

Safety Testing of Human Allogeneic Cells Expanded for Use in Cell-Based Medical Products

Draft Guidance for Industry

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**U.S. Department of Health and Human Services
Food and Drug Administration
Center for Biologics Evaluation and Research
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Cell-Based Medical Products**

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This draft guidance, when finalized, will represent the current thinking of the Food and Drug Administration (FDA or Agency) on this topic. It does not establish any rights for any person and is not binding on FDA or the public. You can use an alternative approach if it satisfies the requirements of the applicable statutes and regulations. To discuss an alternative approach, contact the FDA staff responsible for this guidance as listed on the title page.

I. INTRODUCTION

Allogeneic cells of human origin may be expanded in culture to manufacture medical products consisting of live cells, inactivated cells, cell lysates, or other cell-based materials such as cell-derived particles. We, FDA, are providing you, sponsors of allogeneic cell-based medical products, recommendations for determining the appropriate cell safety testing to support an Investigational New Drug Application (IND)¹ or a Biologics License Application (BLA).² Cell safety testing should be based on a risk analysis that considers the expansion potential of the cells, the reagents that are used to expand the cells in culture, and the number of individuals the cell-based medical product is capable of treating.^{3,4}

This guidance supplements the following two final guidances:

- “Chemistry, Manufacturing, and Control (CMC) Information for Human Gene Therapy Investigational New Drug Applications (INDs); Guidance for Industry,” dated January 2020⁵, and
- “Guidance for FDA Reviewers and Sponsors: Content and Review of Chemistry, Manufacturing, and Control (CMC) Information for Human Somatic Cell Therapy Investigational New Drug Applications (INDs),” dated April 2008.⁶

¹ See generally 21 CFR part 312.

² See generally 21 CFR part 601.

³ Depending on the stage of development, this may be the number of subjects to which the investigational product is capable of being administered in clinical investigations or the number of patients the product is capable of treating following licensure.

⁴ This guidance does not address the measurement or analysis of cell characteristics that may be relevant to biological activity.

⁵ <https://www.fda.gov/media/113760/download>.

⁶ <https://www.fda.gov/media/73624/download>.

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35 In general, FDA’s guidance documents, including this guidance, do not establish legally
36 enforceable responsibilities. Instead, guidances describe the FDA’s current thinking on a topic
37 and should be viewed only as recommendations, unless specific regulatory or statutory
38 requirements are cited. The use of the word *should* in FDA’s guidances means that something is
39 suggested or recommended, but not required.

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42 **II. SCOPE**

43

44 This guidance applies to allogeneic cell-based products that are regulated by the Office of
45 Therapeutic Products of the Center for Biologics Evaluation and Research (CBER) under section
46 351 of the Public Health Service Act (42 U.S.C. 262). This guidance applies to cultured
47 allogeneic cells, including cell banks⁷, that are sources of the intended constituents of the final
48 drug product, as well as combination products that contain an allogeneic cell or cell-based
49 biologic constituent part in combination with a drug and/or device. The recommendations in this
50 guidance also apply to genetically modified allogeneic cells that have been transduced with viral
51 and/or plasmid vectors, and cells that have undergone genome editing. This guidance does not
52 apply to cell substrates that are used during manufacturing of non-cell-based products such as
53 viruses, gene therapy vectors, or recombinant proteins.⁸ Recommendations for feeder or other
54 cells used as reagents during manufacturing are beyond the scope of this guidance.⁹

55
56

57 **III. BACKGROUND**

58

59 Viral and microbial contamination is a potential risk for all cell-based medical products,
60 especially when the cells are cultured extensively during manufacturing. Contamination may be
61 present in the source cells, or the cells may become contaminated with adventitious agents
62 during manufacturing. In addition, genomic changes that result in tumorigenic cells can occur
63 during extensive culture.

64

65 Under Title 21 of the Code of Federal Regulations (CFR), 610.18(c)(1), “Cell lines used for
66 manufacturing biological products shall be:

67

68 (i) Identified by history;

⁷ For the purpose of this guidance, cell bank refers to cells of uniform composition that are stored for further manufacturing.

⁸ The recommendations in this guidance do not apply to products reviewed by CBER’s Office of Vaccine Research and Review or Office of Blood Research and Review, or by the Center for Drug Evaluation and Research or the Center for Devices and Radiological Health.

⁹ Refer to the FDA draft guidance entitled “Considerations for the Use of Human- and Animal-Derived Materials and Components in the Manufacture of Cell and Gene Therapy and Tissue-Engineered Medical Products; Draft Guidance for Industry” dated April 2024 for recommendations on safety testing of feeder cells. <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/considerations-use-human-and-animal-derived-materials-manufacture-cell-and-gene-therapy-and-tissue>. When finalized, this guidance will represent FDA’s current thinking on the topic.

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- 69 (ii) Described with respect to cytogenetic characteristics and tumorigenicity;
- 70 (iii) Characterized with respect to in vitro growth characteristics and life potential; and
- 71 (iv) Tested for the presence of detectable microbial agents.”

72

73 In addition, 21 CFR 312.23(a)(7)(i) requires an IND to include sufficient information to assure
74 the proper identification, quality, purity, and strength of the investigational product. An IND is
75 also specifically required to include a description of the acceptable limits and analytical methods
76 used to assure, among other things, the purity and quality of both the drug substance (DS) and
77 drug product (DP), 21 CFR 312.23(a)(7)(iv)(a)-(b). Potential impurities for cell-based medical
78 products include contamination with viruses or bacteria. Genomic integrity and in vitro growth
79 characteristics are factors that affect the quality of cell-based medical products used under INDs
80 and thus should be evaluated. FDA may place an IND on clinical hold if the IND does not
81 contain sufficient CMC information required under 21 CFR 312.23 “to assess the risks to
82 subjects of the proposed studies” (21 CFR 312.42(b)(1)(iv) and (b)(2)(i)) or if the CMC
83 information indicates that the “[h]uman subjects are or would be exposed to an unreasonable and
84 significant risk of illness or injury” (21 CFR 312.42(b)(1)(i) and (b)(2)(i)).

85

86 The purpose of this document is to provide guidance on safety testing to assist manufacturers in
87 addressing the requirements of 21 CFR 610.18(c)(1), 21 CFR 312.23(a)(7), and other relevant
88 regulations, as applicable, with respect to human allogeneic cells expanded for use in cell-based
89 medical products. FDA’s recommendations for cell safety testing reflect a risk-based approach
90 that takes into consideration both the specific characteristics of the cells and their proposed use.

91

92 We recommend that you provide an integrated assessment of the risk of potential contamination
93 with adventitious agents in the electronic Common Technical Document (eCTD) section 3.2.A.2
94 (Adventitious Agents Safety Evaluation) of your IND or BLA submission (Ref. 1).¹⁰ We also
95 recommend that this section describe risk mitigation measures, including information on the
96 selection, testing, and safety assessment of cells and cell banks.

97

98

99

IV. CONSIDERATIONS FOR CELL SAFETY TESTING

100

101 For allogeneic cells, donor screening and testing must be performed as required in 21 CFR part
102 1271, subpart C, except for those cells that meet the exceptions in 21 CFR 1271.90(a).¹¹ The
103 donor screening and testing information should be provided in the IND or BLA submission.

104

105 In an IND, you must provide a list of all materials used in manufacturing, including the quality
106 or grade of these materials (see 21 CFR 312.23(a)(7)(iv)(b)), which assists FDA in determining
107 if the proposed cell safety testing is adequate to assure the safety of the investigational product.

¹⁰ For information on the submission of an eCTD, see the FDA website <https://www.fda.gov/drugs/electronic-regulatory-submission-and-review/electronic-common-technical-document-ectd>.

¹¹ For more information regarding these requirements, see Testing Donors of Human Cells, Tissues, and Cellular and Tissue-Based Products (HCT/P): Specific Requirements, <https://www.fda.gov/vaccines-blood-biologics/safety-availability-biologics/testing-donors-human-cells-tissues-and-cellular-and-tissue-based-products-hctp-specific-requirements>.

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108 Information describing the source of each reagent and its components, along with a Certificate of
109 Analysis, should be submitted within IND. The origin of animal- and human-derived reagents
110 used in the manufacturing process, and the acceptance criteria for these reagents, will influence
111 the appropriate amount and types of safety testing.¹²

112
113 The nature and extent of cell safety testing needed to provide adequate assurance of product
114 safety will generally depend on the expansion potential of the cells and the number of individuals
115 the cell-based medical product is capable of treating. A description of the culture methods used
116 to expand the cells to manufacture the allogeneic cell-based medical product should be provided
117 in the IND or BLA submission. This information should include the expansion potential of the
118 cells, the passage number that is intended to be used to make the allogeneic cell-based medical
119 product and cell storage conditions.

120

121 **A. Continuous Cell Lines**

122

123 Cellular products may be produced from continuous cell lines, including induced
124 pluripotent stem cells, embryonic stem cells, cancer cell lines, and transformed cell lines.
125 The manufacturing process for a cellular product made from continuous cell lines usually
126 includes the creation of a cell bank to ensure a consistent source material for the
127 manufacture of the cell-based product.

128

129 Cell banks made from continuous cell lines should be tested for adventitious viruses and
130 undergo additional safety evaluations as outlined in section V of this guidance.

131

132 **B. Primary Cells**

133

134 1. Primary Cells Capable of Extensive Expansion in Culture

135

136 If allogeneic primary cells from a single donor can be expanded in culture to a cell
137 number that is sufficient to be administered to many individuals, then we recommend
138 conducting the cell safety testing described in section V of this guidance. This testing
139 should generally be performed on a cryopreserved cell bank. However, some
140 manufacturers of cell-based medical products do not use a cell bank during
141 manufacturing. Instead, they expand the cells extensively and then store them as a
142 cryopreserved lot of drug substance or final product. In this case, the cell safety
143 testing outlined in section V of this guidance should be done on the lot of
144 cryopreserved cells.

145

146 2. Primary Cells Capable of Limited Expansion in Culture

147

¹² Refer to the FDA draft guidance entitled “Considerations for the Use of Human- and Animal-Derived Materials and Components in the Manufacture of Cell and Gene Therapy and Tissue-Engineered Medical Products; Draft Guidance for Industry” dated April 2024 for additional information regarding human- and animal-derived reagents. When finalized, this guidance will represent FDA’s current thinking on the topic.

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148 Primary cells with a limited expansion potential can be expanded to make a cellular
149 therapy product, or to create small to midsize cell banks or a single lot of cells to
150 manufacture cell-based products capable of being administered to a limited number of
151 individuals. In these situations, the complete list of cell safety testing outlined in
152 section V of this guidance is not recommended because full adventitious virus testing
153 is generally not feasible to perform on a small lot of cells or final product. Instead, an
154 abbreviated test matrix, as outlined in section VI of this guidance, is recommended.

155 **C. Cells That Are Administered To A Few Individuals Or A Single Individual**

156 Primary cell cultures that are not subcultivated or primary cell cultures that are
157 subsequently subcultivated for only a very limited number of population doublings are
158 not subject to the provisions in 21 CFR 610.18(c) for cell lines used for manufacturing
159 biological products (21 CFR 610.18(c)(3)). It is not recommended that primary
160 allogeneic cells that are minimally expanded in culture to be administered to only a few
161 individuals, or a single individual, undergo cytogenetic analysis or adventitious virus
162 testing. However, these cells must still be tested for sterility (21 CFR 610.12), purity
163 (e.g., endotoxin) (21 CFR 610.13), as required for lot release of licensed cellular products
164 (21 CFR 610.1). Note that if there are specific safety concerns regarding reagents used
165 during product manufacturing, then adventitious virus testing may need to be performed
166 on the reagents of concern to assure product safety. For example, there may be a safety
167 concern due to the use of animal and human derived reagents because they have the
168 potential to introduce adventitious agents to the cell-based medical product.
169
170
171
172

173 **V. TESTING RECOMMENDATIONS FOR HIGHLY EXPANDED CELLS**

174 This section contains recommendations for testing cell banks of highly expanded primary cells,
175 and cell banks made from continuous cell lines, including pluripotent stem cells, cancer cells,
176 and transformed cells.
177

178 Cell-based medical products may use a one-tier or two-tier cell banking system. A one-tier
179 system consists of a master cell bank (MCB) only, while in a two-tier system there is also at least
180 one working cell bank (WCB) derived from the MCB.
181

182 Some manufacturing schemes may use multiple levels of cell banks; however, it may not be
183 necessary to test all the cell banks for safety as outlined below. For instance, cells used as the
184 cellular starting material for genetically engineered stem cells may be banked but would not be
185 considered the MCB. Instead, the cell bank of genetically modified stem cells would generally
186 be considered the MCB, and the cell safety testing described below should be performed on
187 those genetically modified cells since there is potential for adventitious agent contamination
188 during the genetic modification. Likewise, if donor cells used as starting material for a cell-
189 based product are banked prior to extensive expansion in culture, the highly expanded cells
190 should be considered the MCB and should be used for the safety testing outlined below.
191
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193 Some cell-based medical products do not use a cell bank during manufacturing but may store the
194 final product as a single lot of highly expanded cryopreserved cells that could be used to treat
195 many individuals. In this case, the MCB cell safety testing outlined below should be performed
196 on the lot of cryopreserved cells.

197 198 **A. Master Cell Bank**

199
200 FDA makes the recommendations below regarding safety testing for an MCB. (We note
201 that, for licensure, testing must comply with the general biological products standards in
202 21 CFR Part 610).

- 204 • Sterility testing – Testing for bacterial and fungal sterility may be performed
205 in accordance with United States Pharmacopeia (USP)<71> or using another
206 appropriately qualified and validated method (see 21 CFR 610.12).
207
- 208 • Mycoplasma testing – Mycoplasma testing can consist of culture-based
209 methods per USP<63>. Alternative assays may be used to detect
210 mycoplasma, but such an assay should be shown to have sensitivity that is
211 comparable to the compendial method.
212
- 213 • Human pathogen testing using polymerase chain reaction (PCR) may include
214 testing for human immunodeficiency virus (HIV) -1 &2, human T cell
215 lymphotropic virus (HTLV) 1 & -2, hepatitis viruses B and C,
216 cytomegalovirus (CMV), Epstein-Barr virus (EBV), human parvovirus B19,
217 human papillomavirus (HPV), human herpes viruses (HHV) -6, -7, and 8,
218 John Cunningham (JC) virus, and BK virus, as appropriate. FDA should be
219 consulted for application-specific testing recommendations when cells will
220 be used in immunocompromised individuals.
221
- 222 • In vitro adventitious virus testing – Three cell lines should generally be used:
223 human diploid (e.g., MRC5 cells), monkey kidney (e.g., Vero cells), and
224 another cell line of the same species and tissue type as that used for
225 production (e.g., HeLa cells if the product was made using human cells).
226 However, different cell lines may be appropriate depending on the
227 manufacturing process. For instance, when insect cells are used during
228 manufacturing, BHK21 cells may be used to detect viruses such as
229 rhabdoviruses. In this example, testing for adventitious viruses using BHK21
230 cells would address the recommendation of testing for viruses in cells of the
231 same species in which product production occurs. The BHK21 cells would
232 be the third cell line recommended for adventitious virus testing when used
233 in addition to the human diploid and monkey kidney cell lines.
234
- 235 • In vivo adventitious virus testing is recommended when cells have specific
236 risk factors that are not fully mitigated by other types of testing. Examples of
237 such risk factors include, contact with animals or animal cells, use of animal

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238 or human derived reagents that do not have sufficient safety information, use
239 of cells from unknown sources, and insufficient information on cell culture
240 history and methods. In vivo adventitious virus testing consists of
241 inoculation of adult and suckling mice, and embryonated chicken eggs with
242 cells. Alternatively, a high throughput sequencing method may be used
243 instead of in vivo adventitious virus testing to detect contaminating viruses.
244 However, if a high throughput sequencing method such as next generation
245 sequencing is used, we recommend that you include a description of any
246 qualification or validation studies performed in your regulatory submission.
247 We strongly recommend that if a high throughput sequencing method is used
248 for adventitious virus testing, the proposed method and validation study plan
249 be discussed with FDA prior to implementation.

- 251 • Transmission electron microscopy should be performed to detect virus
252 particles.
- 253
- 254 • Retroviral testing – If the human sourced cells used to make the cell-based
255 medical product are grown on feeder layers of non-human cells, these human
256 cells should be evaluated for the presence of species-specific and endogenous
257 retrovirus.
- 258
- 259 • Species-specific virus testing should be performed. If human sourced cells
260 contact rodent cells or rodent-derived reagents during manufacturing, then
261 testing for mouse/rat/hamster viruses should be performed. Likewise, if the
262 human sourced cells contact simian or insect cells or reagents, then testing
263 for simian or insect viruses should be performed.
- 264
- 265 • Testing for animal viruses consistent with the testing described in 9 CFR
266 113.53 (Requirements for ingredients of animal origin used for production of
267 biologics) and 9 CFR 113.47 (Detection of extraneous viruses by the
268 fluorescent antibody technique) should be performed if animal-derived
269 reagents are used during manufacturing of the cell-based medical product.
270
 - 271 ○ Testing consistent with 9 CFR 113.47 should be performed for
272 bovine derived viruses listed in 113.47(b)(1) and (2), if bovine-
273 derived reagents are used. Note that bovine-derived reagents
274 should be obtained from sources that minimize the risk of
275 transmissible spongiform encephalopathy.
 - 276
 - 277 ○ Testing for porcine-derived viruses consistent with the testing
278 described in 9 CFR 113.53(d) and 9 CFR 113.47(b)(1) and (6)
279 should be performed if porcine-derived reagents are used.
 - 280
 - 281 ○ It may be acceptable to reduce or eliminate testing of the human-
282 sourced cells for animal viruses if the reagent manufacturer

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283 performs and documents adventitious agent testing for the animal-
284 derived reagents consistent with 9 CFR 113.53 and 9 CFR 113.47.
285 Reagent testing documentation should be submitted in the IND or
286 BLA submission.

- 287
- 288 • Testing for the presence of residual viral and plasmid reprogramming vectors
289 used in the creation of induced pluripotent cell lines should be performed on
290 either the cell bank, drug substance, or final product. An acceptance criterion
291 with justification for acceptable levels of residual programming vectors
292 should be established.
293
 - 294 • If a retroviral vector is used to induce gene editing, then the cells used to
295 make the allogeneic cell-based medical product should be tested for the
296 presence of replication competent retrovirus as recommended in the FDA
297 guidance titled “Testing of Retroviral Vector-Based Human Gene Therapy
298 Products for Replication Competent Retrovirus During Product Manufacture
299 and Patient Follow-up; Guidance for Industry” dated January 2020 (Ref. 2).
300
 - 301 • Whole genome sequencing and analysis should be performed on cell banks
302 of continuous cell lines and genome edited cells.
303
 - 304 ○ Cell lines that are cultured extensively often accumulate mutations
305 during cell expansion. Mutations in protooncogenes, such as p53,
306 are of particular concern. Therefore, we recommend that
307 continuous cell lines that contribute cells to the final product be
308 evaluated by performing whole genome sequencing. The whole
309 genome sequencing method used should have a read depth of at
310 least 50X, and at a minimum, the results should be compared to a
311 database of cancer associated mutations. Justification should be
312 provided for the sequencing method, read depth, and for
313 conclusions related to the safety of the product.
314
 - 315 ○ For highly expanded clones of genetically modified cells, whole
316 genome sequencing with at least 50X read depth should be
317 performed to identify off-target genome editing, on-target editing
318 outcomes, vector integration events, and to screen for any
319 mutations of concern.
320
 - 321 • Cytogenetic testing or whole genome sequencing should be performed on
322 highly expanded primary cells that contribute cells to the final product.
323 Whole genome sequencing as described above is the recommended method
324 of testing genome integrity. Alternatively, if cytogenetic testing is
325 performed, G-banding analysis or other sensitive methods should be used to
326 confirm the cells have a normal karyotype. The karyotypes of at least 20
327 cells should be analyzed. An acceptance criterion for cytogenetic test results

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328 with justification for discrepancies should be established. Cytogenetic
329 testing is not recommended for continuous cell lines or highly expanded
330 genetically modified cells that have been subjected to whole genome
331 sequencing as recommended above.
332

- 333 • Cytogenetic testing is not recommended for cells that are only used as
334 sources for products such as cell-derived particles and are not present in the
335 final product. Cytogenetic testing is not recommended for irradiated cells
336 used during manufacturing or that are a part of the final product.
337
- 338 • Tumorigenicity testing, highly expanded cells – Under 21 CFR
339 610.18(c)(1)(ii), cell lines used for manufacturing biological products shall
340 be described with respect to tumorigenicity.
341
 - 342 ○ In cases where the cells present in the final product are
343 phenotypically similar to those in the MCB, the tumorigenic
344 potential of a product may be tested using cells from the MCB.
345 However, tumorigenicity testing may not be necessary if the cells
346 in the MCB are demonstrated to be comparable to cells that were
347 evaluated and tested for tumorigenicity in preclinical studies.
348 Comparability can be evaluated by measuring product specific
349 characteristics that are associated with product performance and
350 safety.
351
 - 352 ○ Tumorigenicity testing, continuous cell lines – Genomic stability
353 and growth characteristics should be evaluated, but cancer cell
354 lines and pluripotent cell lines are generally not tested for
355 tumorigenicity as part of cell safety testing since these cell types
356 are expected to be capable of forming tumors.
357
 - 358 ○ Tumorigenicity testing, irradiated cells – Tumorigenicity testing of
359 irradiated cells is not recommended.
360

361 For detailed information on adventitious virus testing, please refer to section IV.A of the
362 guidance “Guidance for Industry: Characterization and Qualification of Cell Substrates
363 and Other Biological Materials Used in the Production of Viral Vaccines for Infectious
364 Disease Indications,” dated February 2010 (Ref. 3). Sponsors who intend to use novel
365 analytical technology for adventitious agent evaluation of their product should discuss
366 their proposed approach with FDA during the development of the assay.
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370 **B. Working Cell Bank**

371
372 In some cases, the cells from an MCB may be expanded further into WCBs. WCB
373 testing should include, but not be limited to, sterility, mycoplasma, identity, and in vitro
374 adventitious agent tests described in section V.A of this guidance.

375 376 377 **VI. TESTING RECOMMENDATIONS FOR CELLS WITH LIMITED EXPANSION** 378 **POTENTIAL**

379
380 As discussed in section IV.B.2 of this guidance, it is generally unnecessary for primary cells that
381 cannot be expanded in culture extensively to undergo all the testing listed in section V of this
382 guidance, unless there are specific safety concerns that such testing would address.

383
384 Cell banks or product lots made from cells with limited expansion capability should be tested
385 using an abbreviated cell safety test matrix consisting of:

- 386
387
 - Sterility testing;
 - Mycoplasma testing;
 - Human pathogen testing using PCR, as described in section V of this guidance; and
 - In vitro adventitious virus testing as described in section V of this guidance.

391
392 Additional safety testing for cells that come in contact with animal-derived reagents during
393 manufacturing, as described in section V of this guidance, may be appropriate.

394
395 Genome edited cells that are not extensively expanded in culture should undergo targeted
396 sequencing to assess the frequency of editing at confirmed off-target sites and to ensure the
397 desired on-target editing outcome has occurred. If a retroviral vector is used to induce gene
398 editing, then the cells used to make the allogeneic cell-based medical product should be tested
399 for the presence of replication competent retrovirus (Ref. 2).

400
401 If there is a concern that even a limited amount of adventitious virus testing will consume too
402 large a portion of the cellular material available, then testing may be performed on end of
403 production material. The end of production material may be generated by expanding cells from a
404 product lot or a cell bank to a number of cells that is sufficient for the abbreviated cell safety test
405 matrix outlined above in this section.

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Table 1. Cell Safety Testing Recommendations for Allogeneic Cells Expanded for Use in Cell-Based Medical Products

Cell description	Cell culture and preparation	Cells that should be tested	Product Use	Cell safety testing recommended
Embryonic stem cells and allogeneic induced pluripotent cells	Cells are expanded into an MCB and WCBs. WCBs are differentiated into final cellular therapy product.	MCB and WCBs	Potentially, many individuals	The MCB and WCBs should be tested as outlined in section V of this guidance
Immortal cancer cell lines and transformed cell lines	Cells are expanded into an MCB and WCBs. Cell-based product is derived from WCBs.	MCB and WCBs	Potentially, many individuals	The MCB and WCBs should be tested as outlined in section V of this guidance
Primary allogeneic cells capable of extensive expansion (highly expanded)	Cells are expanded to make an MCB. MCB vials are thawed and further expanded to make final product.	MCB and WCBs (if one is made)	Potentially, many individuals	The MCB and WCBs (if one is made) should be tested as outlined in section V of this guidance
Primary allogeneic cells, including some genetically engineered cells, capable of limited expansion before loss of cell quality	Cells are expanded several passages to make a small to midsized MCB or a single lot of cells that is used as the cellular therapy product.	MCB or lot of expanded cells, or end of production cells	Limited number of individuals	MCB or lot of expanded cells, or end of production cells should be tested as outlined in section VI of this guidance
Primary allogeneic cells expanded in culture to make a product for a few subjects or a single subject	Cells are expanded to make product lots of cells capable of being administered to a few subjects or a single subject.	The lot of expanded cells	A few individuals or a single individual	Sterility, mycoplasma, and endotoxin testing

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412 **VIII. REFERENCES**

413

414 1. Guidance for Industry: M4Q: The CTD – Quality, August 2001.

415 <https://www.fda.gov/media/71581/download>.

416

417 2. Testing of Retroviral Vector-Based Human Gene Therapy Products for Replication
418 Competent Retrovirus During Product Manufacture and Patient Follow-up; Guidance for

419 Industry, January 2020. <https://www.fda.gov/media/113790/download>.

420

421 3. Guidance for Industry: Characterization and Qualification of Cell Substrates and Other
422 Biological Starting Materials Used in the Production of Viral Vaccines for Infectious Disease

423 Indications, February 2010. <https://www.fda.gov/media/78428/download>.

424

425