they are
15 biased due to some data heterogeneity, we also can
16 do the sensitivity analysis to evaluate how robust
17 the inference by varying priors.

18 Next. This slide shows us the current
19 status and the enhancement of clinical trials. In
20 current status, we conduct a clinical trial with
21 one drug, one study population, and one trial at a
22 time, and we have discrete-phase drug development,

1 starting from phase 1, then pause to go to phase 2,
2 and then pause to go to phase 3, so it's not
3 efficient. As Dr. Karen Price mentioned, we can
4 use master protocol with a seamless phase design,
5 umbrella basket, or platform trials, and we can
6 also include adaptive randomization to put more
7 patients in more promising arms.

8 Again, the two important things in data
9 analysis is that we should do more frequent interim
10 analysis if the drug is too toxic, or not
11 promising, or highly promising. Then we can have
12 early stoppings, and then we can borrow information
13 from historical data, or across subgroups, or even
14 across different trials.

15 Next. I just want to quickly mention about
16 one adaptive design that we developed called the
17 Model-Assisted Design, which is the design that
18 uses a Bayesian framework. We can have
19 precalculated decision rules such that the design
20 is very easy to conduct, that includes the Bayesian
21 Optimal INterval design, or BOIN design, for dose
22 finding, and also the Bayesian Optimal Phase 2

1 design, or BOP2 design, for complex endpoints.
2 Notice the font in the red color that shows all
3 these designs allow us to incorporate historical
4 data as informative prior.

5 Next. This slide shows you that at
6 MD Anderson, we not only develop methods, but we
7 also provide software. We have more than 30 freely
8 available shiny apps that allow you to design and
9 run clinical trials. All you need to remember is
10 our URL at the bottom, trialdesign.org.

11 Next. For example, we have a family of BOIN
12 designs for single agent, for combined agent, and
13 for late-onset toxicity, and we also have design,
14 like BOIN design, BOIN12, to allow us to find the
15 optimal biological dose.

16 Next. This schema shows us how to choose
17 the right design. On the left, you can see that if
18 you want to find OBD, then you can use only one
19 tool, U-BOIN. If you want to find MTD, you can use
20 iBOIN, TITE-BOIN, or BOIN combination. The one
21 highlighted in purple, I just want to highlight the
22 importance that we use the prior data or historical

1 data.

2 Next. This is a simple diagram for the BOIN
3 design. All you need to do is determine the target
4 probability of toxicity ϕ , and calculate the
5 observed probability of toxicity p_j at dose level j ,
6 and compare with the prespecified boundaries to
7 determine if either dose should be escalated,
8 retain, or de-escalated. The iBOIN design can
9 incorporate prior data, as you can see on the next
10 slide, but I want to make a really important plea,
11 that it is long overdue to abandon the commonly
12 used 3-plus-3 design because there are many better
13 alternatives.

14 Next. So as can be seen here at the left
15 panel, we can see that we have different doses and
16 have different expected probability of toxicity,
17 but then we can put the effective sample size
18 that's corresponding to historical data and how
19 strong the historical data is, and then to model
20 that when we do the decision. On the right, you
21 can see the decision diagram, and with low
22 probability of toxicity, you escalate, and high

1 probability of toxicity, you de-escalate the dose,
2 and if it's the middle, then you retain.

3 Next. At the end, you plug in the number of
4 patients treated and number of dose-limiting
5 toxicities observed, and can calculate the MTD with
6 its confidence intervals. As you can see now, the
7 dose chosen is the dose level 3 MTD.

8 Next. This slide shows that we have
9 different types of model-assisted designs for a
10 phase 2 trial as well, and this we call BOP2 Suite.

11 Next. The BOP2 design allows us to run the
12 phase 2 trial with a unified framework with
13 different endpoints. In particular, depicted on
14 the right, you can see that, for example, in the
15 control arm, the probability of efficacy is assumed
16 to be 0.3, but we have data with the prior efficacy
17 showing that it's corresponding to a sample size of
18 20, and then for the experimental arm, we expect
19 real efficacy, 0.5, but we don't have much data, so
20 we only put the prior effective sample size of 1.

21 Next. This shows you that under the BOP2
22 design, you can have different endpoints; like in

1 example 4, we have objective response and toxicity
2 as endpoints, and we have a different stopping
3 rule. For example, with 10 patients, if we observe
4 two or less responses or five or more toxicities,
5 then we stop the trial and declare the drug is not
6 working. This shows you the importance and
7 feasibility of monitoring the trial during the
8 interim.

9 Next. This shows a graphical presentation.
10 On the left, you can see how the trial evolved in
11 terms of responses, and on the right, in terms of
12 toxicity. The green zones are the go zones, and
13 the pink zones are the no-go zones. In this case,
14 the trial stopped after we observed 35 patients
15 because the drug was too toxic.

16 Next. Just to quickly go over the platform
17 design for adaptive enrichment, since it has been
18 discussed by previous speakers, I'll quickly run
19 through the study schema.

20 Next. The idea for the adaptive platform
21 design is that we can have the control group as the
22 backbone of the platform and experimental

1 treatments as modules, which can be plugged in and
2 out of the platform.

3 Next. For example, if we want to compare
4 experimental 4 with control, we can either use a
5 concurrent control, shown in the light brown, or a
6 historical control, shown in purple. Each one has
7 its own merits and also disadvantages, so we just
8 need to use them carefully.

9 Next. This shows you the normal design with
10 master protocols on the left. This phase is an
11 exploratory kind of design analysis to try to find
12 the signal. After we find the signal, then we can
13 design the confirmatory trial with more focus on
14 the phase 3 trial in the selected patient
15 population.

16 Next. My last part of the talk will cover
17 the Bayesian hierarchical model for synthesizing
18 information for the subgroups analysis in basket
19 trials. Because a clinical trial often has
20 different subgroups, how do we model them such that
21 the information can be borrowed? The Bayesian
22 hierarchical model can synthesize multisources of

1 real-world data.

2 Next. To increase accuracy and also
3 increase the efficiency if we do the right
4 borrowing.

5 Next. This is just to depict that we have
6 five subgroups and different age groups. On the
7 left, you can see the prior distribution of the
8 response rate, and on the right, you can see after
9 borrowing, showing in the red, the posterior
10 distribution. The information tends to move to the
11 center and peaks up.

12 Next. This shows you that if we apply one
13 of the methods we call BaCIS, then you can classify
14 these five subgroups into two clusters, then borrow
15 information within the clusters.

16 Next. In cluster 1, we have 2 arms or
17 2 subgroups, and you can see the red curve is
18 higher than the blue. What does it mean? It means
19 we have more information,

20 Next. Cluster 2 has 3 arms or 3 subgroups,
21 so then, as you can see, the information is closer
22 to the center of the three, and the red curve is

1 higher than the blue curve.

2 Next. We can also use another package
3 called BCHM, Bayesian classified hierarchical
4 model, and in this case you will determine the
5 number of clusters automatically, so in this case
6 we formed 3 clusters.

7 Next. In conclusion, statistics can help us
8 in extracting signals from the noise in the data to
9 avoid bias and increase efficiency. There's no
10 free lunch, but there are some lunch specials if we
11 apply some novel design and analysis. The Bayesian
12 paradigm takes the "we learn as we go" approach,
13 and is particularly useful in rare diseases because
14 it allows flexible, adaptive, and continuous
15 learning, to naturally and easily incorporate and
16 synthesize all relevant information.

17 Bayesian adaptive designs are efficient and
18 robust in the drug development process, but there
19 is one caveat. All signals found need to be
20 validated in prospective trials, so please work
21 closely with statisticians from beginning to end,
22 and apply rigorous statistical methods to maximize

1 the success of every project. Thank you very much.

2 DR. ROSENBLUM: Great. Thank you, Dr. Lee,
3 for an excellent presentation.

4 I'll turn to our third speaker. Dr. Rima
5 Izem is Director of Statistical Methodology in
6 analytics at Novartis. She has expertise in
7 regulatory statistics using causal inference for
8 comparative safety, signal detection, and survey
9 research. She also has experience and expertise in
10 comparative effectiveness in rare diseases at
11 Children's National Research Institute. She works
12 with real-world data, including claims data,
13 electronic health record data, international
14 registries, and electronic clinical outcome
15 assessments. We're honored to have Dr. Rima Izem,
16 and I'll turn it over to you.

17 **Presentation- Rima Izem**

18 DR. IZEM: Thank you so much. I've really
19 enjoyed the workshop so far. My talk will switch
20 gears a little bit from the previous, although we
21 will continue discussing design and analysis that
22 tries to get as much efficiency from the

1 participants in the rare disorders. My talk will
2 focus on leveraging longitudinal data, or in other
3 words, leveraging time.

4 Next slide please. As this is the last talk
5 of the session, I wanted to put the take-home
6 messages right away, and those are the following.

7 First, there are multiple ways to incorporate
8 randomization in your study design beyond a
9 parallel control, and a lot of these designs
10 exploit within-subject comparison, or what I would
11 call sometimes self-control, and that could be used
12 to establish efficacy or safety.

13 In the same fashion, there are also multiple
14 observational study designs beyond the cohort
15 study, or even single arm with an external control,
16 that tries to leverage longitudinal data or
17 repeated measure. However, anytime you move away
18 from randomization, you have to control for
19 multiple sources of bias, like a confounding and
20 selection bias.

21 So why include time or why include
22 within-subject comparison? Because it has a lot of

1 advantages compared to between-subject comparison,
2 in that you're increasing your analysis unit,
3 you're reducing potentially outcome variability,
4 and if you're using an observational study design,
5 you're reducing confounding compared to
6 between-subject comparisons.

7 Next slide please. With that, this is the
8 outline of my talk. I will spend some time
9 discussing how to implement randomization, maybe
10 slightly differently than the speakers earlier
11 today, but maybe less novel; that's my disclaimer.
12 I use this as a motivation before getting into
13 observational study methods because some of them,
14 to make them more rigorous, will try to emulate a
15 randomized design with an observational study, and
16 then I'll finish with some design and analysis
17 consideration.

18 Next slide please. This is basically my
19 outline and my take-home messages all in one slide,
20 but also showing the running thread as I move from
21 randomized to observational studies. In the
22 left-hand side, I show randomized study designs

1 that, again, with randomization, you get a lot of
2 benefits; that is that you control for all
3 confounding, and also you have a really good idea
4 of time zero for a lot of these designs. The
5 parallel arm is probably the most commonly used,
6 where you're randomizing different people to
7 different treatments; however, you can see also
8 that there are other designs that I'll go into with
9 a graphic in the next slide.

10 On the right-hand side are observational
11 study designs. Again, the top one is probably the
12 most commonly known, is the cohort study, or using
13 an external control arm, but the bottom two may be
14 less known, but they're still useful. Now, the
15 reason I have arrows going from one to the other,
16 it's not really to say that they're equivalent, but
17 rather that if you are going to use observational
18 studies like self-control, case series, or cohort
19 study, or sequential control for confounding, it's
20 helpful to keep in mind what would have been the
21 randomized study equivalent, or what would have
22 been ideally, hypothetically, the randomized study

1 that would correspond to this observational study.
2 The reason you do that is that, in itself, with
3 already existing data, emulating a randomized study
4 design is a way to control for multiple sources of
5 bias. And again, I'll be following the same
6 outline of the paper that's shown in the bottom.

7 Next slide please. One more. This is my
8 favorite graphic. It's a little bit dated now
9 because it doesn't include a lot of study designs
10 that you've heard of before this talk, but I really
11 like it because it shows you the diversity of study
12 designs that you can think about, and especially
13 how to incorporate randomization in your design.

14 In A is the parallel group design, but then
15 in B and D are designs that rely solely on
16 within-subject comparisons. In the crossover
17 design that was mentioned earlier today, each
18 person receives both treatments, but what's
19 randomized is the order in which they receive the
20 treatment. So again, it's a randomized design, and
21 yet each person gets to test both treatments.

22 In the N of 1, it's kind of taking the

1 crossover to the limit. You only have one subject,
2 but you have multiple treatment periods. In this
3 case, that's what the N in N of 1 refers to, and
4 then what's randomized in each treatment period is
5 which treatment will be received in that treatment
6 period. So again, that's a randomized design with
7 only one patient.

8 In the bottom in blue are designs that are
9 not only relying on within-subject comparison, but
10 also between-subject comparison; however, they use
11 kind of both. For example, in the randomized
12 placebo phase, what's randomized is whether the
13 patient is receiving treatment right away, after
14 they enter the study, or after a placebo period
15 they will receive the treatment.

16 In the randomized placebo, and the stepped
17 wedge, and the randomized withdrawal, all of the
18 patients receive the treatment, but what's
19 randomized is when they receive it and how long
20 they receive it. So in the analysis stage, the
21 analysis unit is a period rather than a patient, so
22 you're using both within-subject and

1 between-subject comparisons.

2 Next slide please. Just to show you that
3 these designs are feasible, but also that they can
4 lead to significant finding, or statistically
5 significant finding, I'm sharing here an example
6 from urea cycle disorder. This was an N of 1
7 design. Here it used 6-week periods, so there were
8 6 weeks that were split in 3 pairs, and in each
9 pair, the patient was randomized to either receive
10 placebo or active within a given week. This was
11 one particular patient that had a mutation for this
12 rare disorder and had to have a washout period
13 prior to entering the study.

14 There were three outcomes of interest. One
15 was the patient-reported outcome, a questionnaire
16 score to ask about their symptoms, but also there
17 were two laboratory measures of their well-being.
18 Although there were only 6 weeks worth of
19 observation, at least in this study, it was
20 sufficient to show that the treatment of L-arginine
21 significantly improved symptoms, as evidenced by
22 the questionnaire score, but also their glutamine

1 level.

2 Next slide please. If we try to both use
3 within- and between-subject comparisons, there's
4 also another example also in urea cycle disorder
5 that recently used a design that used a
6 hospitalization episode for hyperammonemia as the
7 analysis unit rather than the patient. Every time
8 the patients that were participating in this study
9 were admitted into the hospital, that was
10 considered an analysis unit that was either
11 randomized to receive CARBAGLU or to receive
12 placebo. With only 24 patients, there were
13 42 analysis units in CARBAGLU and 48 in the placebo
14 arm.

15 What we're showing on the right is something
16 that was shared in the label for this drug. We see
17 how over time in the horizontal axis, the
18 proportion of events that occurred in both the
19 treatment arm and also the placebo arm, and in the
20 events here, higher is better, so it's a responder
21 rate. And we see that the response rates, or the
22 two curves, kind of differentiated after day 1 off

1 hospitalization, and it continued to differentiate
2 as time goes on. So it is possible to think of the
3 analysis unit as the treatment episode rather than
4 the patient.

5 Next slide please. In summary, there were a
6 lot more studies that were cited in some of the
7 examples that I showed earlier, that you can go
8 back to, to show not only a stepped-wedge design,
9 but also early withdrawal or delayed therapy. The
10 main advantages of using the designs is, again,
11 increasing the analysis units, maybe reducing
12 variability, and then also getting information on
13 the natural history of the study. However, these
14 designs are not always feasible, and what's the
15 tricky part is to figure out the timing or the
16 duration of these time periods. They should be
17 long enough to observe a change in the outcome, but
18 they shouldn't be too long. They should be short
19 enough to assume that these time periods are
20 independent or may be including some washout
21 period.

22 Next slide. I'll move now to observational

1 longitudinal design, and I apologize, but I'll have
2 to skip some of the slides as we go through this.

3 Next slide. Next. Next slide please. I
4 skipped the cohort design, and that was an example
5 that you can refer to, and I'll be happy to answer
6 any questions you may have on that example during
7 the Q&A, but I wanted to share a few thoughts on
8 using the single arm with external control.

9 Whenever you talk about using or leveraging
10 observational studies, that's probably the first
11 idea that comes to mind, at least for my
12 collaborators in academia, but also industry. They
13 think about conducting a single arm, and then using
14 the observational study instead of randomizing
15 their control. That is, frankly, a very
16 challenging approach, especially in the regulatory
17 setting, and we've reviewed, actually, a lot of
18 applications that tried to make the argument for
19 using an external control, and this is the paper
20 that's referred in the bottom.

21 The difficulty with using external control
22 is that it's very hard to show that two different

1 data sources have similar information. It's very
2 hard to show that there is a comparability of
3 population, treatment, outcome, and frequency of
4 assessment, and that the start and end of follow-up
5 was similar. It's also hard to control adequately
6 for confounding because sometimes the information
7 on confounders is not even collected, or not
8 collected sufficiently frequently, and it's hard
9 when you have to tune the control for confounding
10 methods to prespecify everything at the beginning.
11 So there are more exceptions than rules in using
12 observational study of an external control.

13 Next slide. I'll skip this example, but
14 this was an example of actually the exception. The
15 only thing maybe that I will say here before our
16 skip this slide is what's shown in the label on the
17 left is the effect size, and if you see what effect
18 sizes we're talking about here, we're using an
19 external control that's huge. So definitely, when
20 the effect size is really large, maybe it's
21 worthwhile to discuss having an external control.

22 Next slide please. I've moved very quickly

1 for a cohort study and use of external control
2 because I wanted to spend some time to discuss
3 self-controlled case series, and also maybe
4 sequential control for confounding. The reason I
5 wanted to spend time on those is because I don't
6 think they're used enough, or they're not thought
7 of enough. Especially early on when you're doing
8 proof of concept, they could be leveraging already
9 existing data in your natural history studies to
10 see whether maybe there's a treatment that's
11 promising. It's helpful for those studies, to
12 think of them as the observational study equivalent
13 of crossover, or N of 1, or a sequential
14 randomization study.

15 Next slide please. What are some of the
16 questions that you can ask yourself if you already
17 have a natural history study like some of the ones
18 that were discussed in the workshop yesterday? Can
19 the unit of analysis be subject time rather than
20 just subject? Can the duration of the look-back,
21 that is the medical history, instead of it being
22 static, can it be dynamic? Can you actually look

1 at it longitudinally before diagnosis?

2 Similarly, do you collect information on
3 potential confounders sequentially over time or do
4 you collect this once and for all at baseline when
5 people enter the study, and then you don't look at
6 it again? All of these can inform whether a
7 self-controlled study, or something that augments
8 self-control, with between-subject comparison is
9 good for you.

10 Next please. One successful example that
11 will leverage existing data is the alpelisib, and
12 this is a relatively simple self-controlled study
13 where each person was serving as their control, but
14 you only have two periods, a pre-period before the
15 treatment and a post-period after the treatment.
16 The data here that was used was from a
17 compassionate-use program, so this product was
18 already approved, and this was just to expand the
19 indication for this new population.

20 There were already some findings that
21 supported that maybe this treatment was probably
22 effective, so this study kind of formalized this

1 comparison of the pre-index period to the
2 post-index period. Although the outcome of the
3 responder was not apparent right away, just by
4 calling it a responder because you're comparing to
5 baseline, it is a pre-post comparison that's
6 determining whether this treatment is working or
7 not. So again, just by using pre-post comparison
8 in this case when there was a lot of information
9 already available on this drug, you were able to
10 see that it would work for this rare disorder.

11 Next slide please. One way to generalize
12 this and this type of design, this self-controlled
13 case series, is not really common in rare disease,
14 but it could be used. It's used a lot in
15 postmarket safety to try to leverage observational
16 data for rare outcomes, but I think it has promise.
17 And again, for me the self-controlled case series
18 is the observational study equivalent of the N of 1
19 design, although you could have more than one
20 person, obviously, if you have multiple periods for
21 each person.

22 The study is anchored at the first exposure

1 to therapy, and I'm showing here the exposure
2 period to a particular test drug in yellow;
3 however, you can still use information prior to
4 this first exposure therapy, especially if there
5 was standard of care or exposure to placebo. The
6 periods may have to not be exactly contiguous if
7 the test drug can have an effect beyond when the
8 subject was exposed, so you may need to include
9 washout period. But the point here is that if you
10 have a transient exposure, where a person is
11 receiving the treatment, not just once but multiple
12 times over their journey, then that could be
13 exploited in this analysis to see whether the test
14 drug is effective compared to the placebo drug.

15 Next slide please. A little bit more
16 complex to explain design is what I call sequential
17 cohort entry. Just to give you an idea of how this
18 works, I'm going to be using an example that I
19 worked on in urea cycle disorder. In this example,
20 the interest was to leverage natural history study
21 data from hundreds of patients to see whether liver
22 transplantation was improving multiple outcomes,

1 including hospitalization for hyperammonemia, or
2 quality of life, and survival. The outcomes vary a
3 lot based on the disease severity.

4 What was difficult in this particular
5 leveraging observational data is that the time of
6 intervention was very clear for those who received
7 liver transplantation. That was the day they
8 received the transplantation. However, for those
9 who didn't receive transplantation, finding a start
10 of follow-up was really hard. Because they're
11 defined by not receiving an intervention, there is
12 no anchor and, again, this is an observational
13 study, so you don't have a randomization that sets
14 everything to time zero.

15 Because liver transplantation happened early
16 in age, the time scale here that I'm showing on the
17 red line is age, so you can think about it as age
18 in days. Here, I'm showing 2 days where that
19 intervention had occurred in the database, but of
20 course there were a lot more days. There were
21 around 100 eligible patients that received the
22 liver transplantation.

1 One way to get around thinking about when to
2 start follow-up is to think about this sequentially
3 or as a sequential cohort entry. For example, at
4 63 days, we could ask ourselves who would have been
5 eligible to receive transplantation that maybe
6 either received it at 63 days or later, and then
7 match those who actually received liver
8 transplantation at 63 days with those that did not
9 receive transplantation. By doing this, you're
10 allowing someone who received a liver
11 transplantation later in their life to be a control
12 for someone who received a liver transplantation
13 early in their life. You do this sequentially, and
14 that's a way to solve, first making sure that
15 you're not including immortal time bias, but also
16 control for confounding in every strata.

17 Next slide please. In summary, for a
18 non-randomized comparison, you have similar
19 advantages to randomized comparison. When you're
20 using within-subject comparison, you're augmenting
21 between-subject with within-subject. I'm
22 highlighting here two maybe additional arguments

1 for using self control in longitudinal design,
2 which is that when you're comparing one person to
3 themselves, you're often controlling also for
4 confounding, and also by looking at time periods,
5 you're solving the probability of time zero, making
6 sure that you're starting at the right time.

7 Next slide please. As with randomized
8 study, timing is really important, and that's the
9 trickier part. In addition to the duration, making
10 sure that it's just right, not too long and not too
11 short, you have to also ask yourself whether some
12 of the confounding that you're worried about is
13 actually time-varying because you would need to
14 adjust for that. In terms of the analytical
15 considerations, most of the time, whether you're
16 using self-control, you have to correct for a
17 correlation of measurements within the same
18 subject. So if it's only self-controlled, you just
19 need to use paired tests or paired analysis;
20 however, when you're augmenting between-within,
21 then you probably need to use a hierarchical model
22 for the adjustment.

1 I didn't discuss the case control, but you
2 need to think about anchoring in a lot of these
3 designs.

4 (Alarm sound.)

5 DR. IZEM: Sorry. I had put a timer, and
6 I'm out of time, so next slide.

7 This take-home you've seen before. You can
8 skip a few more slides. I just wanted to put a
9 plug. These are acknowledging all of my
10 collaborators. This is a plug for IDEAL, which is
11 on the same side of the Atlantic, I guess where I
12 am now, which has a lot of very useful work on rare
13 diseases and study designs that is very relevant to
14 the audience today.

15 Then finally, the last slide is my email
16 address if you have any questions. Thank you.
17 Sorry for going a little over.

18 DR. ROSENBLUM: No, that was great. Thank
19 you, Dr. Izem, for the excellent presentation.

20 We'll now turn to our discussion.

21 Professor of Biostatistics at Vanderbilt
22 University, Dr. Frank Harrell, he is an expert

1 biostatistics advisor at CDER, Center for Drug
2 Evaluation and Research at FDA, and he has a wide
3 range of expertise, including development of
4 accurate prognostic and diagnostic models; model
5 validation; clinical trials; observational clinical
6 research; cardiovascular research; technology
7 evaluation; pharmaceutical safety; Bayesian
8 methods; quantifying predictive accuracy; missing
9 data imputation; and statistical graphics and
10 reporting.

11 We're lucky to have Dr. Harrell here today
12 to give his thoughts on the presentations and the
13 topic in general.

14 **Summary of Presentations - Frank Harrell**

15 DR. HARRELL: Thanks very much for the nice
16 introduction, and what a privilege it is to be
17 commenting on the talks by these three amazing
18 speakers. Not only are they leaders in this field,
19 but they presented very complementary work to each
20 other, so I really enjoyed the presentations. I'll
21 start with just a few comments about each one, and
22 then I'll do some general comments.

1 Karen Price, she gave us a great Bayesian
2 background, and I love that little diagram of knew
3 this, saw this, and now know this, and in feeding
4 that back into a continuous learning cycle. She
5 made really a key point, which is that the Bayesian
6 approach makes integration of pre-study knowledge
7 and new data transparent. There's a lot of ad hoc
8 approaches to do that. They're not very
9 satisfactory, and they're very subject to bias of
10 the observer or of the analyst, so Bayes gives,
11 really, the way forward for that.

12 She talked about different kinds of
13 borrowing, and data sources, and a nice overview of
14 knowledge and belief elicitation, which is a very
15 important component of Bayes. She mentioned
16 Bayesian decision rule for a single-arm study,
17 which does require a bit of an arbitrary response
18 probability threshold, so I worry about that a
19 little bit.

20 Just one other comment that I could maybe
21 bother caring about a little, she mentioned
22 Bayesian procedures that have good alpha or type 1

1 assertion probabilities. That's really kind of at
2 odds with optimum decision-making, and in the chat,
3 I put a link to some more details that has a link
4 to a long discussion about this. So controlling
5 alpha is sort of counter to what Bayes is trying to
6 do, which is to maximize the probability of making
7 a right decision.

8 She talked about continuous learning, where
9 you have maybe a sample size goal, and you might be
10 dealing with Bayesian predicted probabilities to
11 decide when you've learned enough. That does
12 require sort of treating an ultimate sample size as
13 a magic quantity, and I would like to argue that
14 one of the best things we can do in the future of
15 clinical trials is to do away with sample size
16 calculations altogether because that's really how
17 we'll really respect continuous learning.

18 Jack Lee had another fascinating talk, and
19 he gave a nice background about bias versus
20 precision, which are always important to keep in
21 your mind as you're looking at any method. He
22 talked about several novel adaptive designs and how

1 information grows as the sample size grows,
2 advantages of the Bayesian paradigm, and he made a
3 push, which I think is very wise, for moving away
4 from discrete drug development phases but making
5 the process more continuous.

6 He talked about different ways to adapt and
7 different reasons to stop early, such as toxicity,
8 futility, efficacy, and handling complex endpoints.
9 Jack has a long track record of developing, really,
10 state-of-the-art free software for helping people
11 use these sometimes complicated ideas in designing
12 studies, so you should look into his software.

13 He talked about minimizing the expected
14 sample size, which is a way to learn faster and
15 maybe save resources also, and the different sorts
16 of environments, platforms, master protocols, and
17 avoiding noisy subgroup analyses. And I loved this
18 quote, "There is no free lunch, but there are lunch
19 specials." I'm writing that one down.

20 Rima Izem really hit something I was hoping
21 somebody would hit in this session, which is
22 capitalizing on time. I think this is extremely

1 important because in the simple case of doing a
2 univariate outcome, let's say you measure a patient
3 outcome at 2 months versus having the outcome
4 assessed at more times, the payoff of having the
5 longitudinal data is huge.

6 She went further than that to talk about the
7 value of within-subject comparisons in general,
8 including a lot of designs that are not used very
9 often and should be used more often. She mentioned
10 how observational longitudinal data, that even
11 though you can have confounding, you probably have
12 a little less confounding when you have
13 time-oriented data. A big plus for the
14 longitudinal way of thinking is increasing the
15 effective sample size, and your unit of analysis
16 might be a patient response measured in one day, or
17 one week, or one month.

18 She mentioned crossover and N of 1 studies.
19 I learned a lot from her talk. There are other
20 things that she touched on which relate to rare
21 disease and ethics, and the push to not randomize
22 because there are things like delayed treatment,

1 there are randomized withdrawal studies, and
2 stepped wedge, which is another kind of delayed
3 treatment design that really should be considered
4 to answer those concerns that a lot of rare disease
5 communities worry about. She mentioned database
6 fitness for purpose and several other good things.

7 I'll just add a few overall comments. I'm
8 not one of these statisticians that's very
9 optimistic about borrowing historical data, so I
10 tend to be very afraid of historical data. I just
11 want to note that when you do use historical data,
12 it takes extreme diligence, and one of the pieces
13 of that is you have to include a lot of historical
14 data that's very unfavorable to what you're trying
15 to show. You cannot be accused of cherry-picking
16 favorable historical data.

17 The use of historical data actually requires
18 you to use the raw data. There's no way I know to
19 do a really satisfactory analysis using only
20 summary statistics of the raw data. For one thing,
21 the raw data should almost always be covariate
22 adjusted. I see a lot of use of historical data

1 where they're not even adjusting for age
2 differences between historical data and the new
3 patients.

4 Jack mentioned the need for validation
5 studies, especially when you're doing things that
6 are complex like adaptation or borrowing data.
7 When you are borrowing data, you really need a
8 validation study, or if you're doing an adaptive
9 study that might actually lower your effective
10 sample size for the favored treatment regime. And
11 if you really need a validation study, it goes
12 against some of what you're trying to gain by
13 having limited sample size now; so how are you
14 going to get a sample size for the validation
15 study? This is an over-generalization, but I think
16 there's less need for a validation study if you're
17 not borrowing historical data.

18 I want to emphasize what I think is the
19 biggest bang for the buck. The biggest bang for
20 the buck is to have a high-resolution outcome
21 variable that has very high test-retest
22 reliability. An example of that -- this is kind of

1 an idealized example -- would be you're looking at
2 bone mineral density measured with a DEXA scan, and
3 the patients get a DEXA scan at 6 months, 7 months,
4 8 months, out to 18 months, so you spend a lot of
5 money per patient but you don't spend a lot of money
6 on a lot of patients.

7 So if you have a high-resolution, high
8 test-retest measure like bone mineral density, it's
9 amazing what sample size you can get away with and
10 learn a tremendous amount of information. So
11 trying to emulate that with the data we usually
12 have, such as having a 60-level clinical outcome
13 scale, or a 60-level patient-oriented outcome
14 scale, and then you have clinical overrides for
15 events that happen to the patients and you do an
16 ordinal analysis that, say, has 62 levels in the
17 ordinal response, now you're starting to
18 approximate what you can get out of the DEXA scan
19 sort of high-resolution data. Then if you measure
20 that multiple times, you'll have much more
21 information, and that increases statistical power a
22 lot. So the idea also was to not have a sample

1 size but to mimic what physicists do, which is to
2 study until you have an answer, you have enough
3 evidence, and that's when you stop.

4 I will go out on a limb and say something
5 nobody will be ready for, which is there's
6 something more valuable to borrow than borrowing
7 data from historical data, and that is to borrow
8 knowledge about how consistently a treatment
9 affects different endpoints, and you could form a
10 Bayesian prior for how the treatment affects
11 mortality versus how it affects functional status
12 and other patient-reported outcomes. Borrowing
13 that kind of information allows you to learn about
14 multiple endpoints when you don't have enough
15 sample size to study any one endpoint, especially a
16 mortality outcome.

17 Then I'll just close by saying, avoid the
18 worst possible mistake. The worst possible mistake
19 in rare diseases, I saw this actually done, and I'm
20 still kind of in a state of disbelief that a
21 sponsor would do this. They had very great
22 difficulty getting patients because of the rare

1 disease. They were able to randomize about
2 80 patients, and they had an established ordinal
3 scale that had maybe seven levels to the scale, but
4 then they dichotomized the outcome and did a
5 responder analysis about whether or not you had
6 like a 2-point improvement from baseline. That was
7 their definition of a binary response.

8 So what they've done is to say, we've got
9 80 patients -- that's not enough really -- but
10 we're going to do a responder analysis and pretend
11 we had 30 patients because that's what that
12 particular responder analysis did. It reduced the
13 effect of sample size from 80 to 30. So whatever
14 you do, don't lose information in making your
15 sample size smaller than it already is.

16 So thanks for listening, and, again, I just
17 tremendously enjoyed these three talks.

18 DR. ROSENBLUM: Thank you, Professor
19 Harrell, for the excellent feedback on the three
20 talks.

21 This is a perfect place to stop, and I'll
22 pass it over to Dr. Dionne Price, but I first want

1 to thank all the speakers and panelists for an
2 excellent second session of this two-day symposium.
3 Thank you all.

4 **Concluding Remarks - Dionne Price**

5 DR. D. PRICE: Thank you, Michael.

6 I started the day by welcoming you all to
7 our workshop, and I was so eager to jump right in,
8 that I neglected to introduce myself. I am Dionne
9 Price, and the Deputy Director of the Office of
10 Biostatistics in the Center for Drug Evaluation and
11 Research. In this role, I also co-lead the CDER
12 and CBER's, Center for Biologics Evaluation and
13 Research, Complex Innovative Trial Design Paired
14 Meeting program and related efforts, and I'm
15 actively engaged in CDER's Accelerating Rare
16 diseases Cures program, so I encourage you all to
17 explore the FDA website for information on both.

18 Now, in our first session today, we heard
19 about small sample sequential multiple assignment
20 randomized trials, adaptive enrichment designs that
21 could be mapped to some rare diseases, and the need
22 to get more data and/or get more information from

1 the same data.

2 Specifically, we may be able to get maximum
3 information available from limited data via
4 adaptive designs with the goal -- and I quote
5 Dr. Stallard -- "of efficient, smarter, and faster
6 trials." The panel also touched on the potential
7 advantage of group-sequential trials with interim
8 analyses, single-arm trials, and sample size
9 considerations.

10 In our second session, which nicely
11 complemented the first session, we heard about
12 Bayesian thinking, borrowing approaches, master
13 protocols, including adaptive platform trials and
14 basket trials; Bayesian adaptive designs; frequent
15 interim analyses; and leveraging longitudinal data
16 using various designs. So if it's not obvious, I
17 will say it. There is no one size that fits all,
18 but we certainly have heard options and some
19 thought-provoking ideas for designs and analysis
20 methods that may aid in drug development for rare
21 diseases.

22 On behalf of the FDA, I would, again, like

1 to thank all of our speakers, our panelists, and
2 our moderators. I would like to thank you, the
3 participants, for your time, your questions, your
4 engagement, your attention throughout both days of
5 the workshop, and I will conclude with a reminder
6 of our June 7th and 8th public workshop on Novel
7 Endpoints for Rare Disease Drug Development, and
8 that link has been added in the chat. So once
9 again, thank you all.

10 (Whereupon, at 11:58 a.m., the workshop was
11 adjourned.)
12
13
14
15
16
17
18
19
20
21
22